





# GPCRs in Human Biology & Pharmacology



800 GPCR are receptors for:

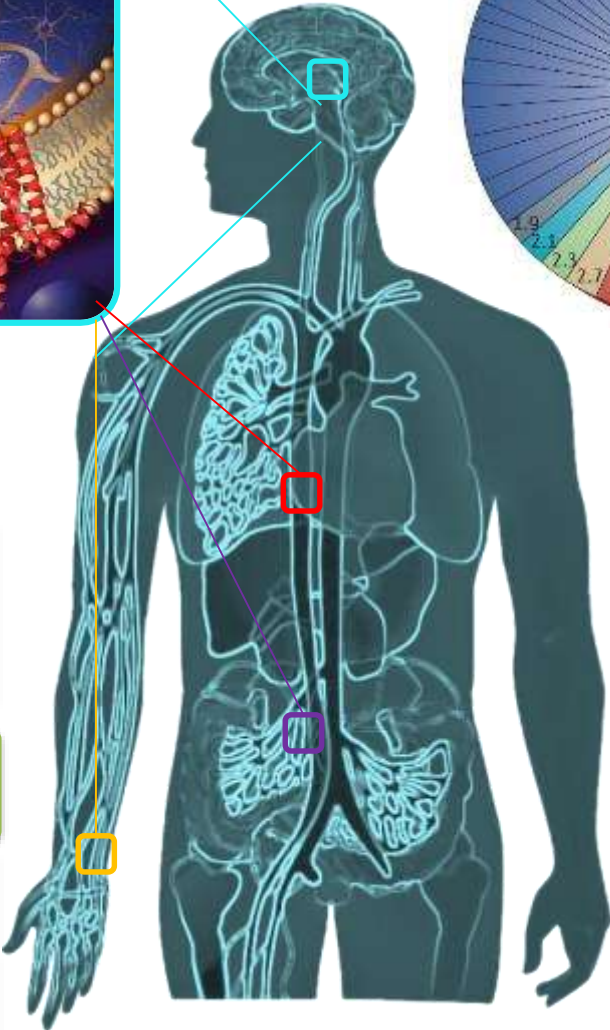
**Neurotransmitters**  
(adrenaline, dopamine, histamine, acetylcholine, adenosine, serotonin, glutamate, anandamide, GABA)

**Hormones & Neuropeptides**  
(opioid, neurotensin, glucagon, CRF, galanin, orexin, oxytocin, neuromedin, melanocortins, somatostatin, ghrelin, TRH, TSH, GnRH, , PTH, THS, LH... >30 total )

**Immune system**  
(chemokine, sphingosine 1 phosphate)

**Development**  
(Frizzled, Adhesion)

**Sensory**  
Light, Taste, Olfactory (388)



Nat Rev Drug Disc 5:993 (2006)

>30% of all drugs

GPCR Targets:  
>120 established  
> 800 potential

Disorders Targeted Clinically

Psychiatric, Learning & Memory, Mood, Sleep, Drug Addiction, Stress, Anxiety, Pain, Social Behavior...

Cardiovascular, Endocrine, Obesity, Immune, HIV, Reproductive ...

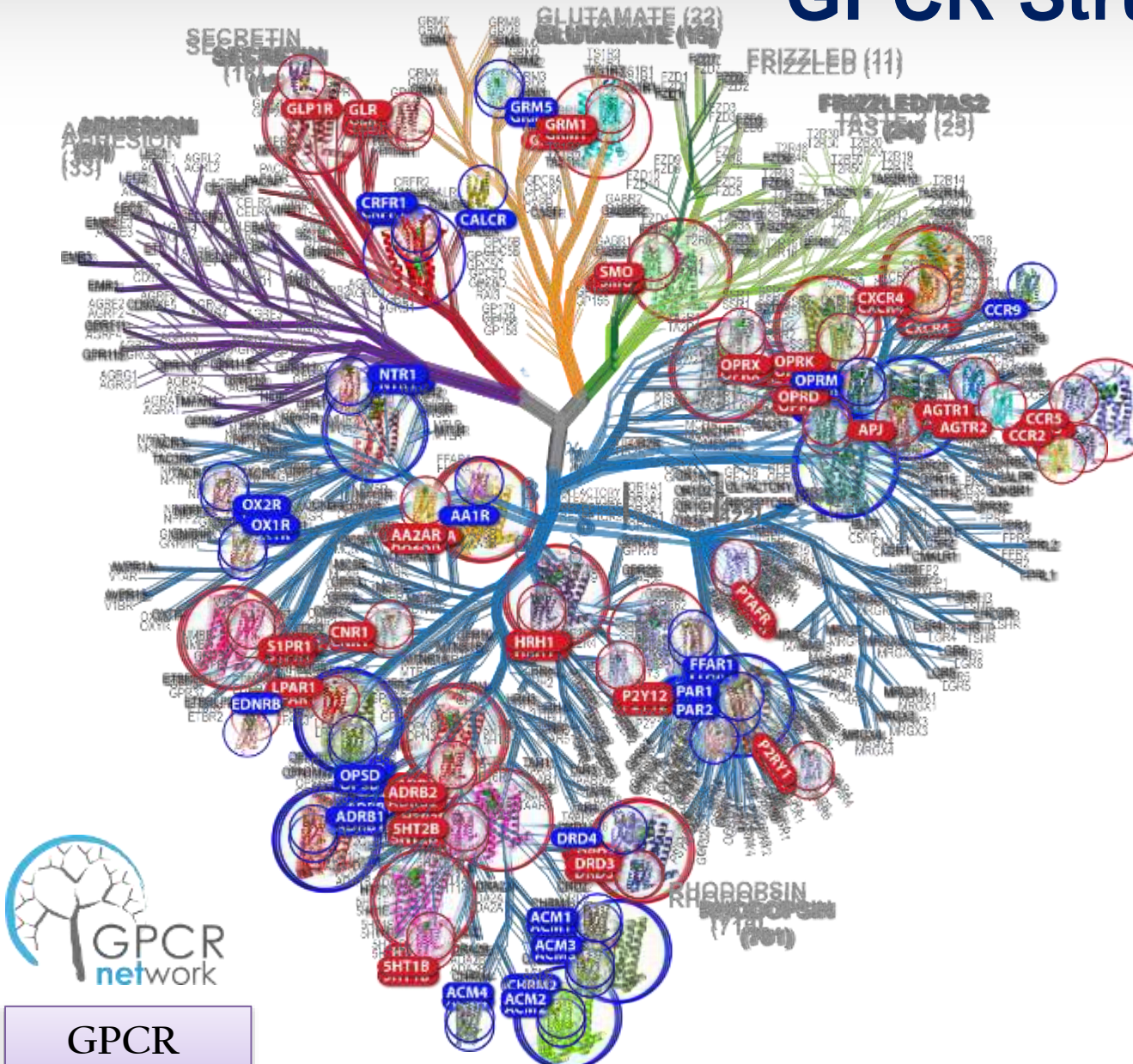
Neurodegenerative and Autoimmune Disease ...

Brain Development, Regenerative Med.

Cancer



# GPCR Structure and Function



Thousands of Ligands - Chemical Diversity

>800 Different Human Receptors  
(largest family in human genome)  
Share 7TM Fold But Diverse Structural Features

Dozens of Effectors



We use structure and modeling to learn:

- Molecular Recognition
- Signaling Mechanisms

and to design: ↓  
New tool compounds  
New receptor properties



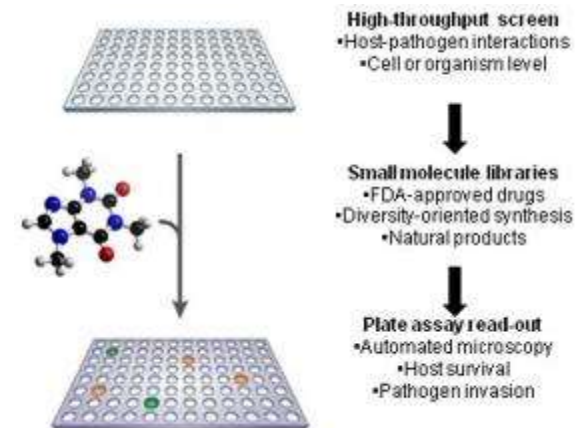
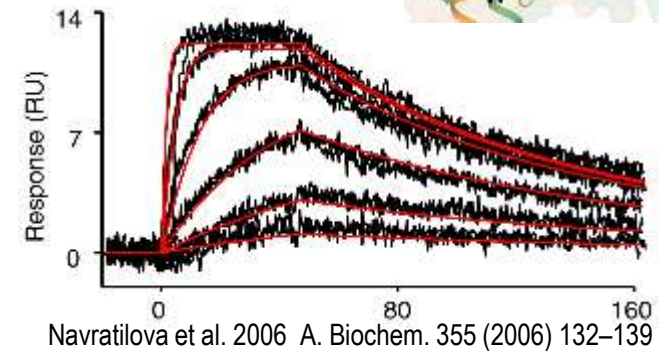
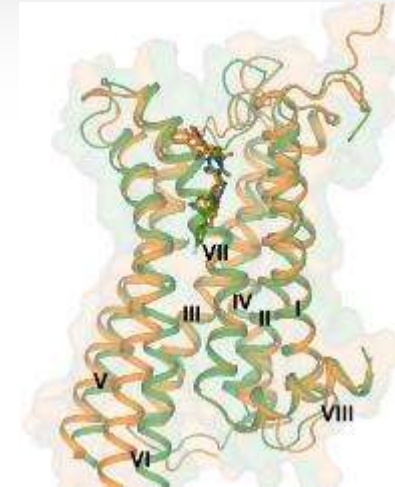
Katritch et al 2013 *Annu Rev Pharm. Tox.* **53**, 531-556  
Stevens et al. 2013, *Nat Rev Drug Discov*, **12**: 25-34

# Outline

- **Rational prediction of stabilizing mutations:  
CompoMug**
- **New insights into GPCR function and allosteric mechanisms**
- **Structure-Based ligand discovery for GPCRs**

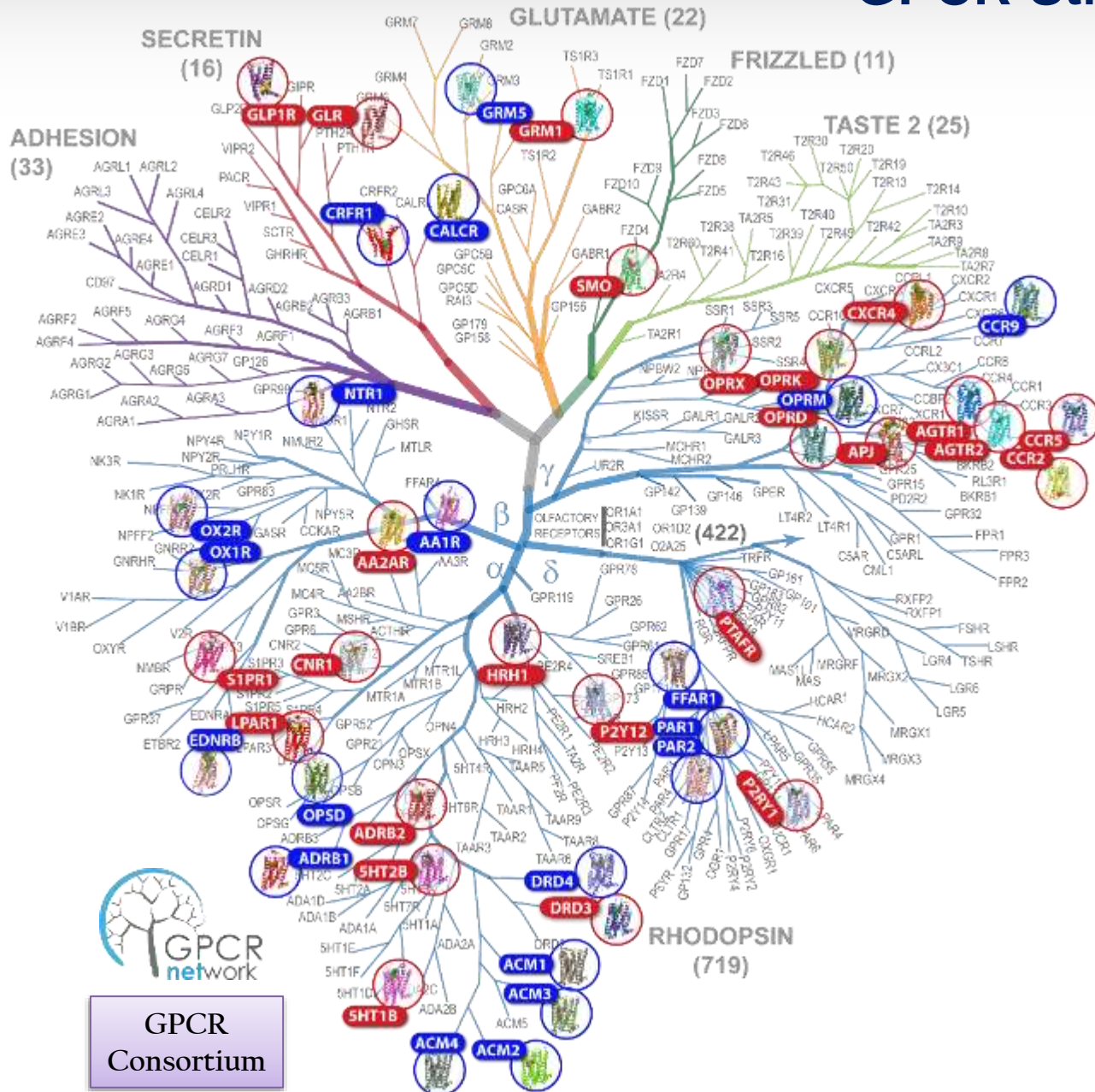
# Why Stabilizing GPCRs?

- **Crystallography:**
  - Synergistic with fusions and truncations
  - Reduces heterogeneity
  - Allows stabilization of active or inactive states
  - Allows co-crystallization with low-affinity ligands
- **Biophysical characterization**
  - SPR
  - NMR
- **Drug discovery**
  - More robust assays for HTS
  - NMR-based ligand screening
  - Ligand soaking for large-scale SBDD





# GPCR Structure and Function



- > 250 structures
- > 55 unique GPCRs

All required some protein engineering:

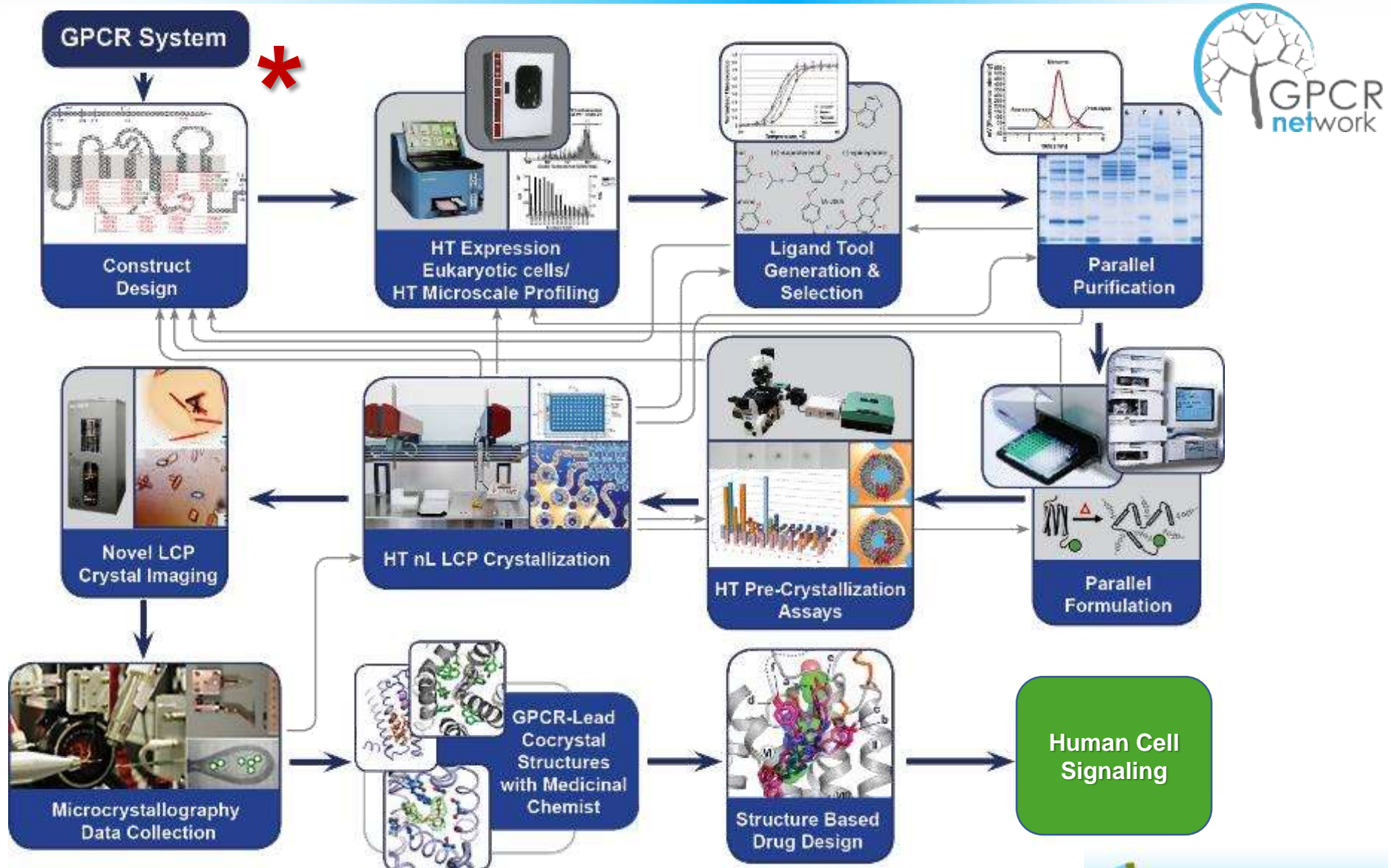
- 47 with fusion proteins
- 36 with mutations
- 25 with both
- All have truncations



# GPCR Network Structure-Function Pipeline



Multistage iterative process

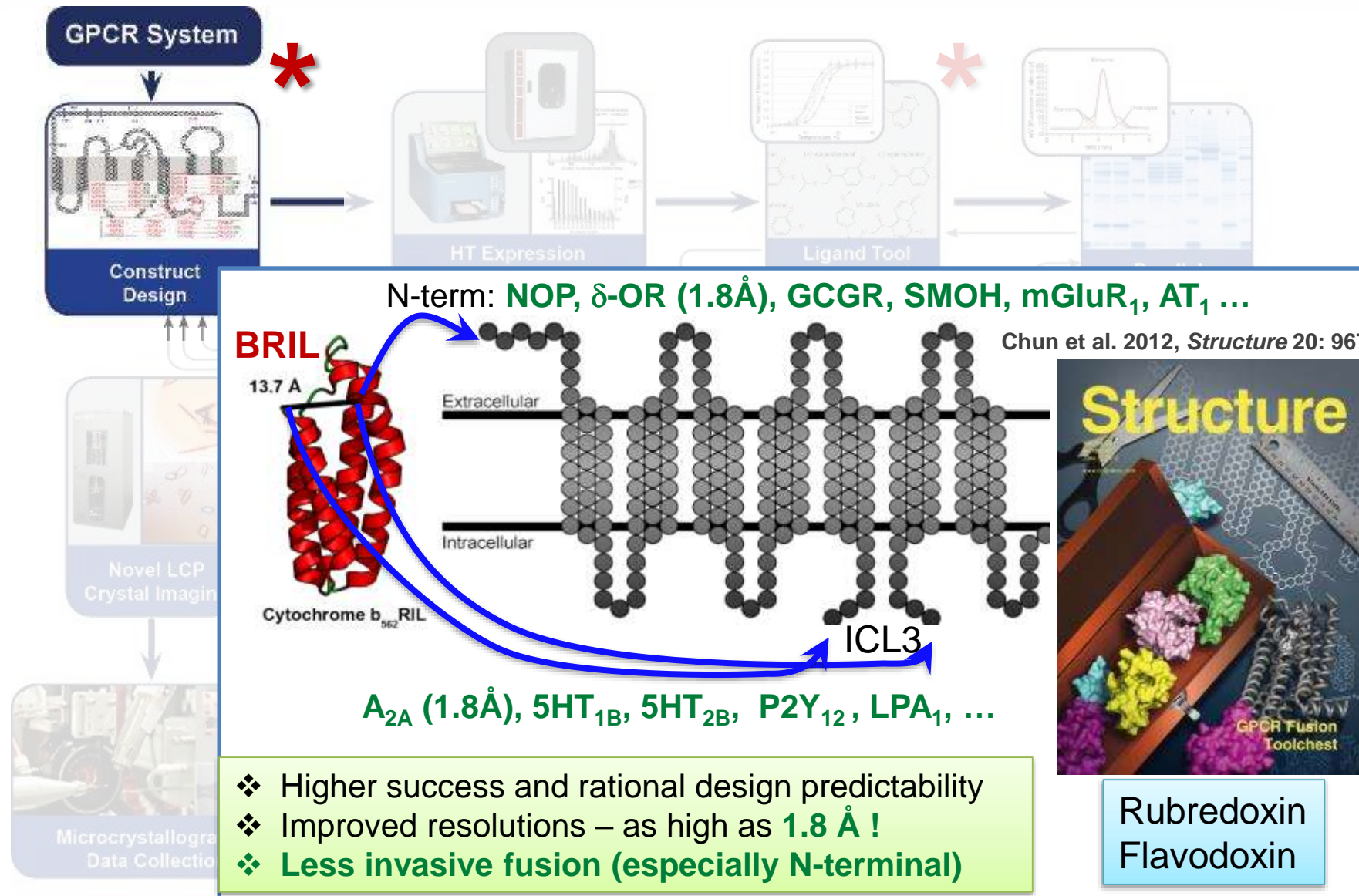




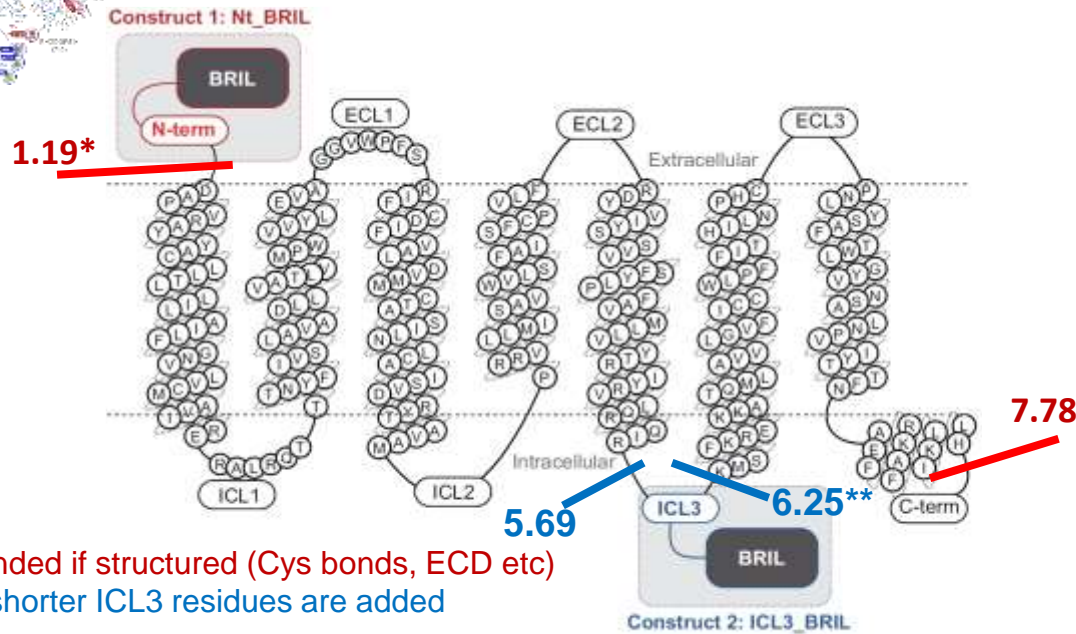
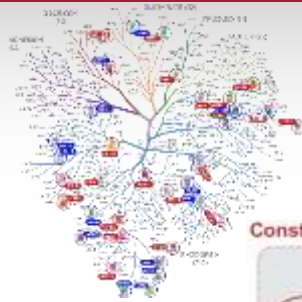
# GPCR Network Structure-Function Pipeline



## New Rationally Designed Protein Fusions

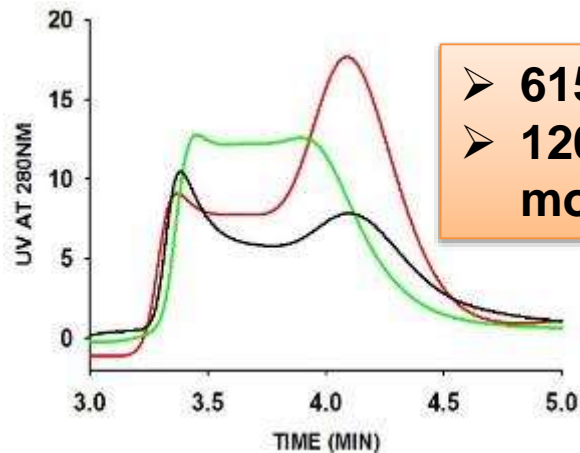


# Initial Constructs: GPCR-826 project



\* Extended if structured (Cys bonds, ECD etc)

\*\*For shorter ICL3 residues are added

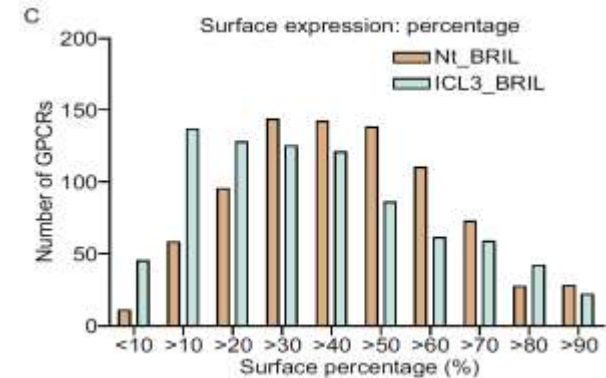
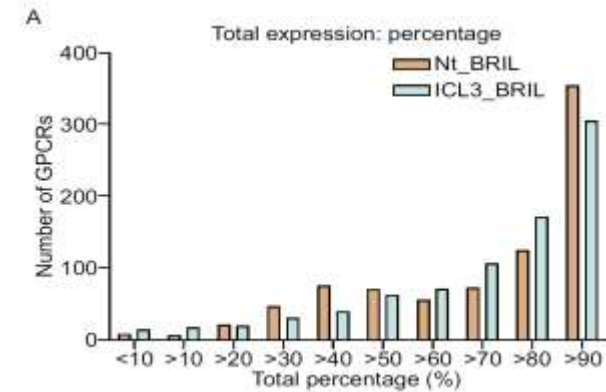


➤ 615 GPCRs purified

➤ 120 have strong monomer fractions

Surface Expression >30%:

- 80% (718) of Nt\_BRIL
- 60% (480) of ICL3\_BRIL



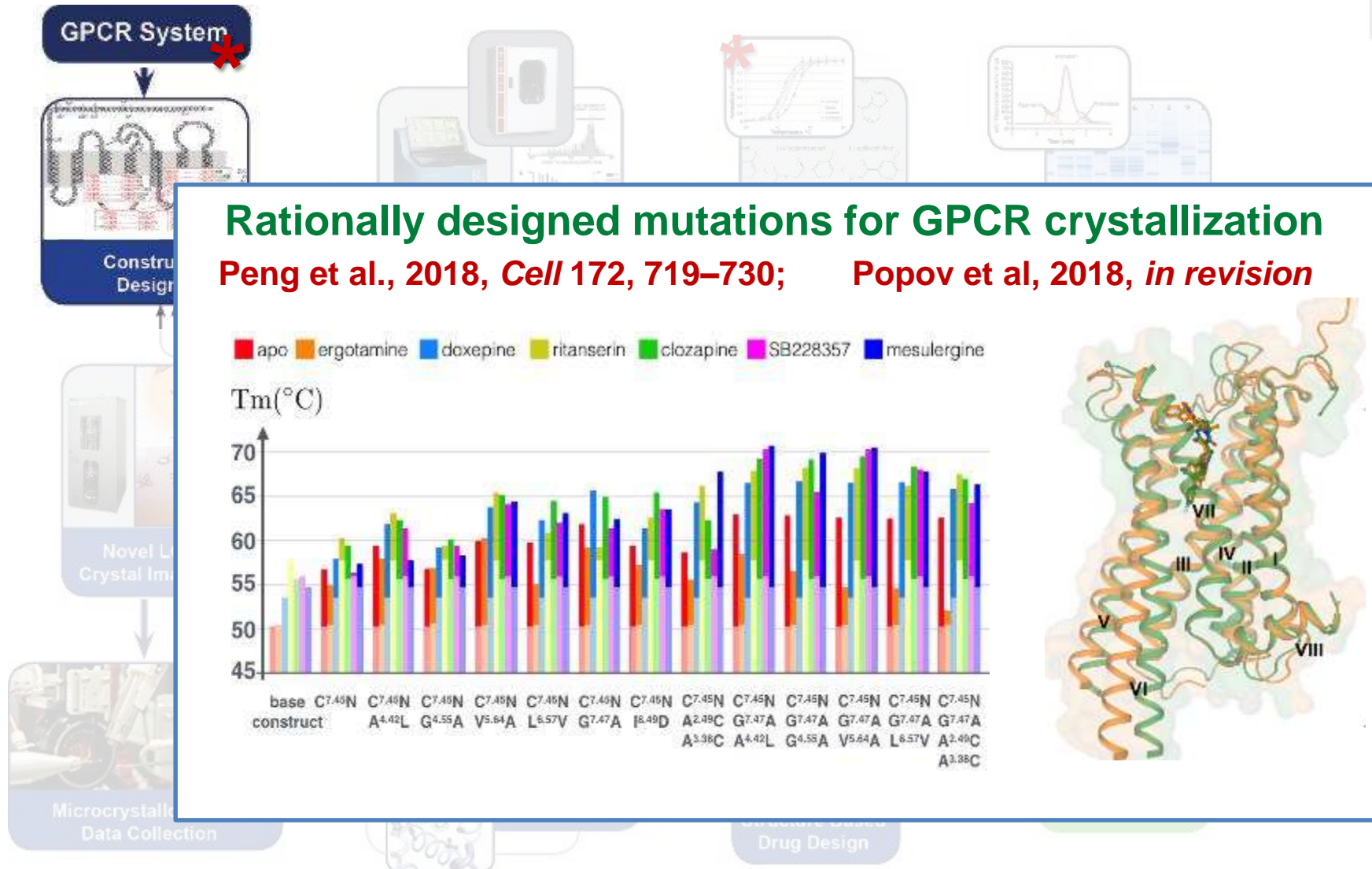
Xuechen Lv et al, Protein & cell (2016)

# GPCR Network Structure-Function Process

CompoMug: Rationally Designed Mutations



GPCR Consortium





# Stabilizing mutations: Experimental approaches

## Alanine/Leucine scanning

Number of mutants: 300-2000

- $\beta$ 1AR (Serrano-Vega et al., 2008, Warne et al., 2009)
- Adenosine A2A receptor (Magnani et al., 2008, Lebon et al., 2011, Robertson et al., 2011, Dore et al., 2011)
- Neurotensin NTSR1 (Shibata et al., 2009, 2013)
- mGLUR<sub>5</sub> (Dore et al., 2014)
- CRFR1 (Hollenstein et al., 2013)
- FFAR1 (Hirozane et al., 2014)

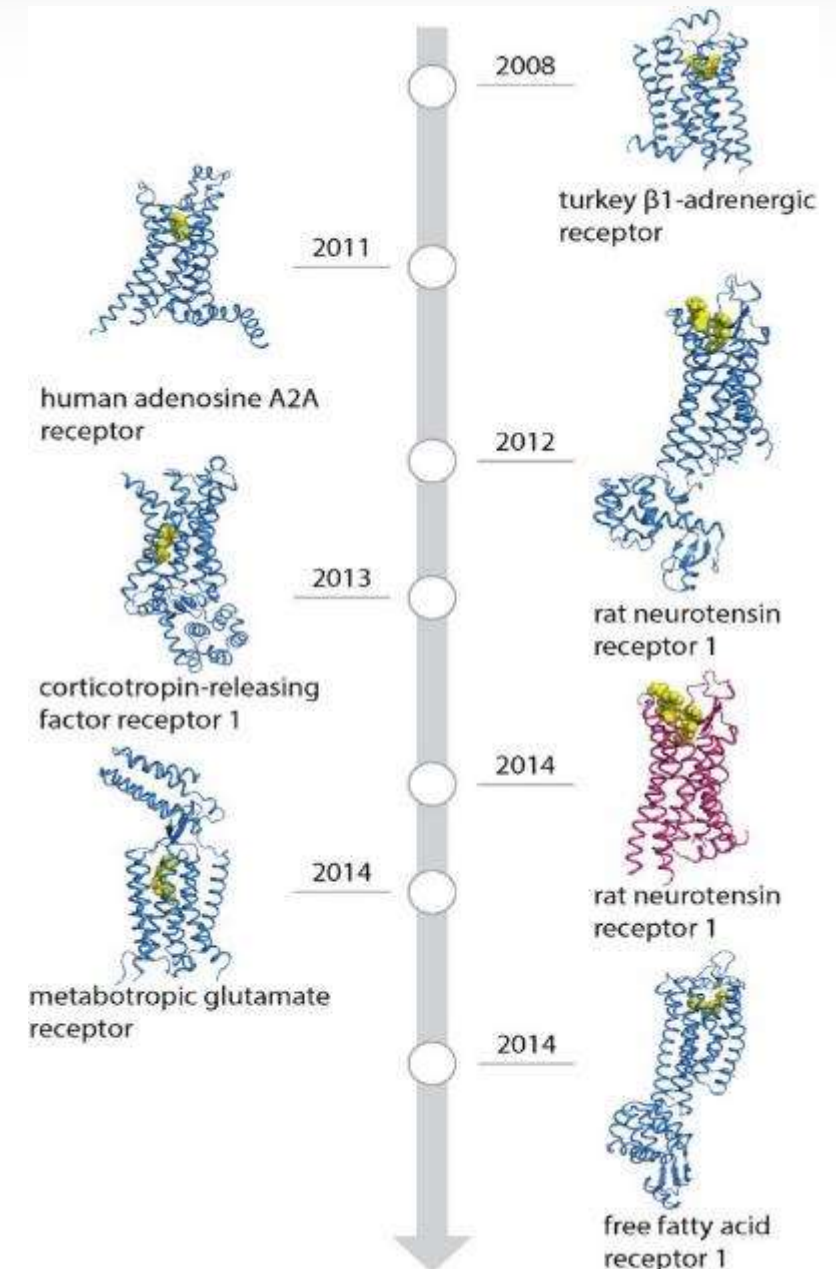
## Directed evolution

Number of mutants: > 1,000,000

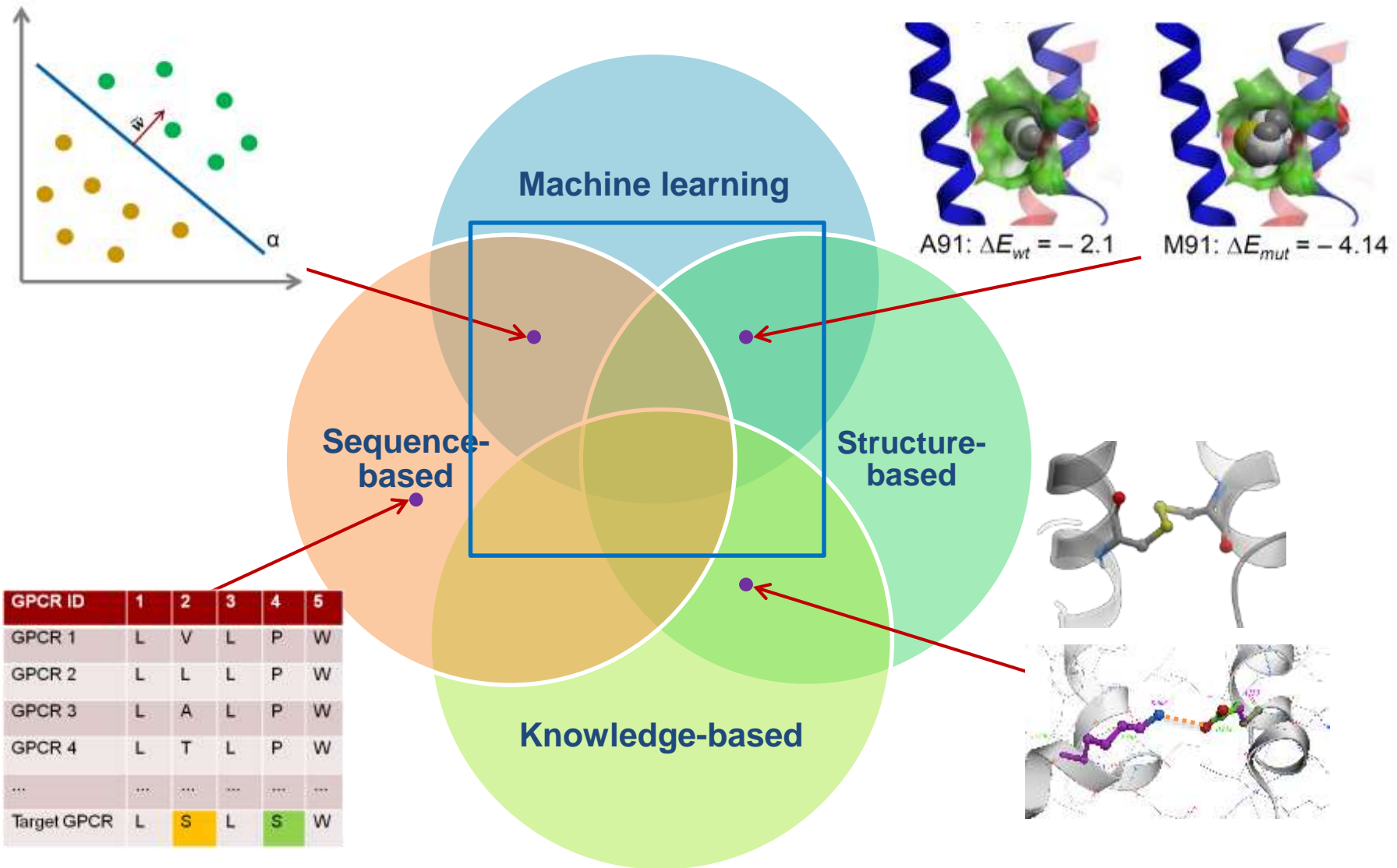
- NTSR1 (Sarkar et al., 2008, Dodevsky et al., 2011)
- $\alpha$ 1AR,  $\alpha$ 1BR (Dodevsky, Plückthun, 2011)
- Tachykinin receptor NK1 (Dodevsky, Plückthun, 2011)

**One might hope that in the future it might be possible to design thermostabilizing mutations, computationally predict them or transfer them from other receptors ...**

Heydenreich et al., 2015, Frontiers in Pharmacology



# ComPOMuG: Computational Predictions of Optimizing Mutations in GPCRs



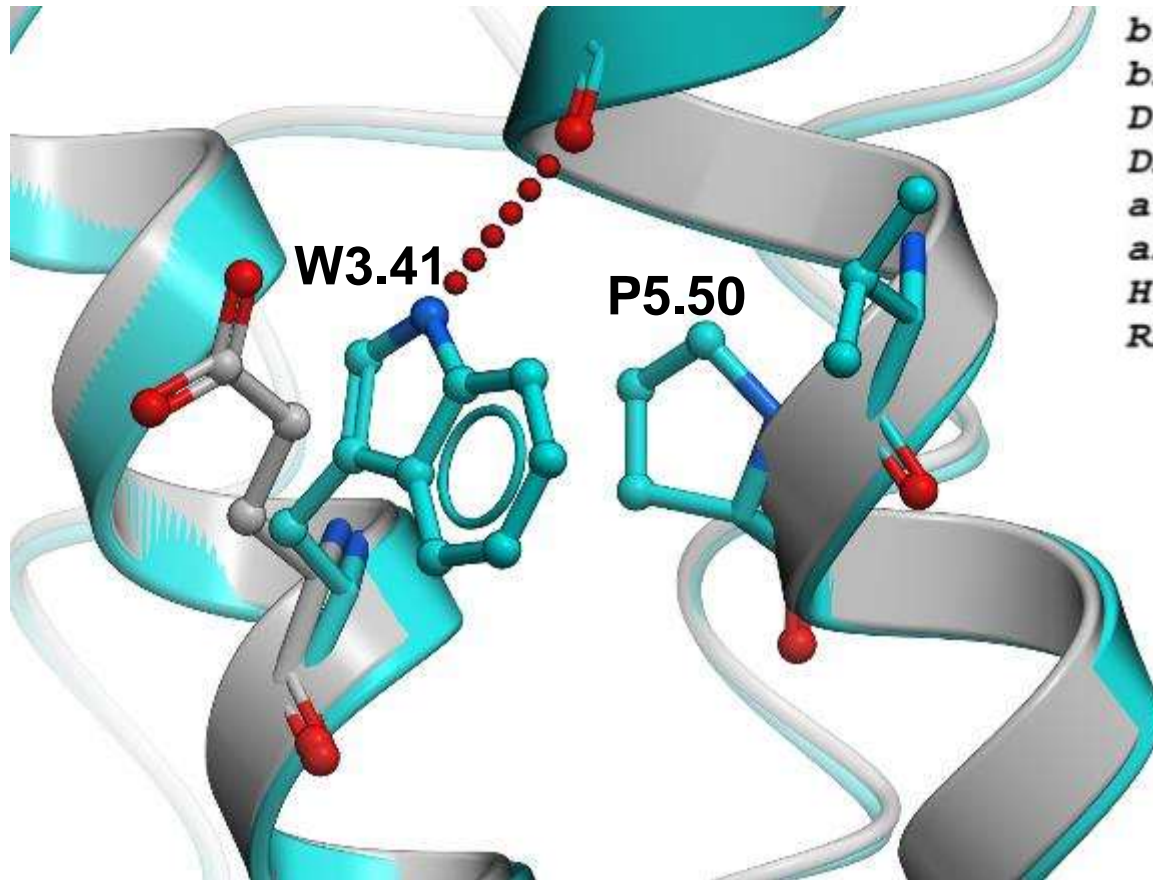
# Knowledge Based (Class A only)

- 2 or more structures with this mutant
- Known transferable position
- Most of them destroy/modify a conserved **functional** site

Position	Mutation	Role	Receptor (PDB ID)
3.41	X->W	stabilization of TM3 - TM5 interface	5HT2B (4IB4), 5HT1B (4IAR), ADRB1 (5A8E), ADRB2 (3NY8), CXCR4 (3ODU), DRD3 (3PBL)
2.50	D->N	Sodium pocket	AA2AR (5WF5)
3.39	S->A	Sodium pocket	AA2AR (5WF5)
7.49	D->N	Sodium pocket	P2RY1 (4XNV), P2Y12 (4PXZ)
3.40	I->V, A	P-I-F microswitch motif	ADRB1 (4BVN), APJ (5VBL)
3.49	D,G->A	DRY motif	FFAR1 (5TZR), NTR1 (4XES)
5.58	Y-> A	Conserved activation microswitch	FFAR1 (5TZR), ADRB1 (4BVN)
6.37	L->A	Interferes with DRY motif function	AA2AR (5IU4), NTR1 (4GRV)



# X3.41W – “Roth” Mutation



(c)

	3.30		3.40		3.50
<i>b1AR</i>				*	
<i>b2AR</i>				*	
<i>D1AR</i>				*	
<i>D2AR</i>				*	
<i>a1aR</i>				*	
<i>a2bR</i>				*	
<i>HT5A</i>				*	
<i>Rho</i>				*	

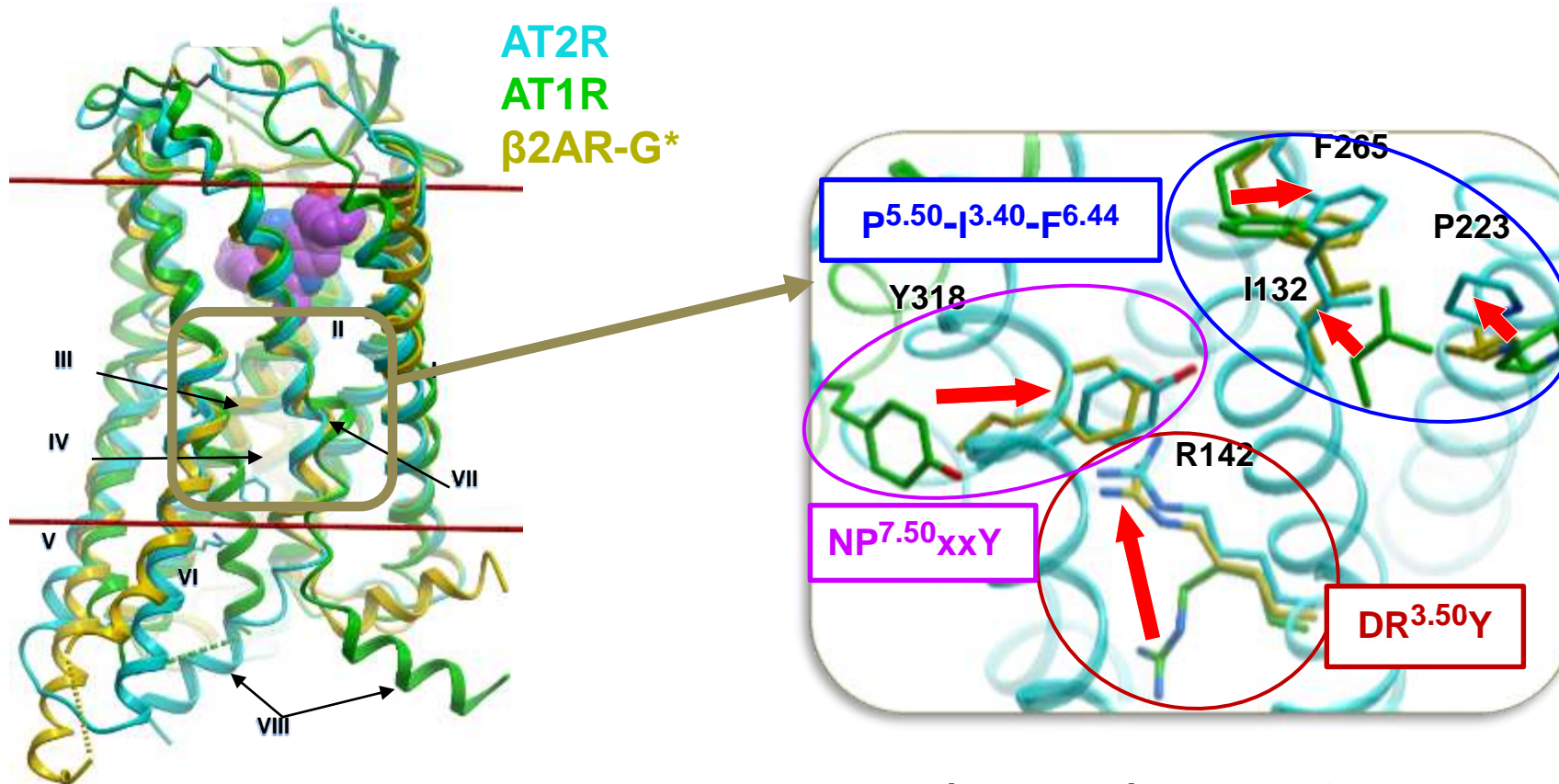
TM3

- Stabilizing or neutral in most Class A GPCRs
- Helped to solve >6 receptors, including 2 in active-like state

# Activation Related Changes in Microswitches

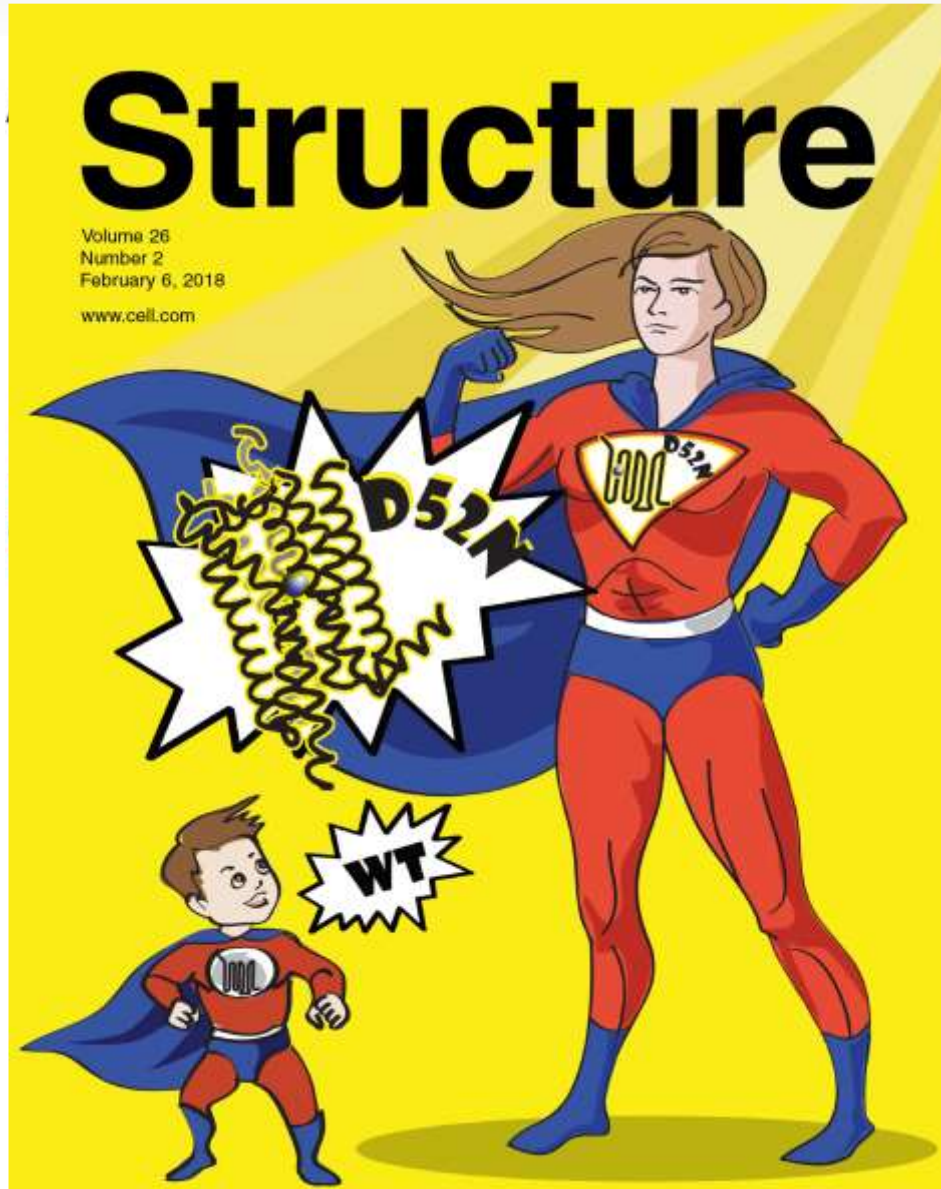
Removing switches can decouple from agonist and limit natural motions of receptor

3.40	I->V, A	P-I-F microswitch motif
3.49	D,G->A	DRY motif
6.37	L->A	Interferes with DRY motif function



Zhang H et al, *Nature*. 2017;544:327-32.

# Stabilizing mutations in conserved Na<sup>+</sup> site



	APO T <sub>M</sub> °C	NECA T <sub>M</sub>	Theophylline T <sub>M</sub>	ZM241385 T <sub>m</sub>
WT	47	53	53	62
D2.50A	47	58	54	60
S3.39A	50	61	55	63
N7.49A	54	64	56	64
<b>D2.50N</b>	<b>58</b>	<b>64</b>	<b>59</b>	<b>66</b>

D52N–UK432097 (agonist)  
S91A–UK432097 (agonist)



Katritch et al 2014, *TiBS*, **39**, 233  
White et al 2018, *Structure* **26**, 259



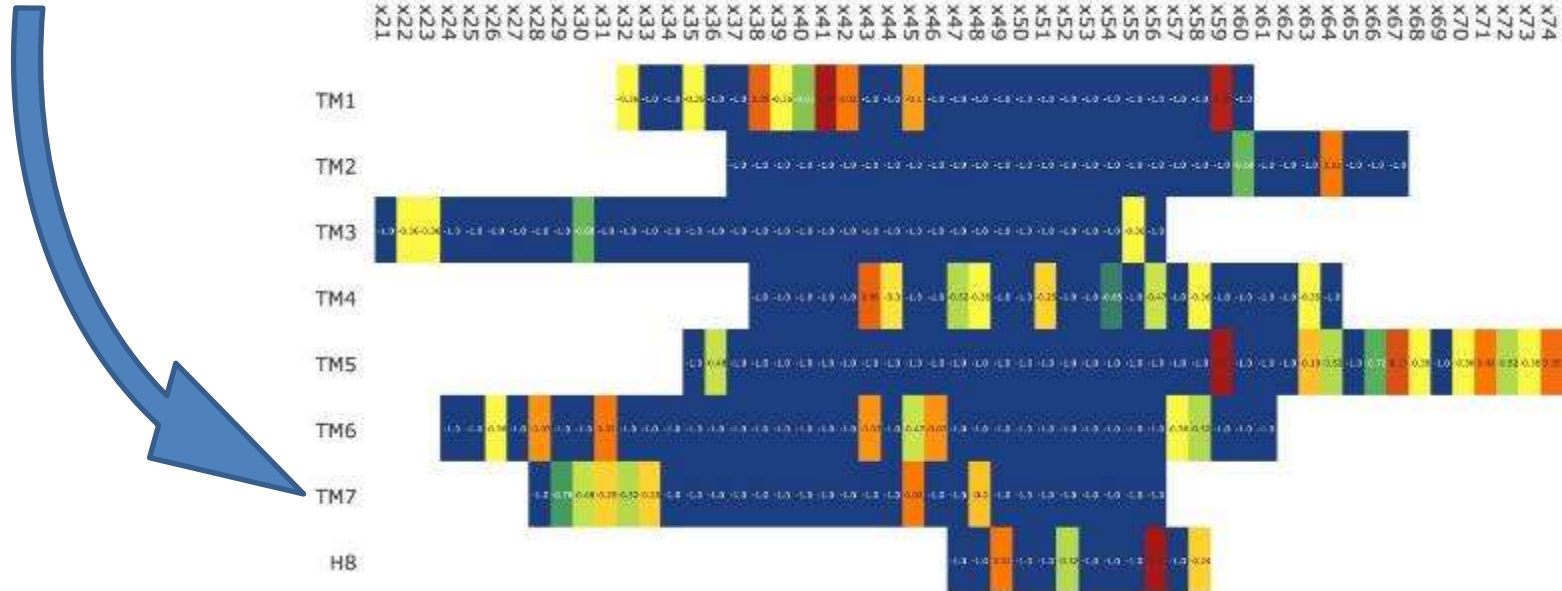
# Sequence-Based

- Goal: identify (and replace) “outliers”: residues which are rarely observed at certain position
- Some positions are highly variable – take into account relative conservation
- Results depend on the GPCR set – use 3 levels of sequence clustering:
  - GPCR Class or branch (SI  $\geq$  25%)
  - GPCR family (SI  $\geq$  30%)
  - GPCR subfamily (SI  $\geq$  35%)

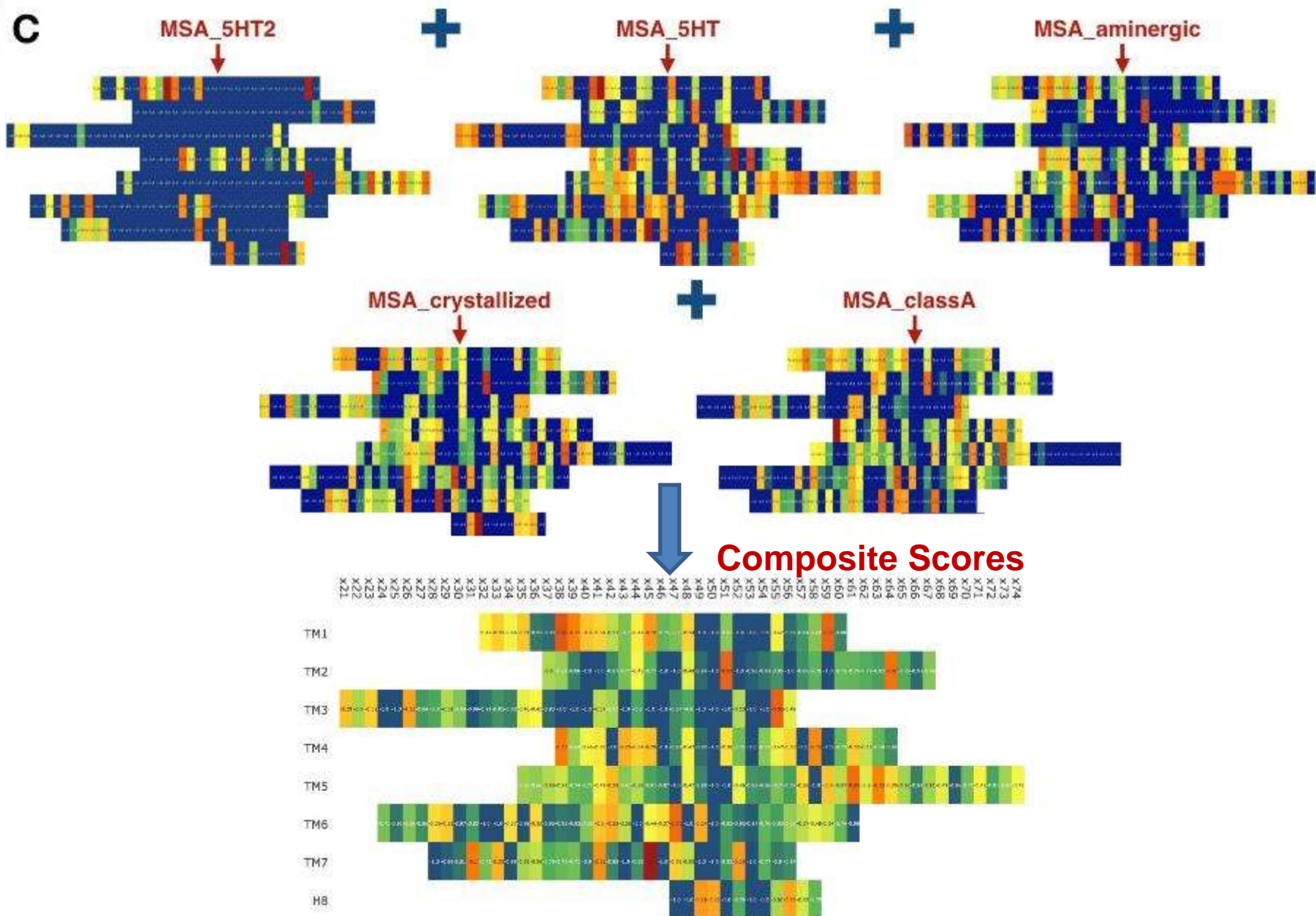
GPCR ID	1	2	3	4	5
GPCR 1	L	V	L	P	W
GPCR 2	L	L	L	P	W
GPCR 3	L	A	L	P	W
GPCR 4	L	T	L	P	W
...	...	...	...	...	...
Target GPCR	L	S	L	S	W

# Sequence-Based: Building Scoring Matrix

UniProt ID	...	7.36x35	7.37x36	7.38x37	7.39x38	7.40x39	7.41x40	7.42x41	7.43x42	7.44x43	7.45x45	7.46x46	7.47x47	7.48x48	7.49x49	7.50x50	...
5ht2a_bovin	...	N	V	F	V	W	I	G	Y	L	S	S	A	V	N	P	...
5ht2a_canif	...	N	V	F	V	W	I	G	Y	L	S	S	A	V	N	P	...
5ht2a_crigr	...	N	V	F	V	W	I	G	Y	L	S	S	A	V	N	P	...
5ht2a_drome	...	S	L	F	L	W	L	G	Y	F	N	S	T	L	N	P	...
5ht2a_human	...	N	V	F	V	W	I	G	Y	L	S	S	A	V	N	P	...
5ht2a_macmu	...	N	V	F	V	W	I	G	Y	L	S	S	A	V	N	P	...
5ht2a_mouse	...	N	V	F	V	W	I	G	Y	L	S	S	A	V	N	P	...
5ht2a_pig	...	N	V	F	V	W	I	G	Y	L	S	S	A	V	N	P	...
5ht2a_porpy	...	N	V	F	V	W	I	G	Y	L	S	S	A	V	N	P	...
5ht2a_rat	...	N	V	F	V	W	I	G	Y	L	S	S	A	V	N	P	...
5ht2b_drome	...	S	L	F	L	W	L	G	Y	F	N	S	T	L	N	P	...
5ht2b_human	...	E	I	F	V	W	I	G	Y	V	S	S	G	V	N	P	...
5ht2b_mouse	...	E	I	F	V	W	I	G	Y	V	S	S	G	V	N	P	...
5ht2b_rat	...	Q	I	F	V	W	V	G	Y	V	S	S	G	V	N	P	...
5ht2b_tetfl	...	E	I	F	S	W	V	G	Y	V	S	S	G	I	N	P	...
5ht2c_canlf	...	N	V	F	V	W	I	G	Y	V	C	S	G	I	N	P	...
5ht2c_human	...	N	V	F	V	W	I	G	Y	V	C	S	G	I	N	P	...
5ht2c_mouse	...	N	V	F	V	W	I	G	Y	V	C	S	G	I	N	P	...
5ht2c_pantr	...	N	V	F	V	W	I	G	Y	V	C	S	G	I	N	P	...
5ht2c_rat	...	N	V	F	V	W	I	G	Y	V	C	S	G	I	N	P	...

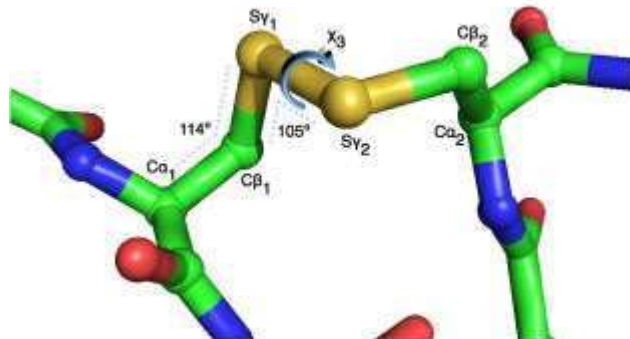
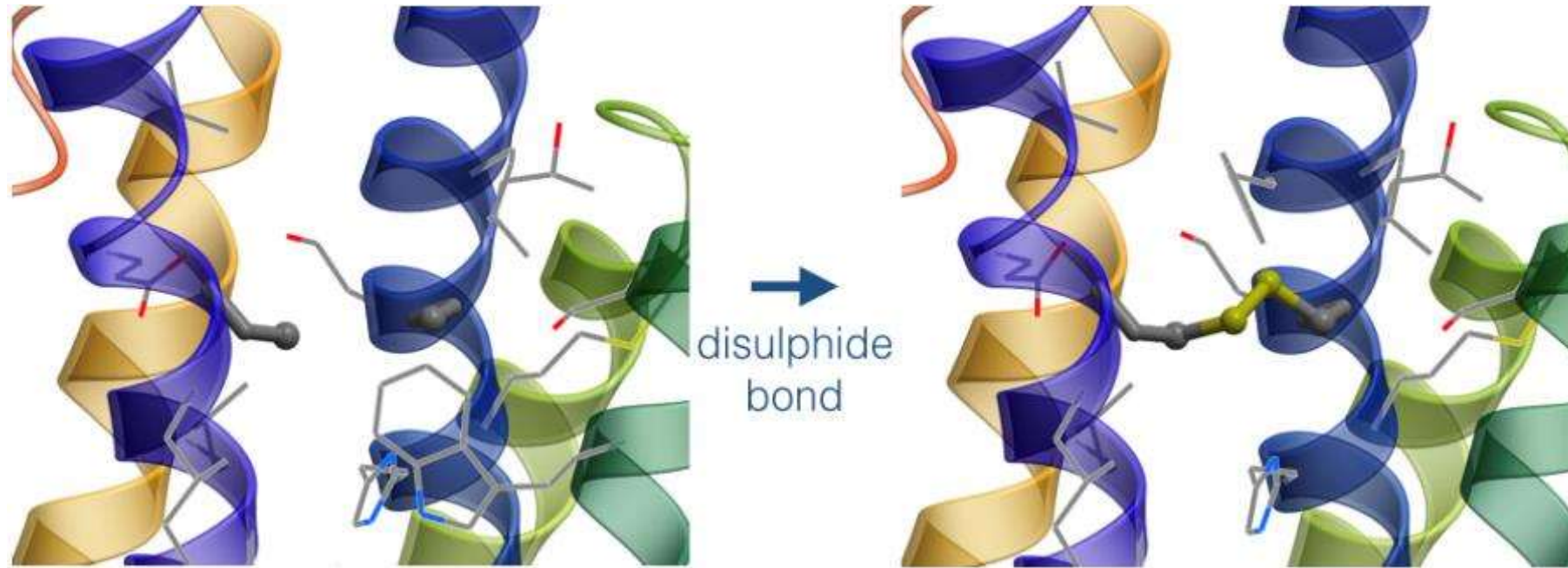


# Sequence-Based: Building Scoring Matrix



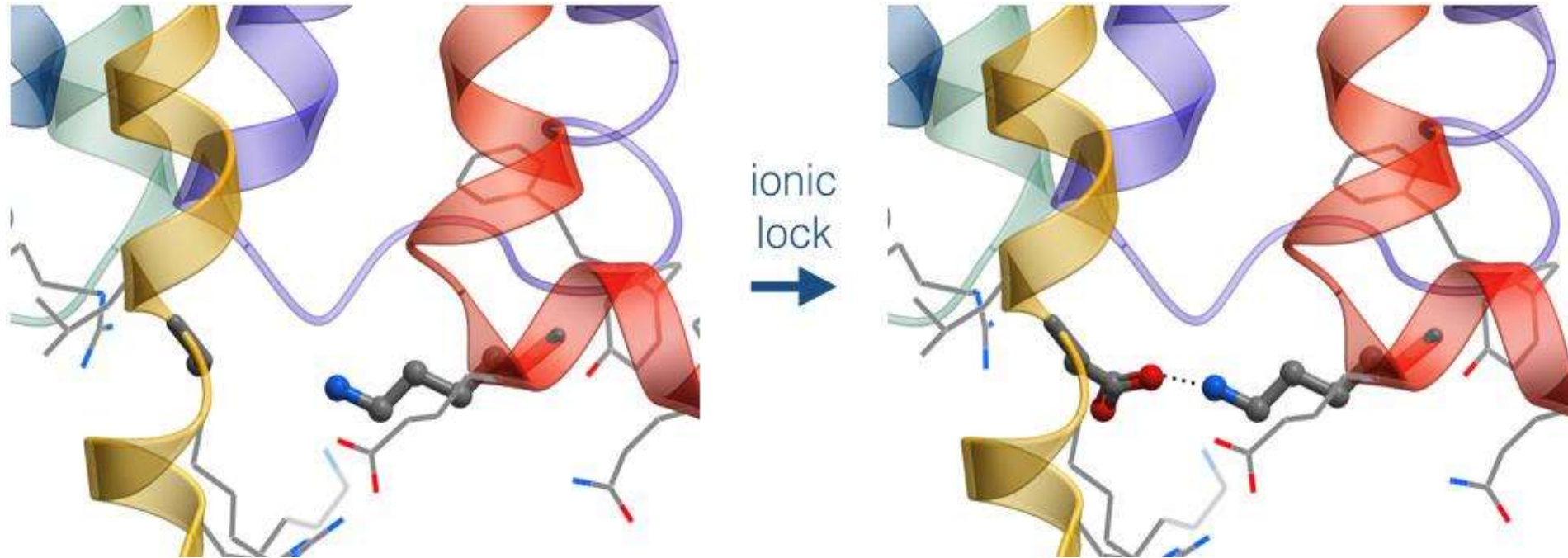


# Structure-based: Design of Disulfide Bridges



- Find a pair of residues with permissive geometry
- Full energy optimization and scoring energy strain

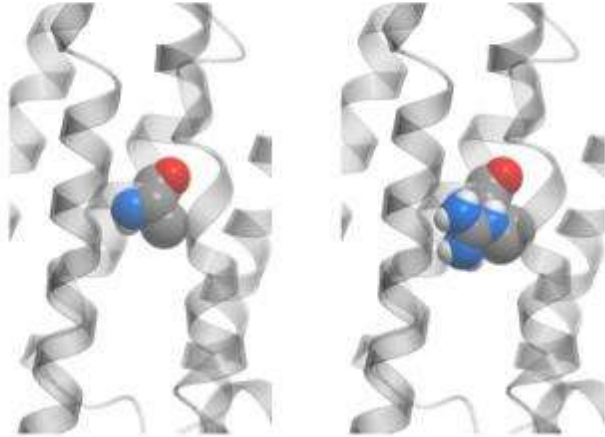
# Structure-based: Design of ionic locks



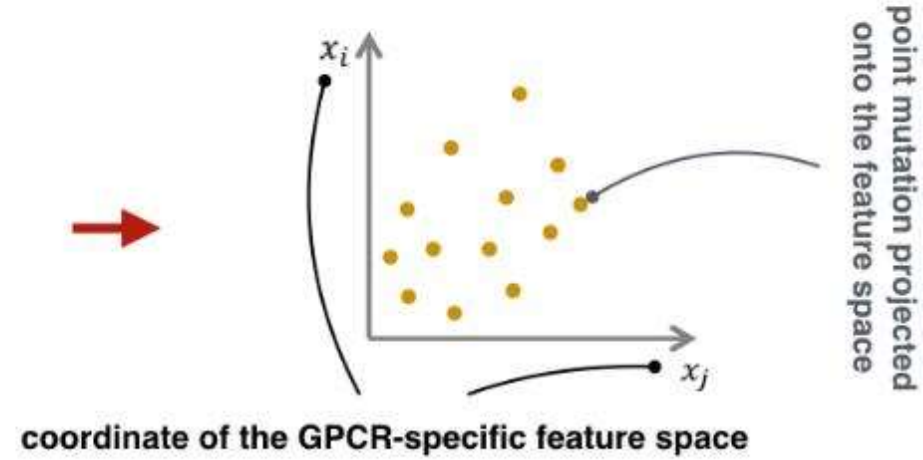
- Find a pair of residues with permissive geometry
- Full energy optimization and scoring energy strain

# Machine Learning

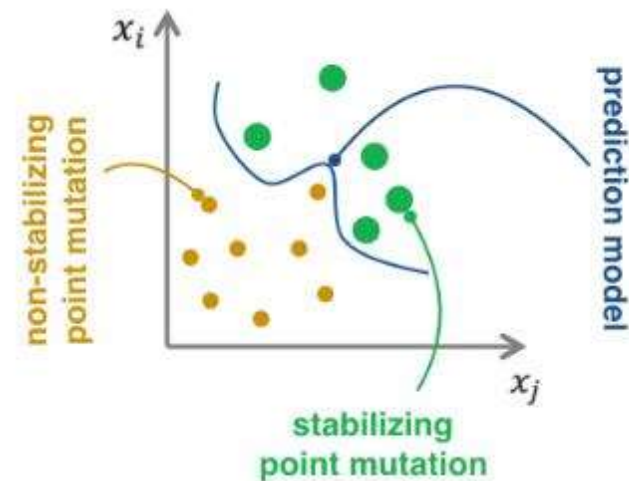
**A** structural models of point mutations



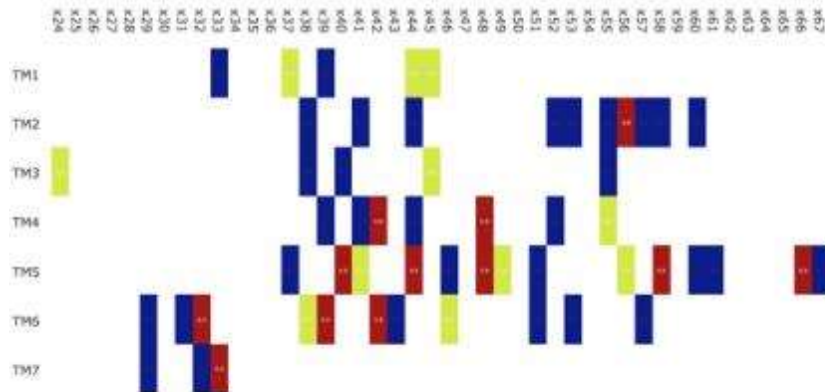
**B** projection into the feature space



**C** applying of prediction model

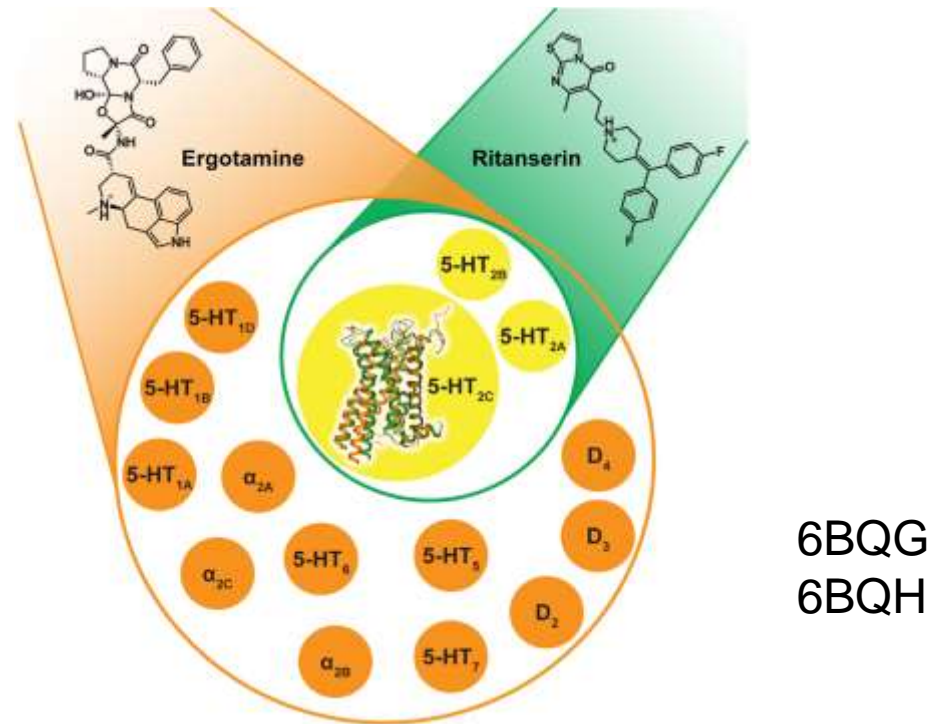


**D** scoring matrix





# Prospective Application to Mutant Discovery: 5HT<sub>2C</sub> structures

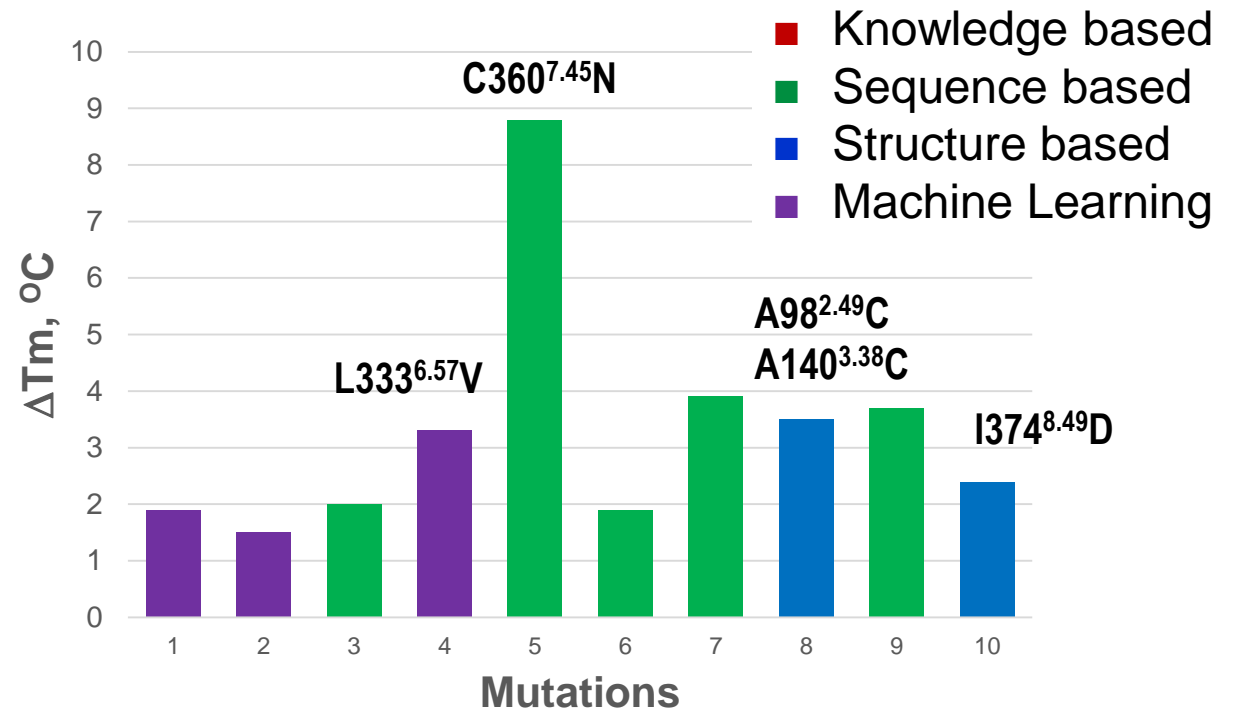


Peng Y et al. (2018) 5-HT<sub>2C</sub> Receptor Structures Reveal the Structural Basis of GPCR Polypharmacology. *Cell* 172(4):719-30

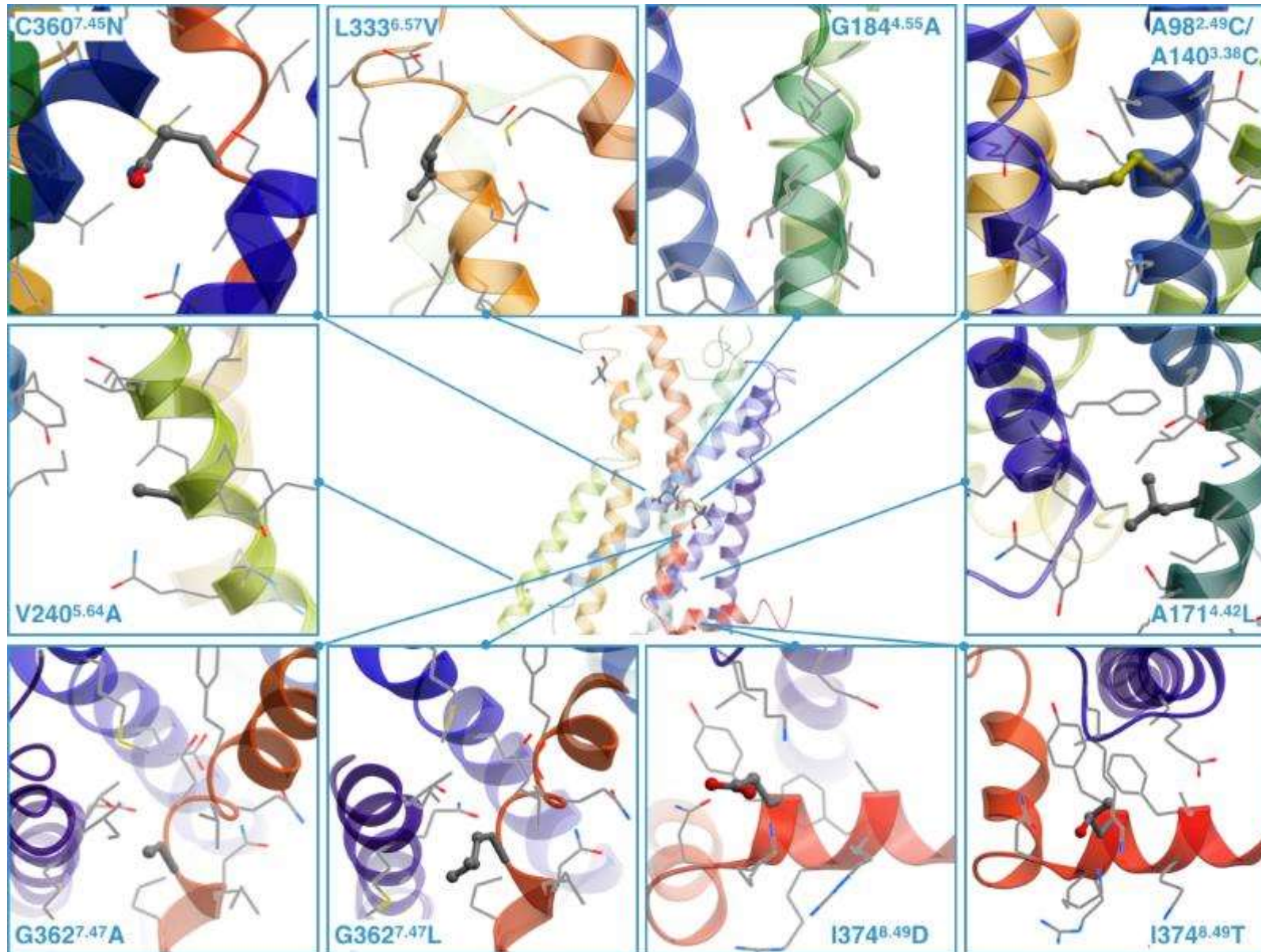
# CompoMug prospective screening: 5HT<sub>2C</sub>

Mutation	CompoMug Module	aSEC* quality	Tm (°C) ±SEM	ΔTm (C)
WT			50.4 ± 0.8	0.0
I62 <sup>1.41</sup> V	Seq-based	~		-0.7
G69 <sup>1.48</sup> A	Seq-based	-		-1.4
D99 <sup>2.50</sup> N	Knowledge	-		-
H85 <sup>12.51</sup> E	Struct-based	N/A		-
G103 <sup>2.54</sup> A	Seq-based	-		-4.4
Y125 <sup>3.23</sup> K	Seq-based	-		-2.0
Y125 <sup>3.23</sup> V	Seq-based	~		-0.7
M143 <sup>3.41</sup> W	Knowledge	-		0.6
R157 <sup>3.55</sup> T	ML & Seq-based	-		-1.8
R157 <sup>3.55</sup> Q	Seq-based	-		-2.0
T169 <sup>4.40</sup> K	Seq-based	+		0.2
<b>A171<sup>4.42</sup>L</b>	<b>ML</b>	<b>~</b>	<b>52.3 ± 1.2</b>	<b>1.9</b>
I172 <sup>4.43</sup> A	Seq-based	-		1.1
I172 <sup>4.43</sup> F	Seq-based	~		0.6
<b>G184<sup>4.55</sup>A</b>	<b>ML</b>	<b>+</b>	<b>51.9 ± 0.1</b>	<b>1.5</b>
N203 <sup>ECL2</sup> D	Struct-based	-		-2.6
F220 <sup>5.45</sup> I	ML	~		0.0
F224 <sup>5.48</sup> Y	ML & Seq-based	-		-3.3
C235 <sup>5.59</sup> F	Seq-based	~		0.1
L236 <sup>5.60</sup> R	ML & Seq-based	N/A		-
<b>V240<sup>5.64</sup>A</b>	<b>Seq-based</b>	<b>+</b>	<b>52.4 ± 0.5</b>	<b>2.0</b>
V240 <sup>5.64</sup> S	Seq-based	+		0.3
G314 <sup>6.38</sup> A	ML	-		-4.0
<b>L333<sup>6.57</sup>V</b>	<b>ML &amp; Seq-based</b>	<b>+</b>	<b>53.7 ± 0.6</b>	<b>3.3</b>
K348 <sup>7.32</sup> A	Seq-based	-		-4.4
<b>C360<sup>7.45</sup>N</b>	<b>Seq-based</b>	<b>+</b>	<b>59.2 ± 0.5</b>	<b>8.8</b>
<b>G362<sup>7.47</sup>L</b>	<b>Seq-based</b>	<b>+</b>	<b>52.3 ± 0.7</b>	<b>1.9</b>
<b>G362<sup>7.47</sup>A</b>	<b>Seq-based</b>	<b>+</b>	<b>54.3 ± 0.7</b>	<b>3.9</b>
L370 <sup>7.55</sup> D	Struct-based	-		-2.3
K373 <sup>8.48</sup> E	Struct-based	-		-0.4
<b>I374<sup>8.49</sup>D</b>	<b>Struct-based</b>	<b>+</b>	<b>53.9 ± 0.8</b>	<b>3.5</b>
<b>I374<sup>8.49</sup>T</b>	<b>Seq-based</b>	<b>+</b>	<b>54.1 ± 0.9</b>	<b>3.7</b>
Y375 <sup>8.50</sup> F	Seq-based	-		-2.4
N381 <sup>8.56</sup> R	Sequence-based	~		0.6
T67 <sup>1.46</sup> C/G103 <sup>2.54</sup> C	Struct-based	-		-
V74 <sup>1.53</sup> C/A96 <sup>2.47</sup> C	Struct-based	-		-
A87 <sup>2.38</sup> C/A171 <sup>4.42</sup> C	Struct-based	~		-
<b>A98<sup>2.49</sup>C/A140<sup>3.38</sup>C</b>	<b>Struct-based</b>	<b>~</b>	<b>52.8 ± 1.0</b>	<b>2.4</b>
T369 <sup>7.54</sup> C/Y375 <sup>8.50</sup> C	Struct-based	N/A		-

- 40 mutations tested
- 10 improved both Tm & SEC (~25%)
- Best shift ΔTm ~ 9°C
- 3 components contributed, but not Knowledge-based

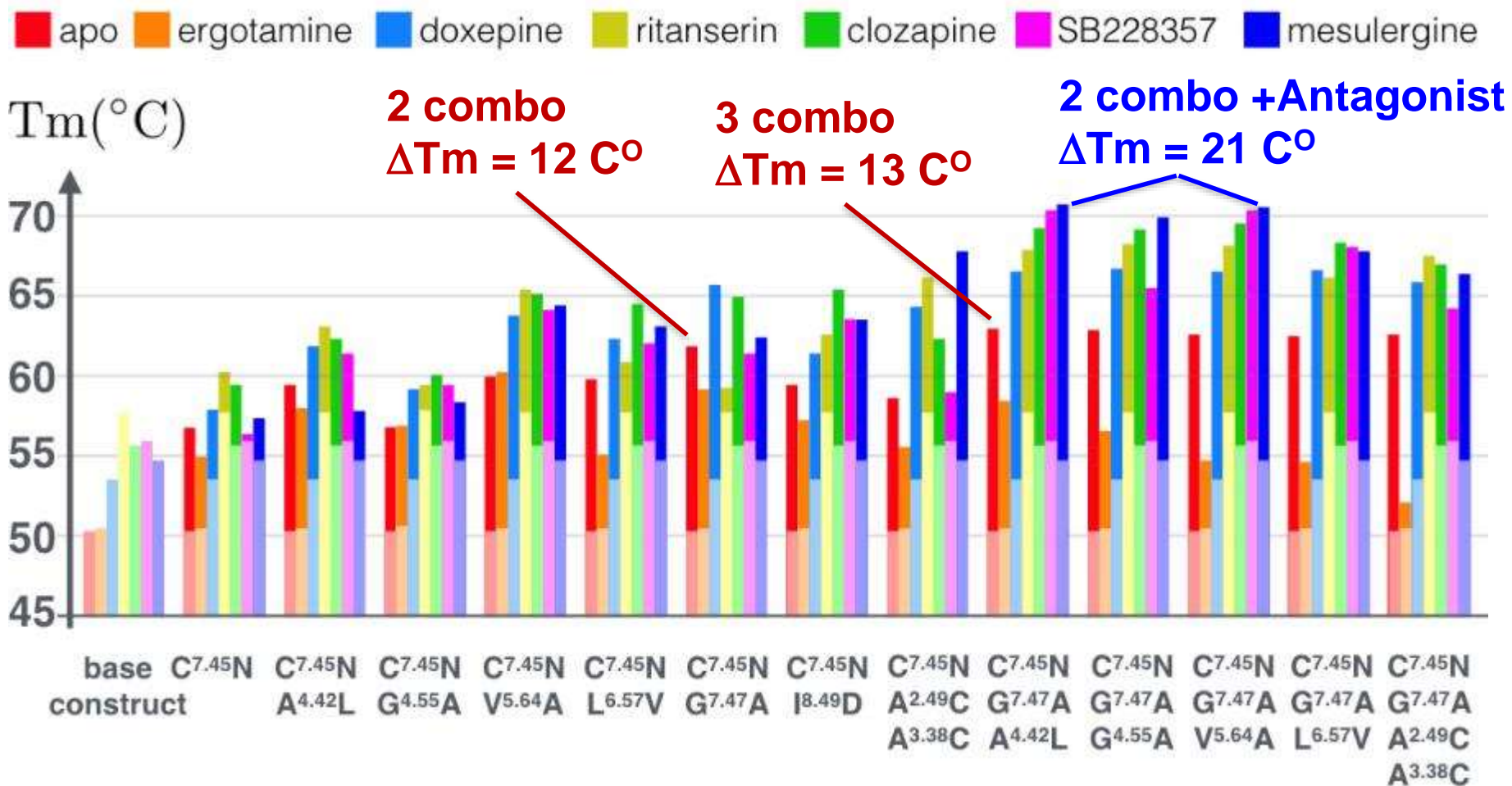


# 5HT<sub>2C</sub>: Mutations in the structural model



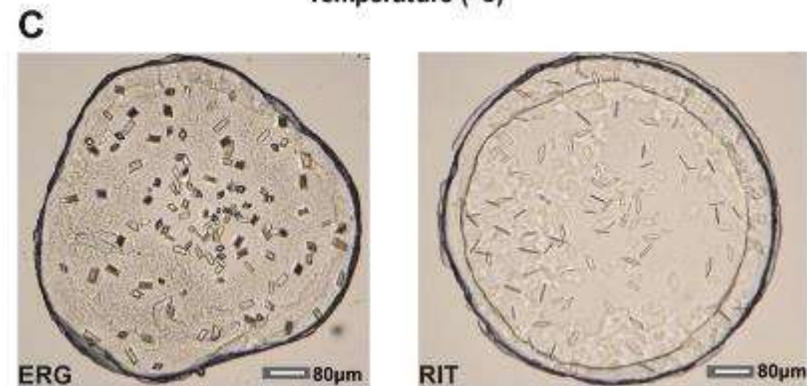
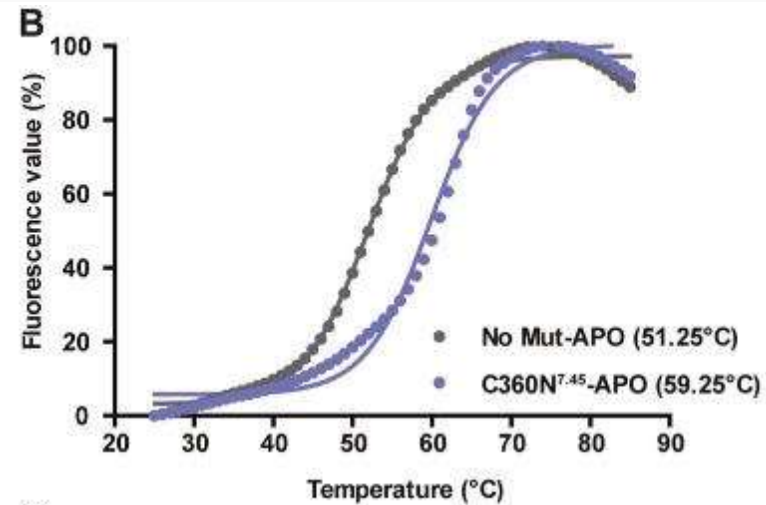
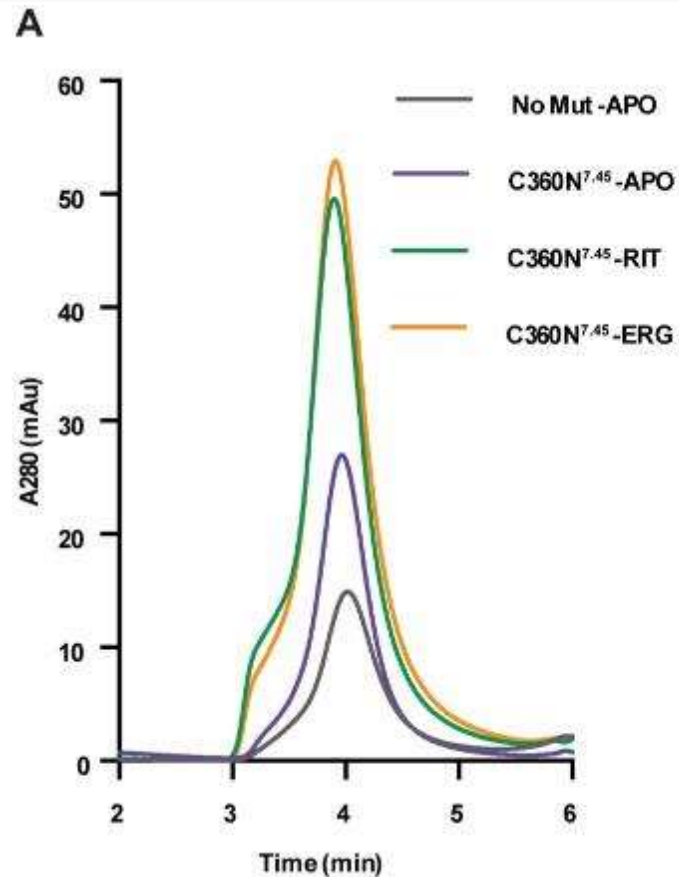


# 5HT<sub>2C</sub>: Combined mutations in complex with ligands

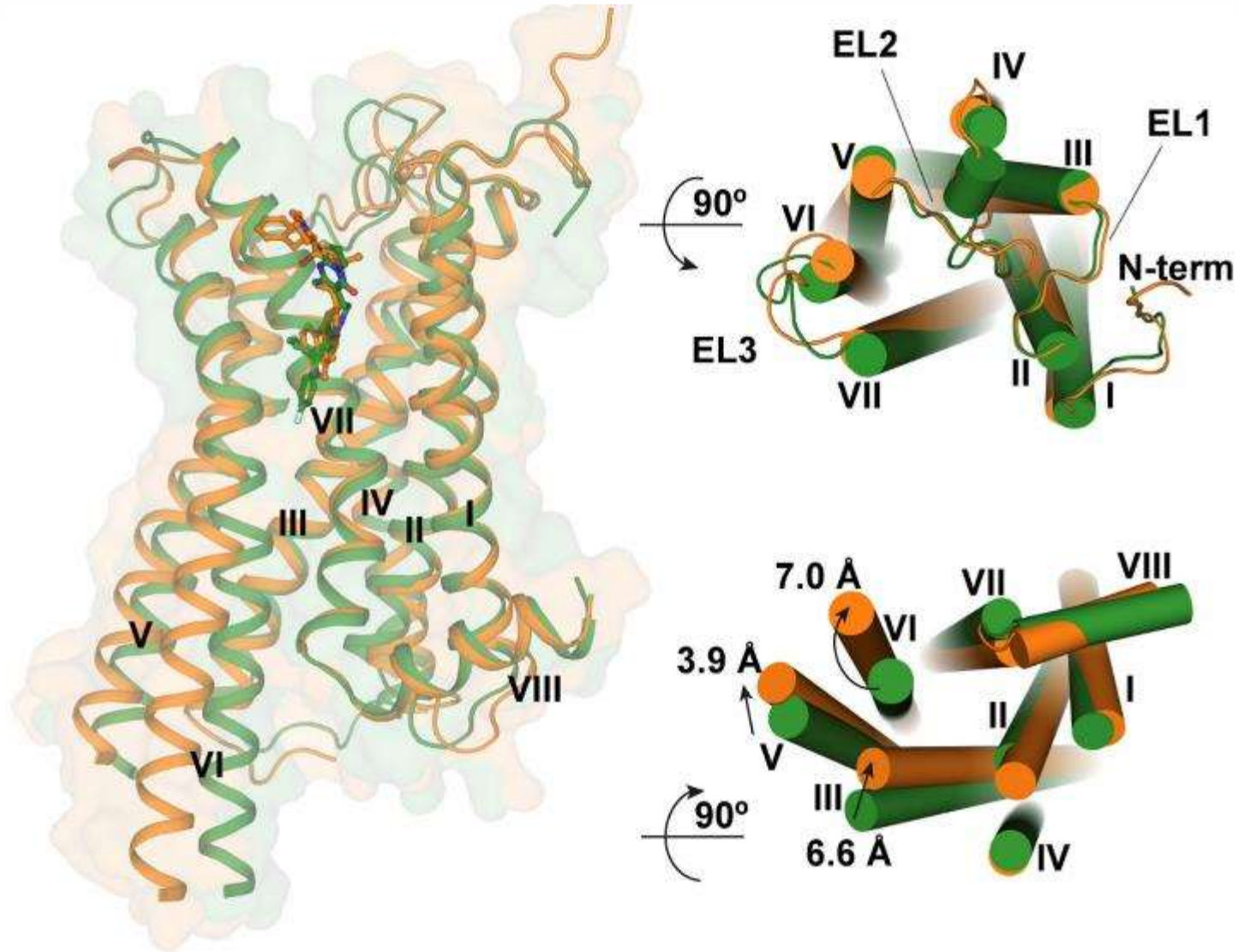


# 5-HT<sub>2C</sub>: Crystals and structures

- Combo mutations yield first 5-HT<sub>2C</sub> crystals with antagonists
- Single C360<sup>7.45</sup>N allowed crystallization of both agonist and antagonists
- Structures for agonist Ergotamine (3.0 Å) and antagonist Ritanserin (2.7 Å)



# 5-HT<sub>2C</sub> inactive and active-like structures





# More CompoMug predictions tested

Target	# tested single mutants	Hit rate	Best $\Delta T_m$ , C°	Best $\Delta T_m$ , C° combined mutations	Crystallized /Solved	Comment
5HT2C	40	25%	9	13	Yes/Yes	1 mutant in structure
Target #2	40	25%	6	7	Yes/Yes	2+4 mutants in structure
Target #3	60	17%	10			
Target #4	36	30%	5	9	Yes/Yes	5 mutations in structure
Target #5	40	20.0%	4			
Target #6	60	12%	4			
Target #7	40	10.0%	3	4		
Target #8	60	7%	4			
Target #9	60	25%	3	16	Yes/Yes	non Class A; 4 mutants
Target #10	60	11%	3			
EP3	30	15%	0		Yes/Yes	Only Improved Diffraction
Taste /Class C	40-60	0%				

# CompoMug: Summary & Outlook

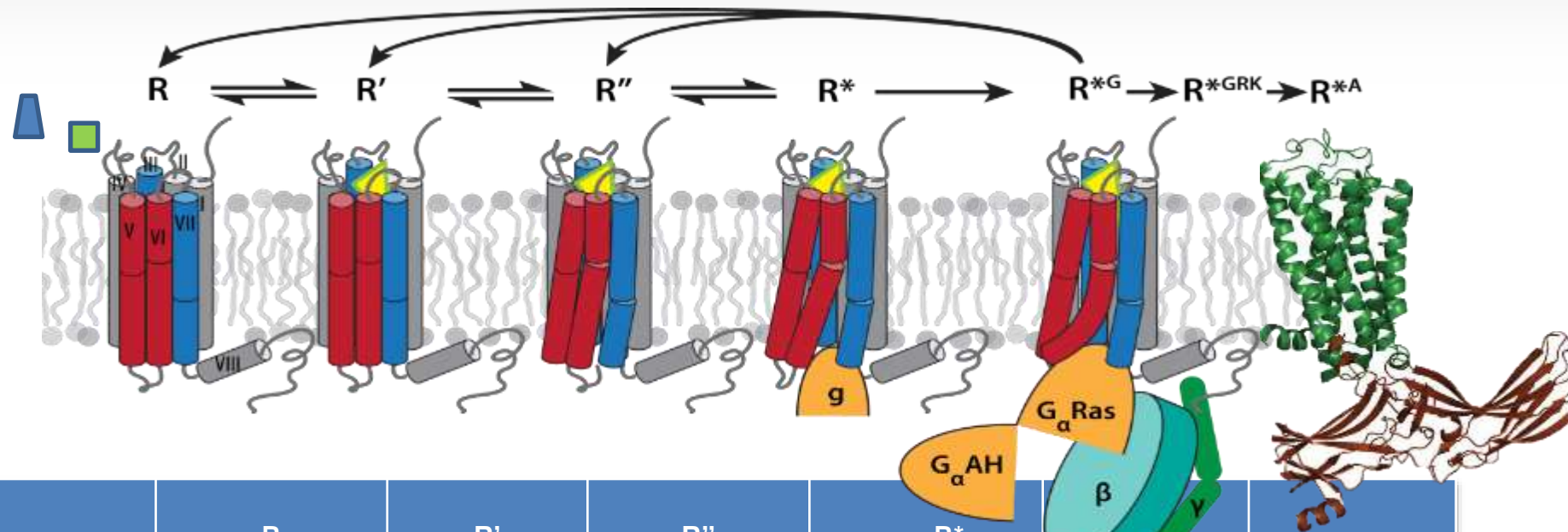
- **CompoMug** already shows 15-30% “hit rate” for most GPCRs targets, and up to 16C° combined  $\Delta T_m$
- Complementary to other engineering approaches
- All 4 modules make important contributions:
  - **Sequence-based** module most universally applicable
  - **Energy-based predictions** can improve with each GPCR structure (better homology models)
  - **Machine learning** continuously can improve with each new stability dataset obtained
- **Utility for both SBDD and for assay development!**

# Outline

- Rational prediction of stabilizing mutations:  
CompoMug
- **New insights into GPCR function and allosteric mechanisms**
- Structure-Based ligand discovery for GPCRs



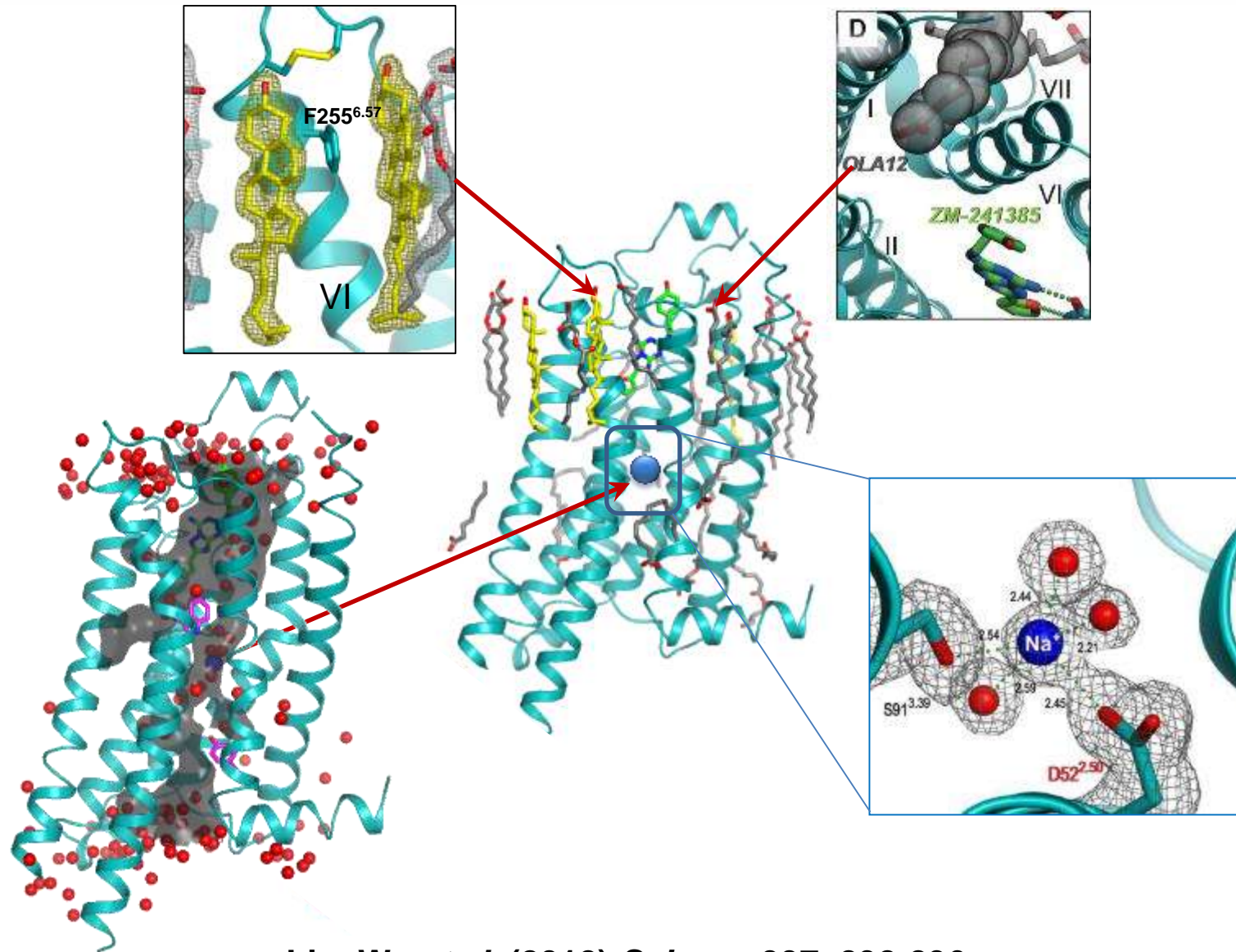
# Dynamic Mechanisms of GPCR action on atomic level



Receptor	R Inactive Ground State	R' Inactive Agonist Bound	R'' Active-Like Agonist Bound	R* Active, G-protein or Mimic Binding	R*G G-protein Signaling	R*A β-arrestin Complex
Adenosine A <sub>2A</sub>	3EML,3REY, 3RFM, 3PWH, <b>4E1Y</b>	--	<b>3QAK</b> ,2YDO,2YDV, 4UHR, 4UG2	--	<b>5G53</b>	
β <sub>1</sub> -Adrenergic	2VT4,2YCW,2YCX, 2YCY,2Y CZ	2Y00 -2Y04	--	--	--	
β <sub>2</sub> -Adrenergic	2RH1, 2R4R, 2R4S,3D4S	3PDS	--	3P0G <sup>c</sup> (3.5)	<b>3SN6</b>	
Rhodopsin	1F88,1U19	2G87, 2HPY	3CAP <sup>f</sup> ,	3DQB,2X72,3PQR, 3PXO	--	<b>4ZWJ</b>
Muscarinic	<b>3UON</b>			<b>4MGS,4MGT</b>		
Opioid	<b>4DKL</b>			<b>5C1M, κ-OR</b>		
CB1	<b>5U09</b>		<b>5XRA</b>			
P2Y12	<b>4NTJ</b>	<b>4PXZ</b>				
NTSR1, ETB <sub>4</sub> 5HT1B, 5HT2B		<b>5GLH</b>	<b>4GRV,</b> <b>4IAQ,4IAR, 4IB4</b>			

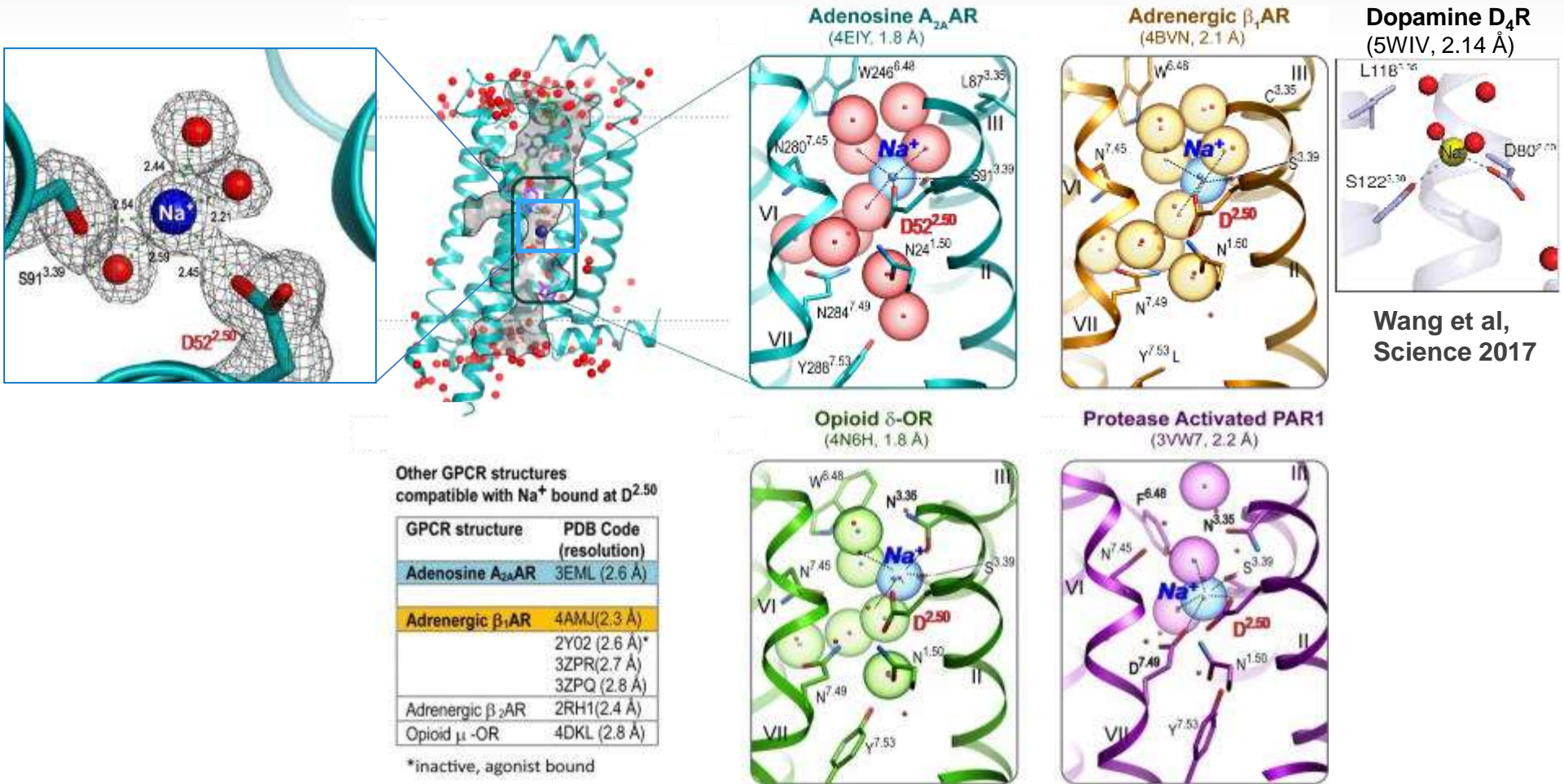
Updated from Katritch, Stevens, Cherezov,  
*Annual Rev. Pharmacology & Toxicology*, 2013

# Endogenous allosteric molecules in 1.8 Å Structure of human A<sub>2A</sub>AR



Liu, W., et al. (2012) *Science* 337, 232-236

# Allosteric Na<sup>+</sup> site in class A GPCR structures

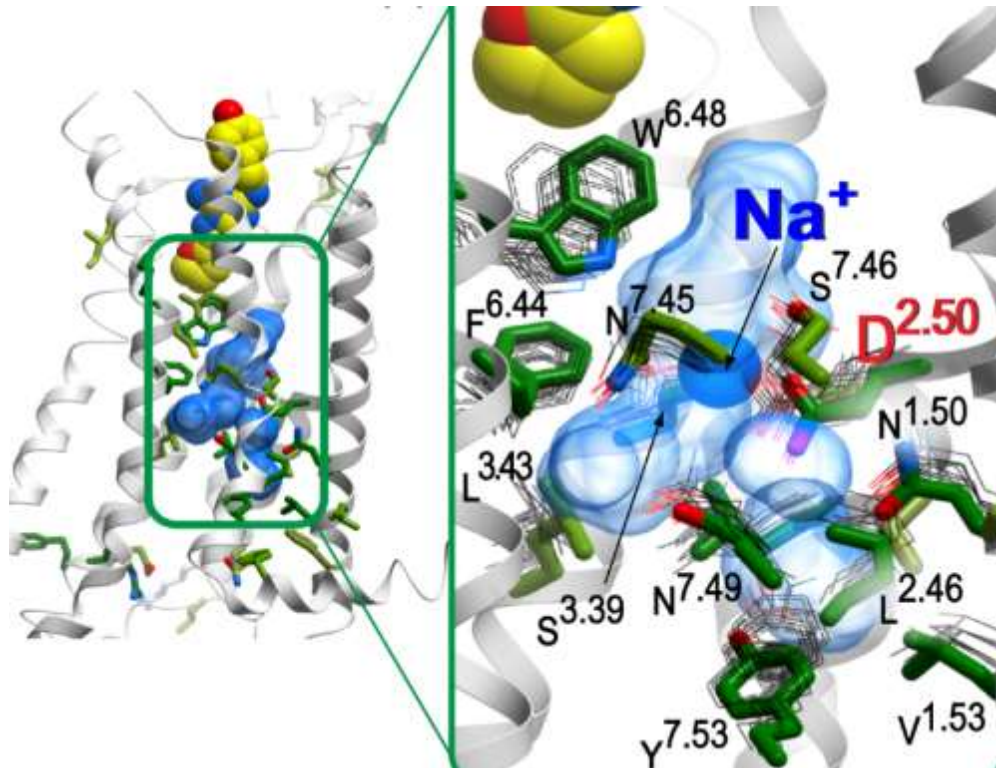


- Na<sup>+</sup> crystallographically observed in α, δ, γ branches of Class A
- The pocket highly conserved in most class A GPCRs



# Na<sup>+</sup> Pocket Conservation in Class A GPCRs

- Largest conserved 3D cluster (15 residues of 34 most conserved residues)
- Includes **NPxxY** in helix VII and **FxxCW** in helix VI, but **not DRY** in helix III
- **D2.50** conserved in 95% of GPCRs



(c)

	1.50	1.53	2.46	2.47	2.49	2.50	3.35	3.39	3.43	6.44	6.48	7.45	7.46	7.49	7.50	7.53
<b>A<sub>2A</sub>AR*</b>	N	V	L	A	A	D	L	S	L	F	W	N	S	N	P	Y
<b>β<sub>1</sub>AR*</b>	N	V	L	A	A	D	C	S	L	F	W	N	S	N	P	Y
<b>β<sub>2</sub>AR</b>	N	V	L	A	A	D	C	S	L	F	W	N	S	N	P	Y
<b>ACM2</b>	N	V	L	A	A	D	V	S	L	F	W	N	S	N	P	Y
<b>ACM3</b>	N	V	L	A	A	D	A	S	L	F	W	N	S	N	P	Y
<b>5HT<sub>1B</sub></b>	N	V	L	A	T	D	C	S	L	F	W	N	S	N	P	Y
<b>5HT<sub>2B</sub></b>	N	V	L	A	A	D	F	S	L	F	W	S	S	N	P	Y
<b>H<sub>1</sub>R</b>	N	V	L	S	A	D	A	S	V	F	W	N	S	N	P	Y
<b>D<sub>3</sub>R</b>	N	V	L	A	A	D	M	S	L	F	W	N	S	N	P	Y
<b>S1P<sub>1</sub>R</b>	N	V	L	A	S	A	S	L	L	F	W	N	S	N	P	Y
<b>Rho</b>	N	T	L	A	A	D	G	A	L	F	W	A	A	N	P	Y
<b>δ-OR*</b>	N	V	L	A	A	D	N	S	L	F	W	N	S	N	P	Y
<b>κ-OR</b>	N	V	L	A	A	D	N	S	L	F	W	N	S	N	P	Y
<b>μ-OR</b>	N	V	L	A	A	D	N	S	L	F	W	N	S	N	P	Y
<b>NOP</b>	N	V	L	A	A	D	N	S	L	F	W	N	S	N	P	Y
<b>CXCR4</b>	N	V	L	S	A	D	N	S	L	F	W	H	C	N	P	Y
<b>NTSR1</b>	N	T	L	A	S	D	C	T	V	F	W	S	S	N	P	Y
<b>PAR1*</b>	N	A	L	A	A	D	N	S	M	F	F	S	C	D	P	Y

α

γ

β

δ

# Only 5% of Class A GPCRs lack D2.50

	Uniprot ID	Description*	Protein name	#2.50	Other Acid
1	VNRL4_HUMAN	PSEUDOGENE	Putative vomeronasal receptor-like protein 4	H	
2	CCBP2_HUMAN	DECOY RECEPTOR D6	Chemokine-binding protein 2	N	
3	CCRL2_HUMAN	Non-signaling	C-C chemokine receptor-like 2	N	
4	GNRR2_HUMAN	NON_FUNCTIONAL	Putative gonadotropin-releasing hormone II receptor	F	
5	GPR26_HUMAN	Orphan, Constitutively active	GPCR 26		
6	GPR78_HUMAN	Orphan, Constitutively active	GPCR 78		
7	GP161_HUMAN	Orphan, negative regulator of Shh pathway	GPCR 161	N	
8	LGR4_HUMAN	ORPHAN, binds Frizzled and LRPs	Leucine-rich repeat-containing GPCR 4	N	
9	LGR5_HUMAN	ORPHAN, binds Frizzled and LRPs	Leucine-rich repeat-containing GPCR 5	N	
10	LGR6_HUMAN	ORPHAN, binds Frizzled and LRPs	Leucine-rich repeat-containing GPCR 6	N	
11	NTR2_HUMAN	Low affinity to NT, constitutively active [19, 20]	Neurotensin receptor type 2	G	
12	GP141_HUMAN	Probable GPCR	Probable G protein coupled receptor 141	H	
13	GP146_HUMAN	Probable GPCR	Probable GPCR 146	A	
14	GP148_HUMAN	Probable GPCR	Probable GPCR 148	Y	
15	GP150_HUMAN	Probable GPCR	Probable GPCR 150	F	
16	GP153_HUMAN	Probable GPCR	Probable GPCR 153	V	
17	GP162_HUMAN	Probable GPCR	Probable GPCR 162	V	
18	GP176_HUMAN	Probable GPCR	Probable GPCR 176 (HB-954)	I	
19	GPR21_HUMAN	Probable GPCR	Probable GPCR 21	A	
20	GPR22_HUMAN	Probable GPCR	Probable GPCR 22	V	
21	GPR33_HUMAN	Probable GPCR	Probable GPCR 33	Y	
22	GPR52_HUMAN	Probable GPCR	Probable GPCR 52	A	
23	GPR62_HUMAN	Probable GPCR	Probable GPCR 62	A	
24	GPR75_HUMAN	Probable GPCR	Probable GPCR 75	L	
25	GPR82_HUMAN	Probable GPCR	Probable GPCR 82	N	
26	O10J6_HUMAN	Putative olfactory	Putative olfactory receptor 10J6	K	
27	OR2G2_HUMAN	Olfactory	Olfactory receptor 2G2	Y	E3.39
28	OR4F6_HUMAN	Olfactory	Olfactory receptor 4F6	N	E3.39
29	OR5BH_HUMAN	Olfactory	Olfactory receptor 5B17	G	E3.39
30	OR8J1_HUMAN	Olfactory	Olfactory receptor 8J1	N	E3.39
31	OR8J2_HUMAN	Olfactory	Olfactory receptor 8J2	N	E3.39
32	OR8J3_HUMAN	Olfactory	Olfactory receptor 8J3	N	E3.39
33	GP143_HUMAN	Signals	GPCR 143 (Ocular albinism type 1)	A	D3.39
34	GPBAR_HUMAN	Signals	G protein coupled bile acid receptor 1	G	D7.50
35	GNRHR_HUMAN	Signals	Gonadotropin-releasing hormone receptor	N	D7.49
36	OPSB_HUMAN	Opsin, no Na <sup>+</sup> binding	Blue-sensitive opsin	G	

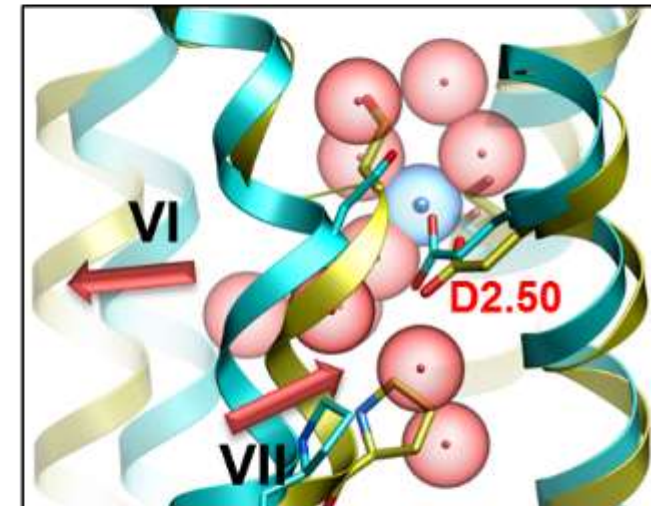
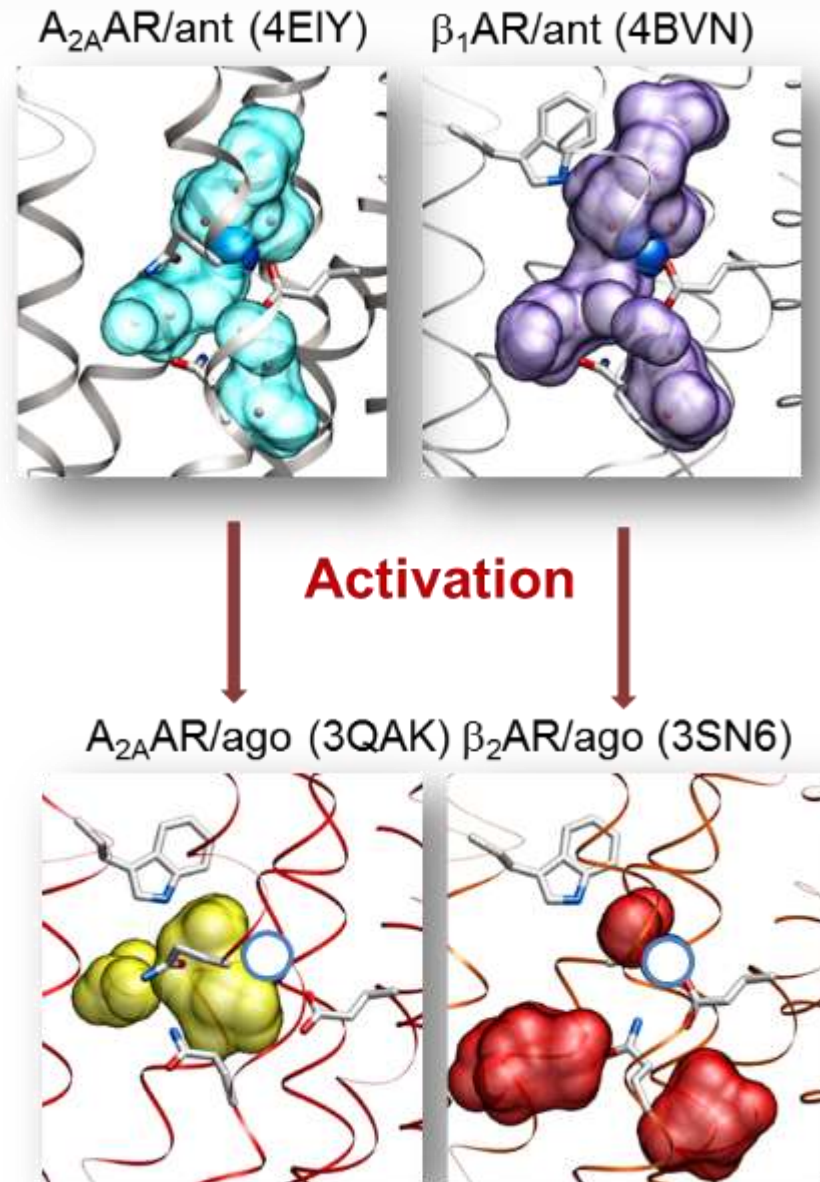
Orphans,  
Constitutive,  
Non-signaling

Probable/  
Putative

Have alternative  
acidic side chain  
In the pocket

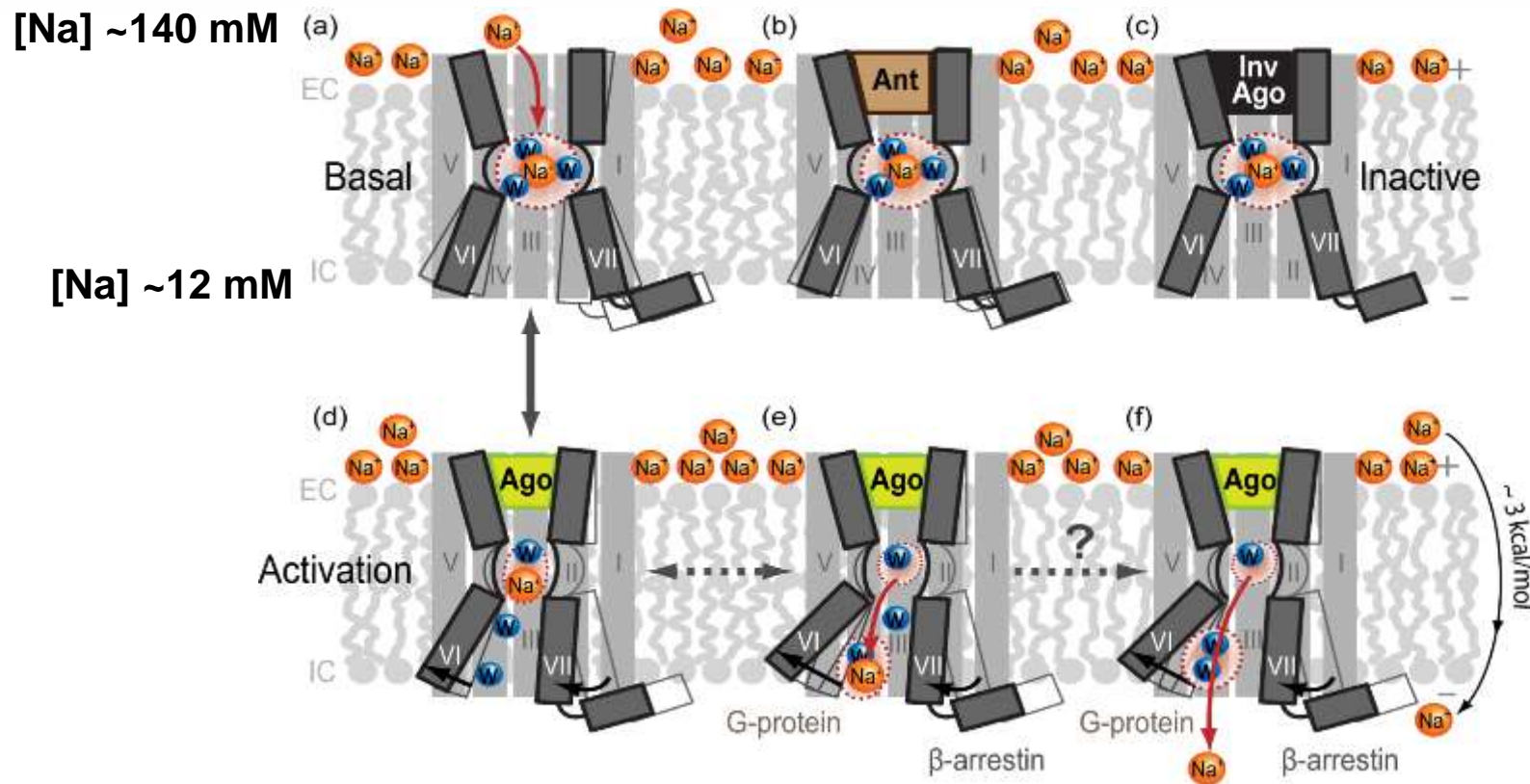
BlueOpsin

# Collapse of the Na<sup>+</sup> pocket in GPCR Activation

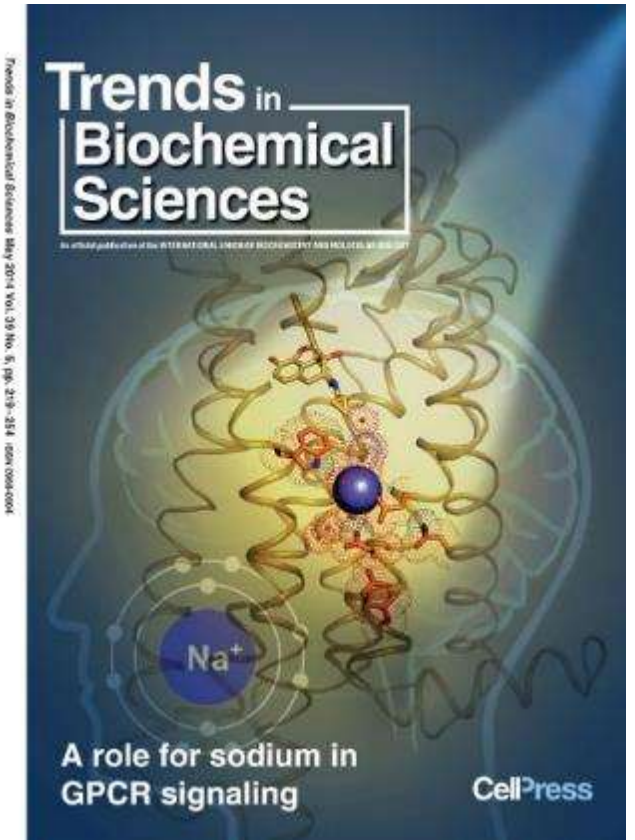




# Na<sup>+</sup> Plays a Key Role in Activation Mechanism

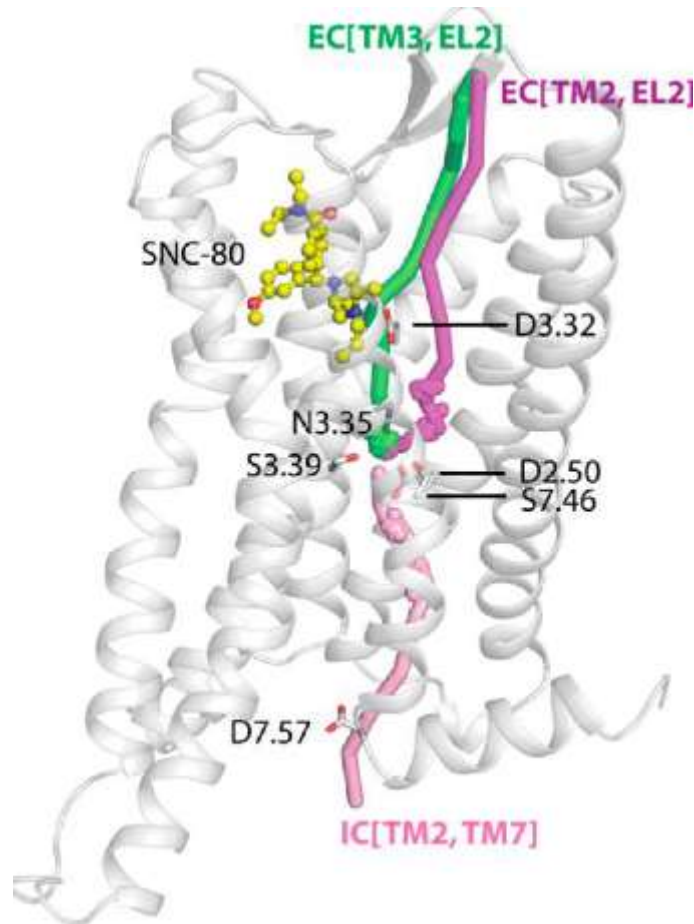


**Allosteric Sodium in GPCRs:  
One of the ten Science Signaling “Breakthroughs of 2014”**



Fenalti et al. 2014, Nature 506, 191-196  
Katritch et al. (2014) *TiBS* 39:233-44

# But where does it go upon activation?



- “In MD simulations of activation transition, Na<sup>+</sup> does not leave the allosteric pocket but rather kept its coordination with residues D2.50, N3.35, and S3.39”
- Used Accelerated MD, but only 15% follow the intracellular path

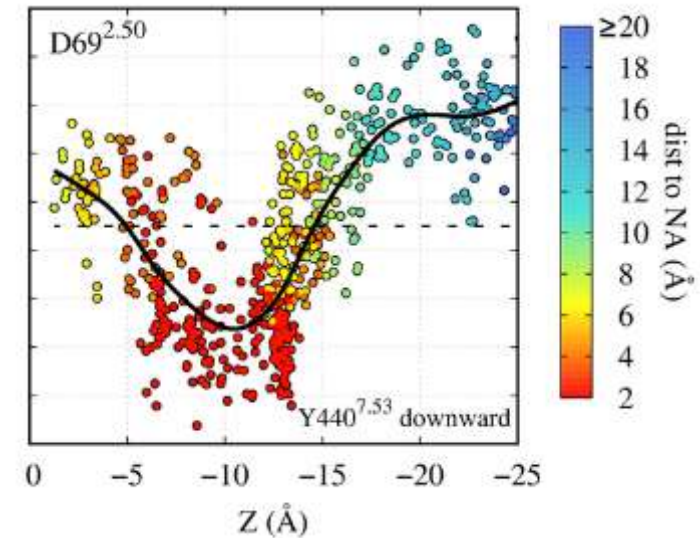
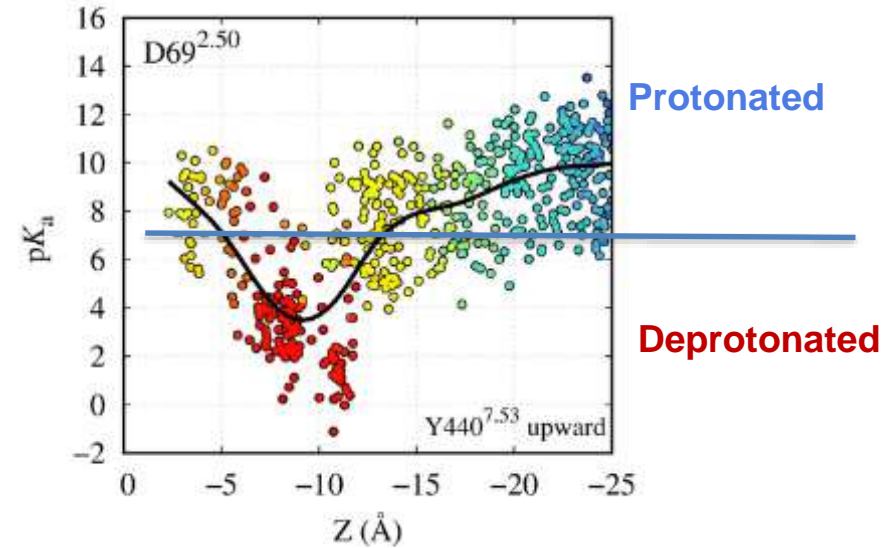
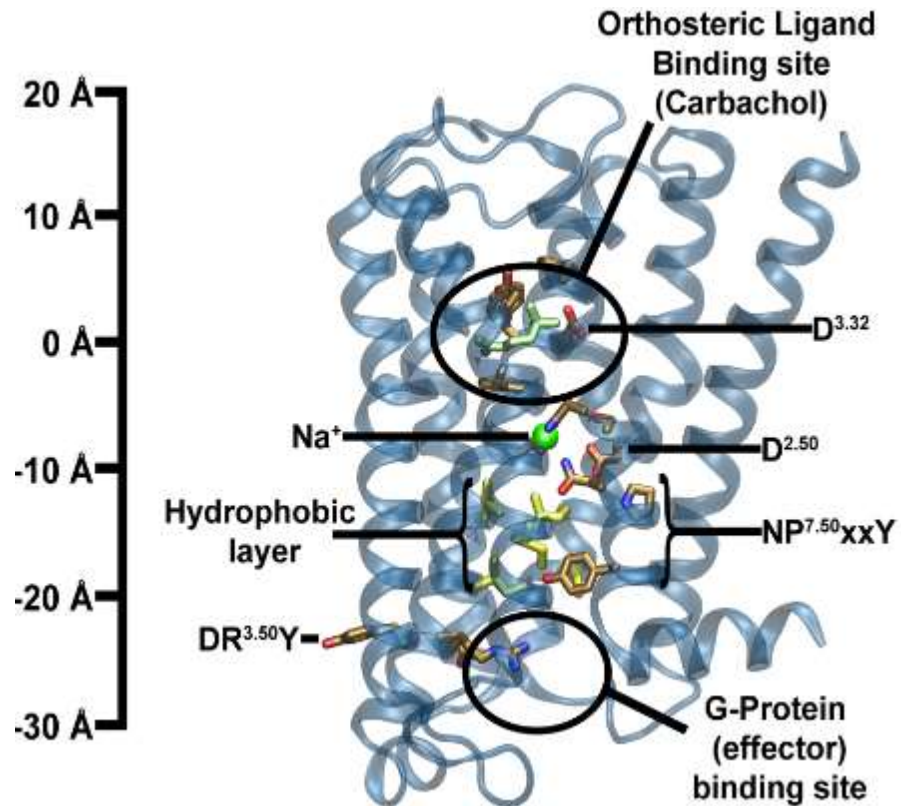
These MD simulations did not take into account:

1. Electrostatic and Na<sup>+</sup> concentration gradient
2. Possibility of D2.50 protonation

# Is protonation of D2.50 involved in Sodium transition?

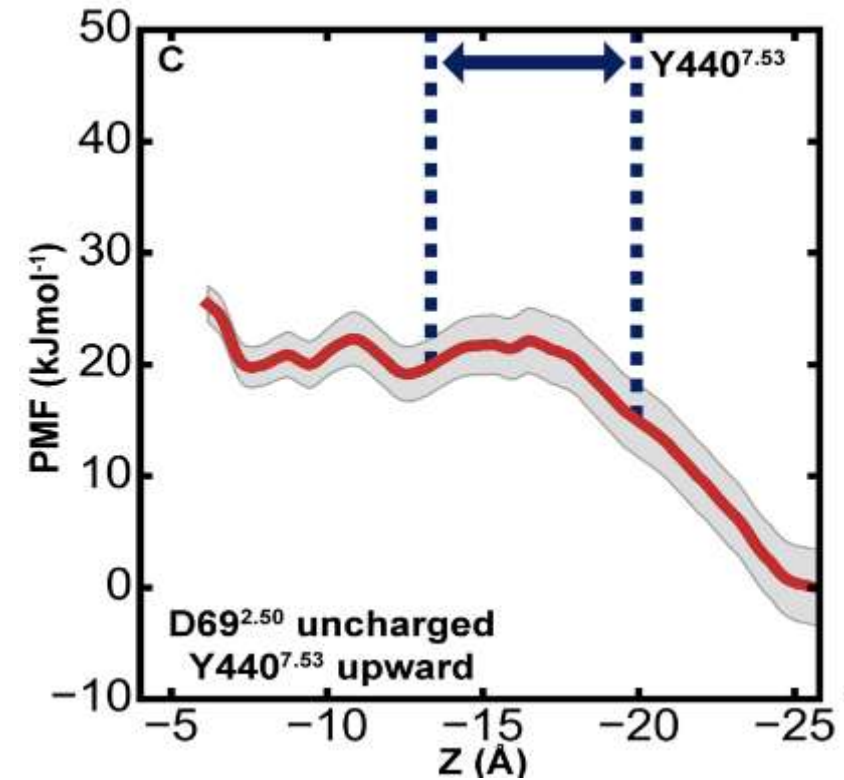
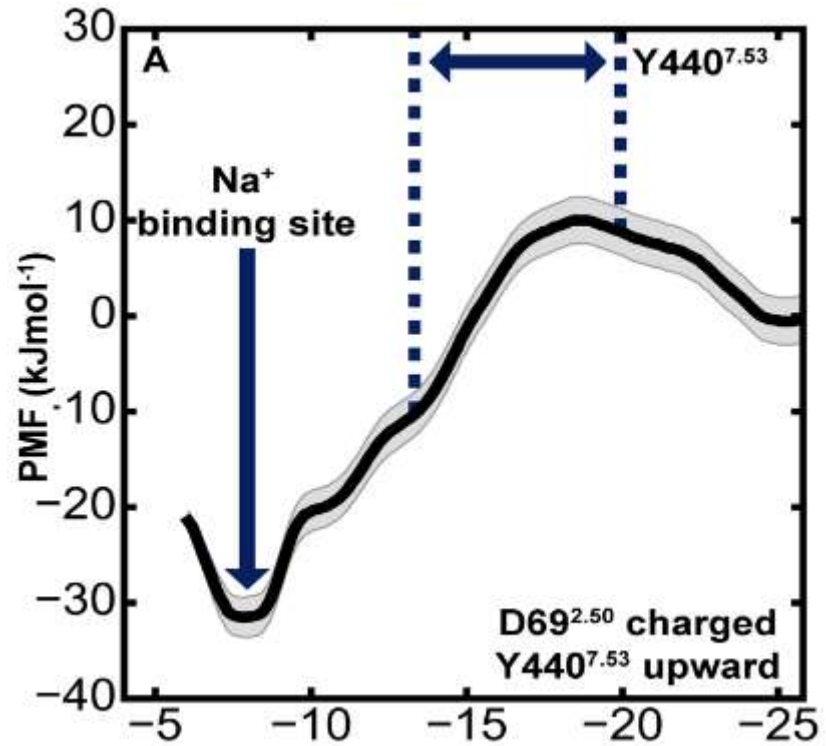


Calculated  $pK_a$  of D2.50 strongly depends on direct contact with  $\text{Na}^+$





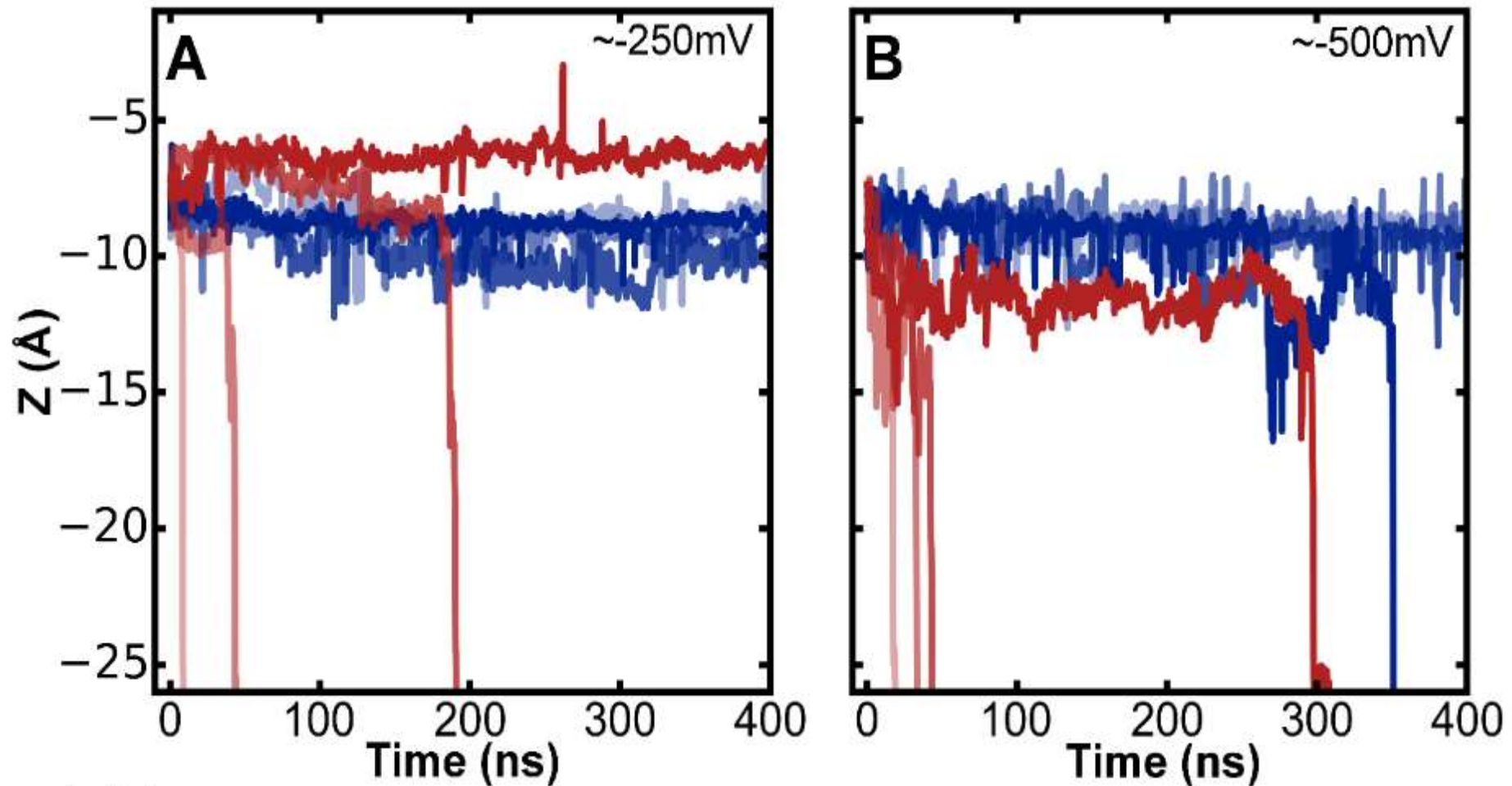
# Protonation Changes Na<sup>+</sup> Potential along the path



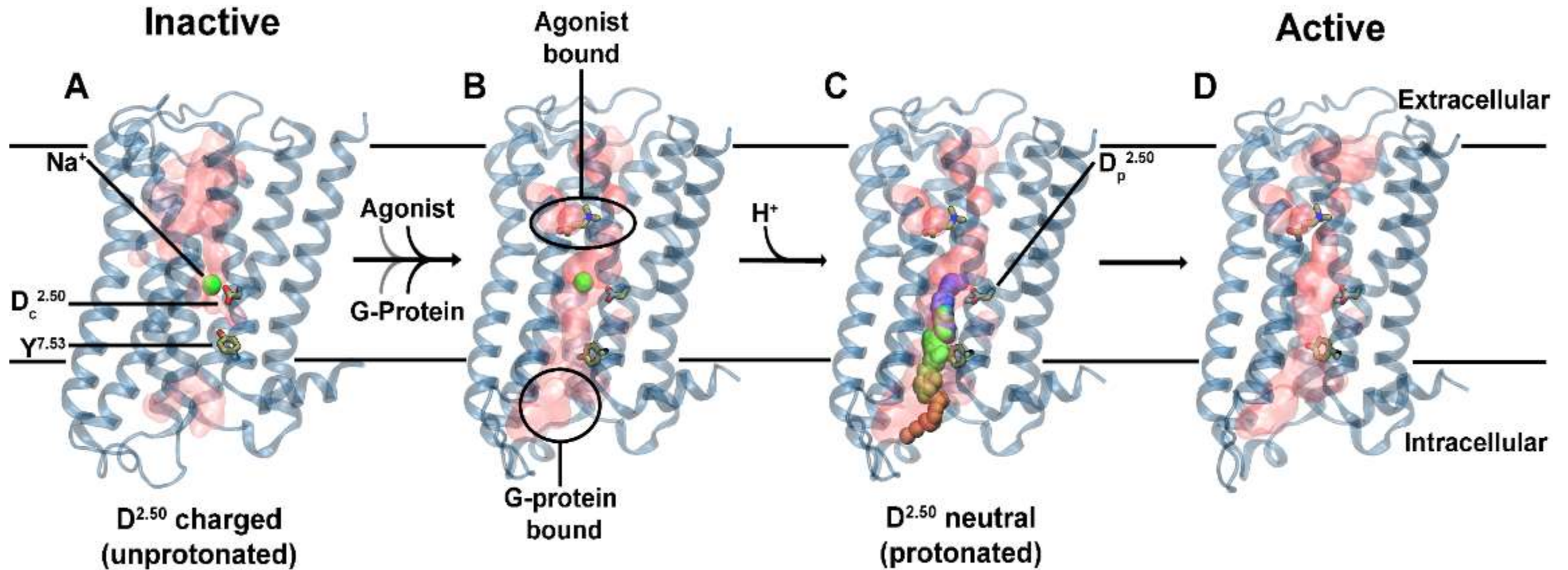
# Na<sup>+</sup> ion egress to the intracellular side.

D2.50 protonated

D2.50 charged



# Updated Na<sup>+</sup> mechanism





## Summary & Outlook : **Conserved Na<sup>+</sup> site in GPCRs**

- **Sodium site conserved in most class A GPCRs**
- **Sodium coordinating residues and Na<sup>+</sup> itself are involved in GPCR activation mechanism**
- **Sodium can travel through the GPCR “channel” along the voltage and concentration gradient (one Na<sup>+</sup> at a time!)**
- **Sodium transfer may be coupled with D2.50 protonation**
- **Sodium site can be exploited in ligand discovery and receptor stability design**

# Outline

- Rational prediction of stabilizing mutations:  
CompoMug
- New insights into GPCR function and allosteric mechanisms
- **Structure-Based ligand discovery for GPCRs**

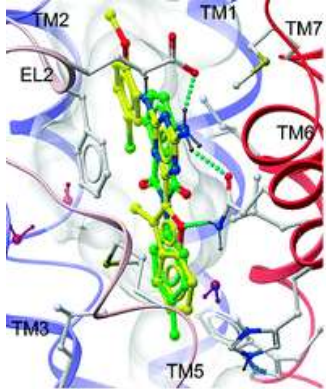
# Can we use structures in GPCR ligand discovery?

Journal of Medicinal Chemistry Article

J. Med. Chem. 2010, 53, 1799–1809 1799  
DOI: 10.1021/jm901647p

## Structure-Based Discovery of Novel Chemotypes for Adenosine A<sub>2A</sub> Receptor Antagonists

Vsevolod Katritch,<sup>\*,1,2,3</sup> Veli-Pekka Jaakola,<sup>1,4</sup> J. Robert Lane,<sup>1</sup> Judy Lin,<sup>1</sup> Adrian P. Hoerman,<sup>1</sup> Mark Yeager,<sup>2,4</sup> Inna Kufareva,<sup>1,5</sup> Raymond C. Stevens,<sup>\*,4</sup> and Ruben Abagyan<sup>\*,1,6</sup>



Journal of Medicinal Chemistry

## Optimization of Adenosine 5'-Carboxamide Adenosine Receptor Agonists Using Structure-Based Design and Fragment Screening

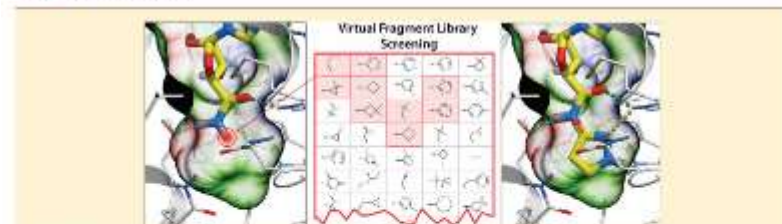
Dilip K. Tosh,<sup>†</sup> Khai Phan,<sup>†</sup> Zhan-Guo Gao,<sup>†</sup> Andrei A. Gakh,<sup>†</sup> Fei Xu,<sup>†</sup> Francesca Delloriano,<sup>†</sup> Ruben Abagyan,<sup>‡</sup> Raymond C. Stevens,<sup>‡</sup> Kenneth A. Jacobson,<sup>\*,§,¶</sup> and Vsevolod Katritch<sup>\*,§</sup>

<sup>†</sup>Molecular Recognition Section, Laboratory of Bioorganic Chemistry, National Institute of Diabetes and Digestive and Kidney Diseases, National Institutes of Health, Bethesda, Maryland 20892, United States

<sup>‡</sup>Department of Molecular Biology, The Scripps Research Institute, 10550 North Torrey Pines Road, La Jolla, California 92037, United States

<sup>§</sup>University of California, San Diego Skaggs School of Pharmacy and Pharmaceutical Sciences, 9500 Gilman Drive, La Jolla, California 92093, United States

Supporting Information



**ABSTRACT:** Structures of G protein-coupled receptors (GPCRs) have a proven utility in the discovery of new antagonists and inverse agonists modulating signaling of this important family of clinical targets. Applicability of active-state GPCR structures to virtual screening and rational optimization of agonists, however, remains to be assessed. In this study of adenosine 5' derivatives,

Journal of Medicinal Chemistry Article

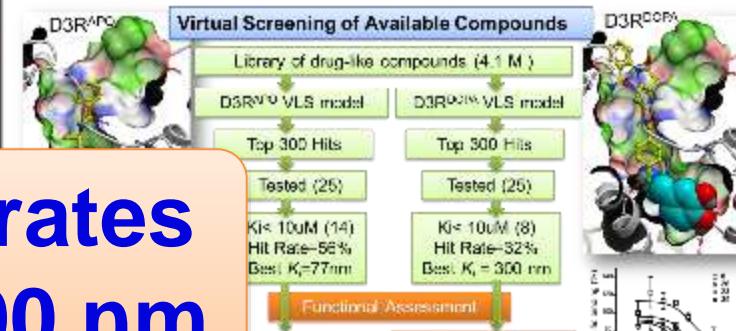
Journal of Medicinal Chemistry Article

## Structure-Based Ligand Discovery Targeting Orthosteric and Allosteric Pockets of Dopamine Receptors<sup>2</sup>

J. Robert Lane, Pavel Chubukov, Wei Liu, Meribell Canals, Vadim Cherezov, Ruben Abagyan, Raymond C. Stevens, and Vsevolod Katritch

Department of Integrative Structural and Computational Biology, Scripps Research Institute, La Jolla, California (P.C., W.L., V.C., R.C.S., V.K.); Drug Discovery Biology, Monash Institute of Pharmaceutical Sciences, Monash University, Parkville, Victoria, Australia (J.R.L., M.C.); and Skaggs School of Pharmacy and Pharmaceutical Sciences, and San Diego Supercomputer Center, University of California, San Diego, La Jolla, California (R.A.)

Received June 10, 2013; accepted September 10, 2013



30-70% hit rates  
Best  $K_i < 100 \text{ nm}$   
 $LE > 0.35$

## Exploring a 2-Naphthoic Acid Template for the Structure-Based Design of P2Y<sub>14</sub> Receptor Antagonist Molecular Probes

Evgeny Kiselev,<sup>†</sup> Matthew O. Barrett,<sup>‡</sup> Vsevolod Katritch,<sup>§</sup> Silvia Paoletta,<sup>†</sup> Clarissa D. Weitzer,<sup>†</sup> Kyle A. Brown,<sup>†</sup> Eva Hammes,<sup>†</sup> Andrew L. Yin,<sup>†</sup> Qiang Zhao,<sup>§</sup> Raymond C. Stevens,<sup>§</sup> T. Kendall Harden,<sup>†</sup> and Kenneth A. Jacobson<sup>\*,†</sup>

<sup>†</sup>Molecular Recognition Section, Laboratory of Bioorganic Chemistry, National Institute of Diabetes and Digestive and Kidney Diseases, National Institutes of Health, Bethesda, Maryland 20892, United States

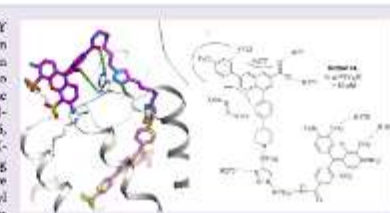
<sup>‡</sup>Department of Pharmacology, University of North Carolina, School of Medicine, Chapel Hill, North Carolina 27599, United States

<sup>§</sup>CAS Key Laboratory of Receptor Research, Shanghai Institute of Materia Medica, Chinese Academy of Sciences, 555 Zuchongzhi Road, Pudong, Shanghai 201203, China

<sup>¶</sup>Department of Integrative Structural and Computational Biology, The Scripps Research Institute, 10550 North Torrey Pines Road, La Jolla, California 92037, United States

Supporting Information

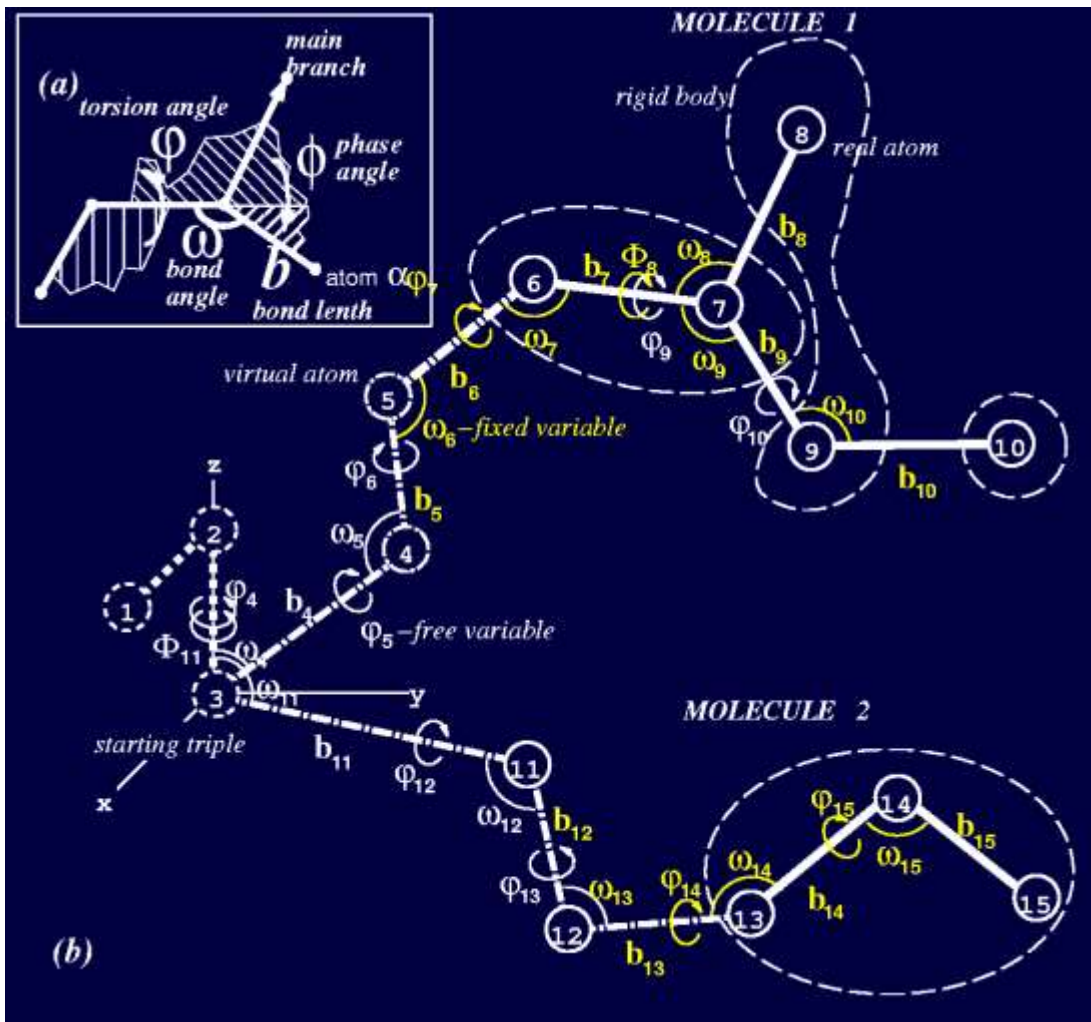
**ABSTRACT:** The P2Y<sub>14</sub> receptor (P2Y<sub>14</sub>R), one of eight P2Y G protein-coupled receptors (GPCR), is involved in inflammatory, endocrine, and lysocaine processes and is an attractive pharmaceutical target. The goal of this research is to develop high-affinity P2Y<sub>14</sub>R fluorescent probes based on the potent and highly selective antagonist 4-(4-(piperidin-4-yl)-phenyl)-7-(4-(trifluoromethyl)-phenoxy)-2-naphthoic acid (6, 997N). A model of hP2Y<sub>14</sub>R based on recent hP2Y<sub>12</sub>R X-ray structures together with simulated antagonism docking suggested that the piperidine ring is suitable for fluorophore conjugation while preserving affinity. Chain-elongated allyl or amino derivatives of 6 for click or amide coupling were





# Core Modeling Technology: Internal Coordinate Mechanics (ICM)

$$E(\alpha) = \Delta E_{FF} + \Delta E_{EN} + \alpha_1 N_{at} + \alpha_2 \Delta E_{HB} + \alpha_3 \Delta E_{SE} + \alpha_4 \Delta E_{EL} + \alpha_5 \Delta E_{SO}$$



Internal coordinates:

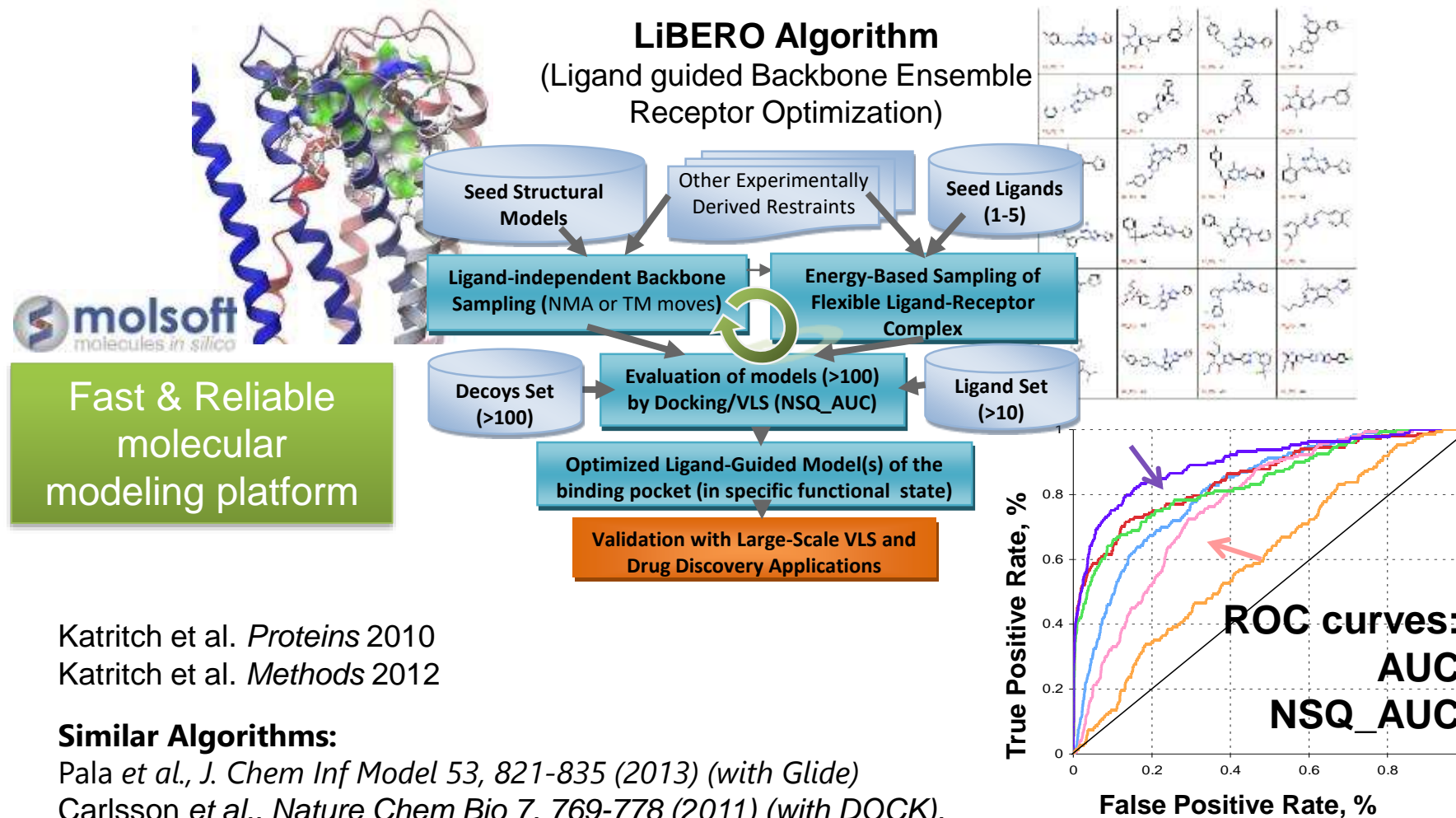
- + Eliminates “high frequency” modes
- + Efficient & fast global energy optimization algorithm
- + Large radius of convergence
- + Accurate force field and free energy func (solvation/entropy)
- + **Easy to perform sampling of both ligand and receptor conformations**

## ICM References:

- Abagyan et al. (1994)  
“ICM - a new method for protein modeling..”  
*J. Comp. Chem.* 15, 488-506
- Katritch et al (2003).  
ICFF: New Internal Coordinate Force Field  
*J Comp. Chem.* 24:254-65

# Ligand-Guided Model Optimization

- Improve accuracy and reliability of docking
- Improve VLS performance, especially for lower resolution structures or homology models
- Reduce ligand bias, or shift bias towards desirable ligand/scaffold
- Develop VLS models for different functional states



Katritch et al. *Proteins* 2010  
Katritch et al. *Methods* 2012

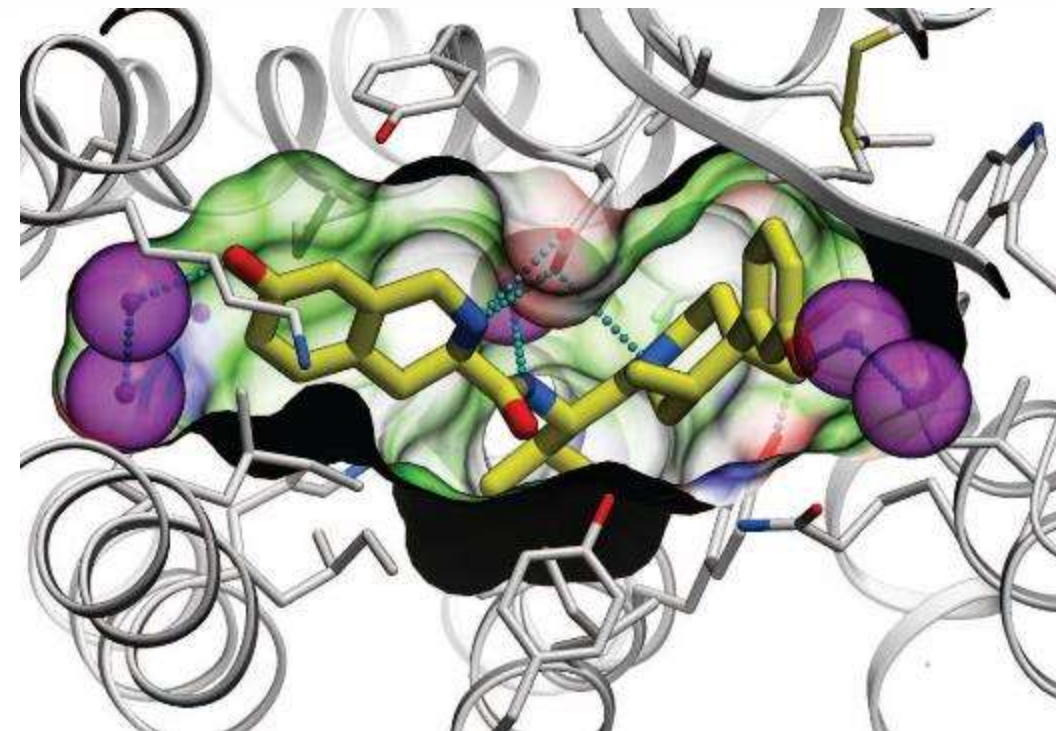
## Similar Algorithms:

Pala et al., *J. Chem Inf Model* 53, 821-835 (2013) (with Glide)  
Carlsson et al., *Nature Chem Bio* 7, 769-778 (2011) (with DOCK).

# $\kappa$ -Opioid Receptor as a Therapeutic Target

- $\kappa$ -OR full agonists (e.g. SalA) lead to hallucination and dysphoria
- $\kappa$ -OR antagonists: potential antidepressants, anxiolytics and anti-addiction drugs
- JDTic: high affinity selective antagonist, (but cardiac side effects in clinic)
- G-protein biased  $\kappa$ -OR agonists (no  $\beta$ -arrestin signaling): non-addictive analgesia
- **New  $\kappa$ -OR ligand chemotypes needed for both antagonist and biased agonist functional profiles**

Buda et al Neuropsychopharmacology 2015.  
Carroll and Carlezon (2015) JMC 56: 2178–2195.



$\kappa$ -OR structure with JDTic (4DJH)  
Wu et al 2012, Nature

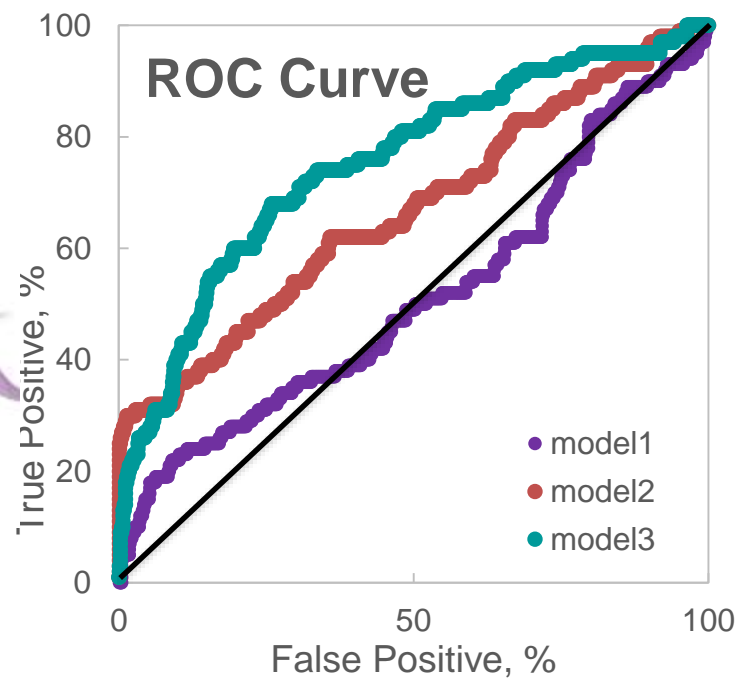
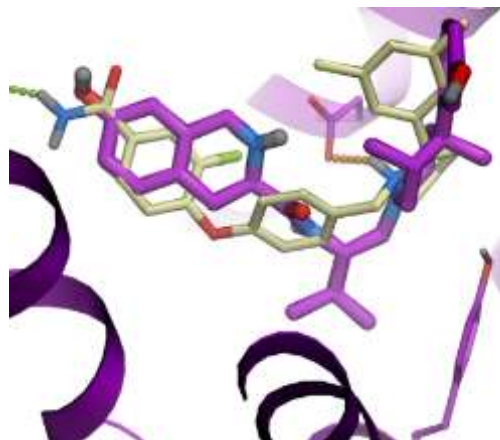
**Hard target for VLS – few previously published VLS hits all  $>\mu\text{M}$  range**



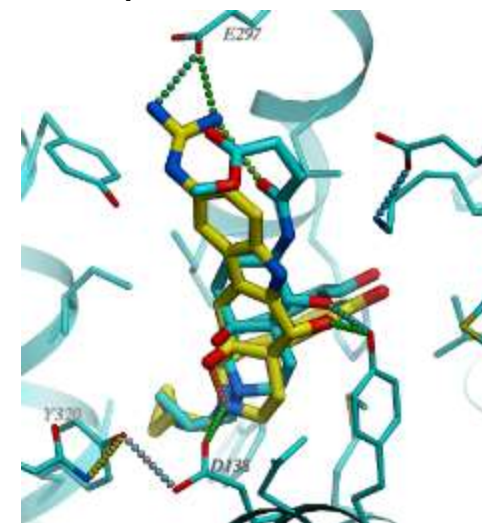
# Virtual Ligand Screening for $\kappa$ -OR: Model Optimization



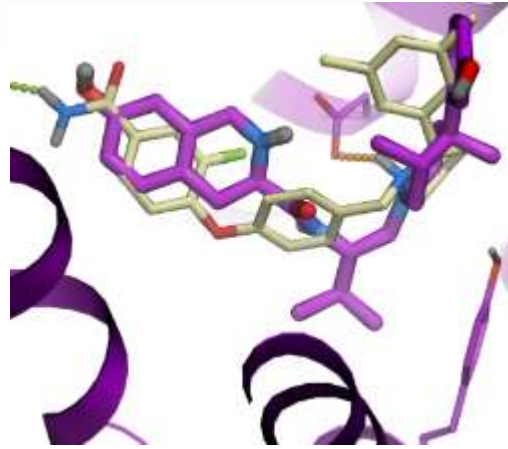
**Model 1**  
JDTic-bound



**Model 3**  
Morphinan-bound



# Virtual Ligand Screening for $\kappa$ -OR New Chemotype Discovery



Three Ligand-Optimized VLS models

Virtual Fragment-like and Lead-like Library  
(5M compounds)

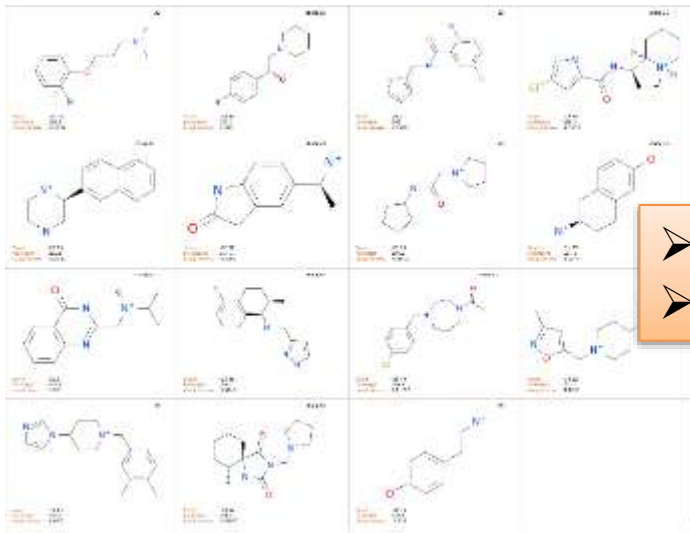
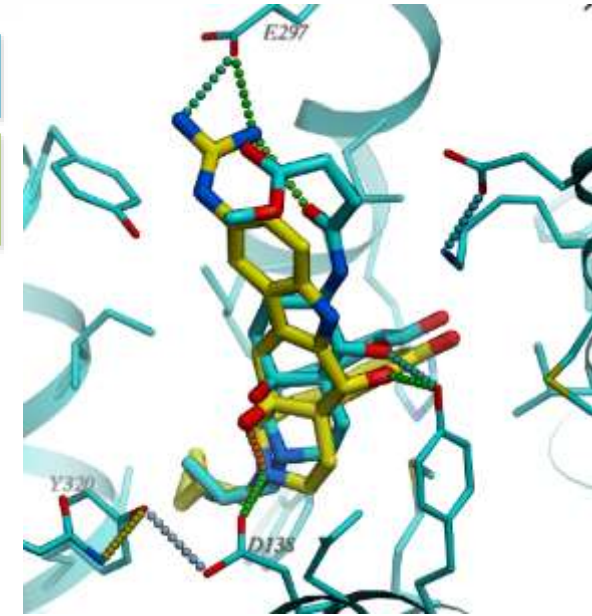
Novel Candidate hits with  $\Delta\Delta G < 30$  (~200)

Ordered and tested (43)

$K_i(\kappa\text{-OR}) < 30\mu\text{M}$  (27)

$K_i(\kappa\text{-OR}) < 10\mu\text{M}$  (14)

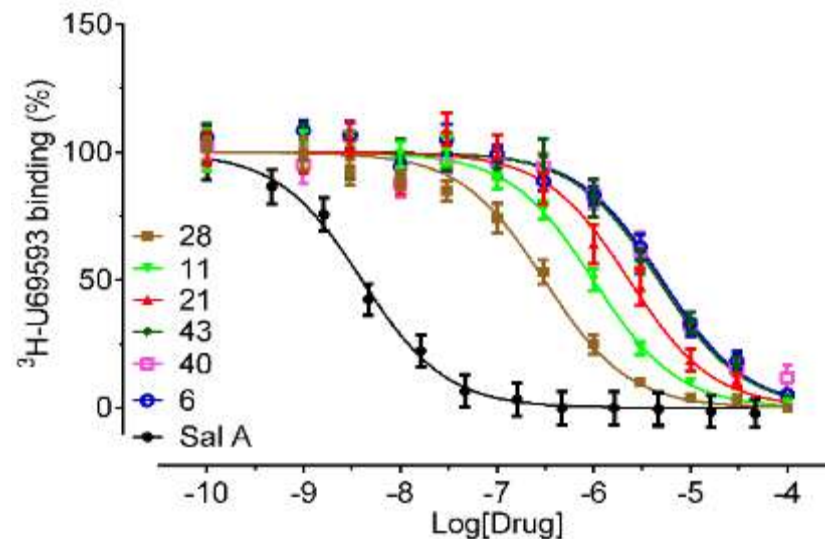
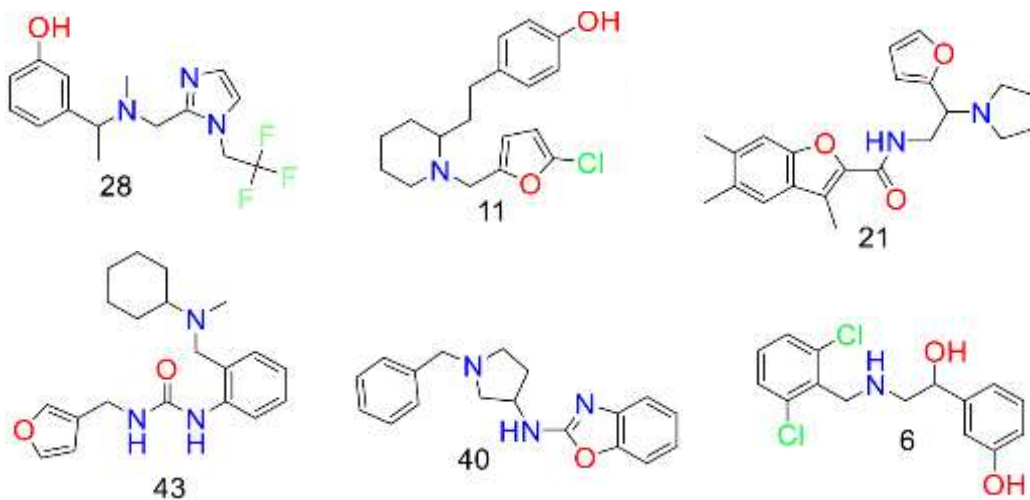
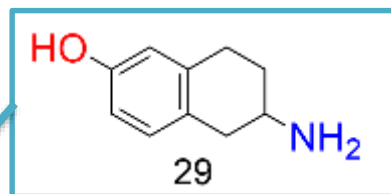
- Best  $K_i < 200$  nm, Ligand Efficiency **LE**  $> 0.45$
- Agonists and antagonists, best  $EC_{50} = 260$  nm



# > 6 New Distinct Chemotypes of $\kappa$ -OR Ligands

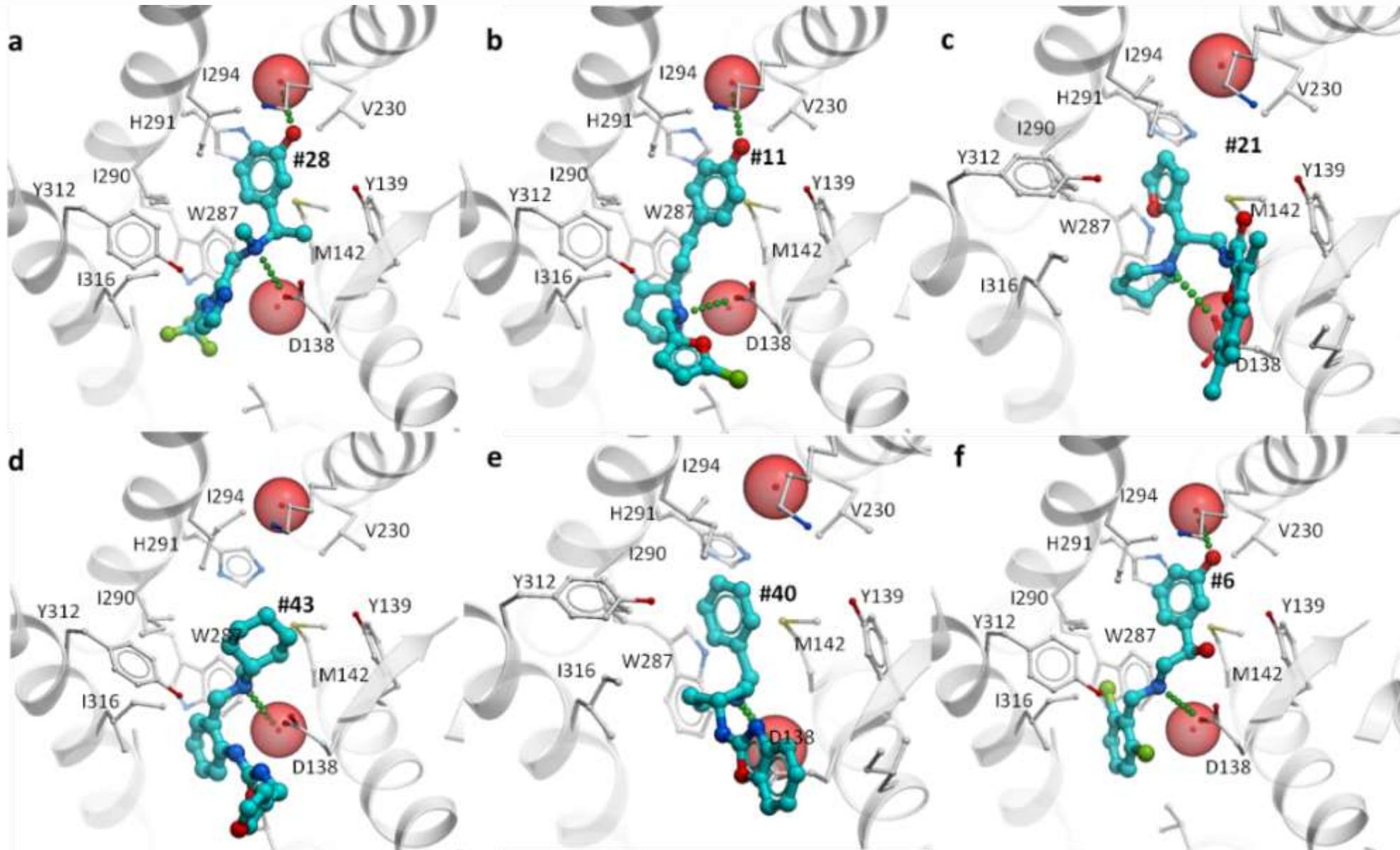
Comp#	Model <sup>a</sup>	Inhibit% <sup>b</sup>	$\pm$	SEM <sup>c</sup>	$pK_i$	$\pm$	SEM	$K_i(\mu\text{M})$	LE <sup>d</sup>	Tanimoto <sup>e</sup>
28	1	96.9	$\pm$	1.2	6.82	$\pm$	0.16	0.2	0.43	0.42
11	1	95.3	$\pm$	2.5	6.30	$\pm$	0.12	0.5	0.40	0.19
21	3	77.5	$\pm$	6.8	5.95	$\pm$	0.26	1.1	0.31	0.29
43	2	76.5	$\pm$	4.7	5.64	$\pm$	0.16	2.3	0.32	0.39
40	3	77.5	$\pm$	5.6	5.59	$\pm$	0.22	2.6	0.36	0.40
6	1	80.1	$\pm$	4.2	5.58	$\pm$	0.17	2.6	0.39	0.39
17	2	53.0	$\pm$	7.8	5.52	$\pm$	0.12	3.0	0.32	0.52
16	2	60.7	$\pm$	14.7	5.52	$\pm$	0.17	3.0	0.37	0.38
20	2	49.1	$\pm$	16.4	5.21	$\pm$	0.14	6.2	0.30	0.42
35	2	49.8	$\pm$	8.4	5.19	$\pm$	0.17	6.4	0.36	0.51
3	1	70.2	$\pm$	3.9	5.18	$\pm$	0.18	6.6	0.36	0.26
8	1	73.7	$\pm$	4.0	5.09	$\pm$	0.22	8.1	0.29	0.34
29	2	48.6	$\pm$	9.6	5.08	$\pm$	0.12	8.3	0.59	0.15
22	2	47.3	$\pm$	7.7	5.01	$\pm$	0.13	9.8	0.28	0.39

“Similar” to JD<sub>Tic</sub> core  
tetrahydronaphthalene vs.  
tetrahydroisoquinoline



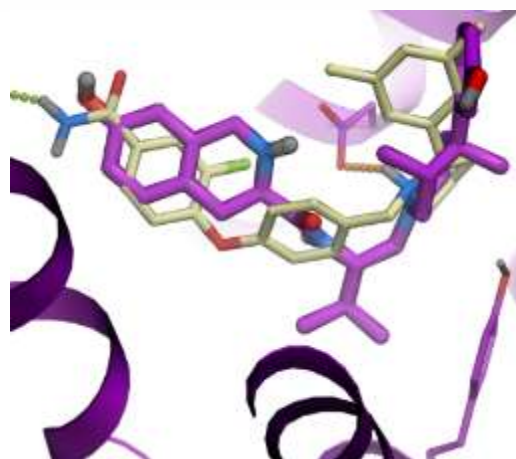


# > 6 New Distinct Chemotypes of $\kappa$ -OR Ligands



# Virtual Ligand Screening for $\kappa$ -OR

## First Round of Optimization



Three Ligand-Optimized VLS models

Virtual Fragment-like and Lead-like Library  
(5M compounds)

Novel Candidate hits with  $\Delta\Delta G < 30$  (~200)

Ordered and tested (43)

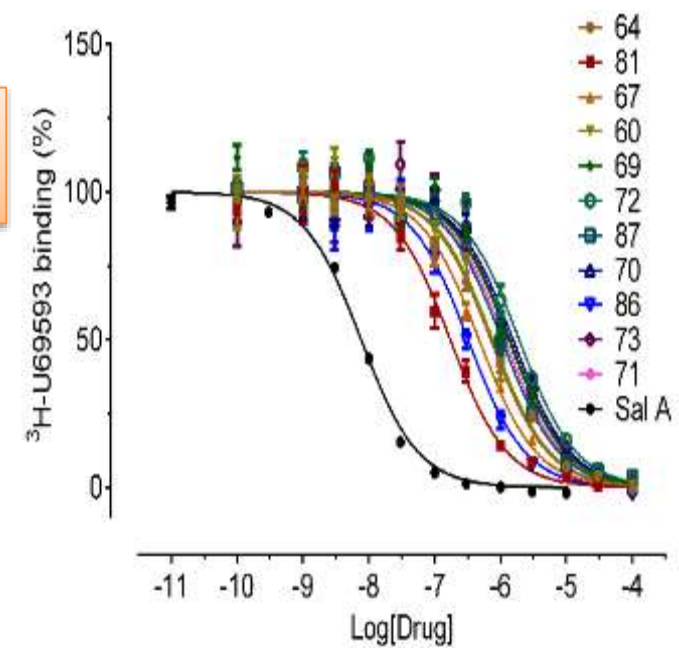
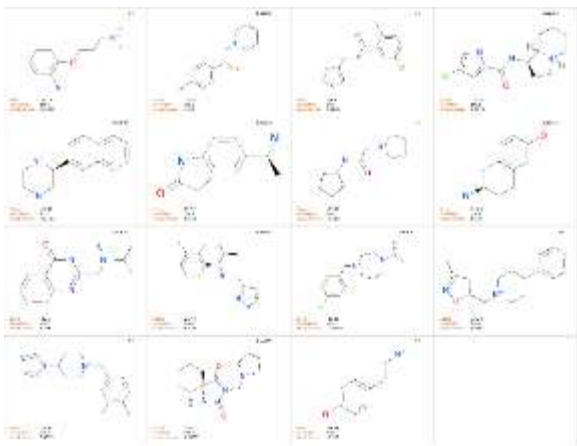
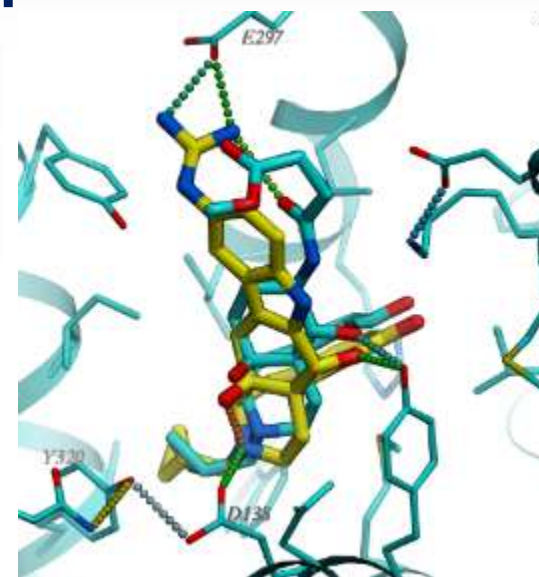
$K_i(\kappa\text{-OR}) < 30\mu\text{M}$  (27)

$K_i(\kappa\text{-OR}) < 10\mu\text{M}$  (14)

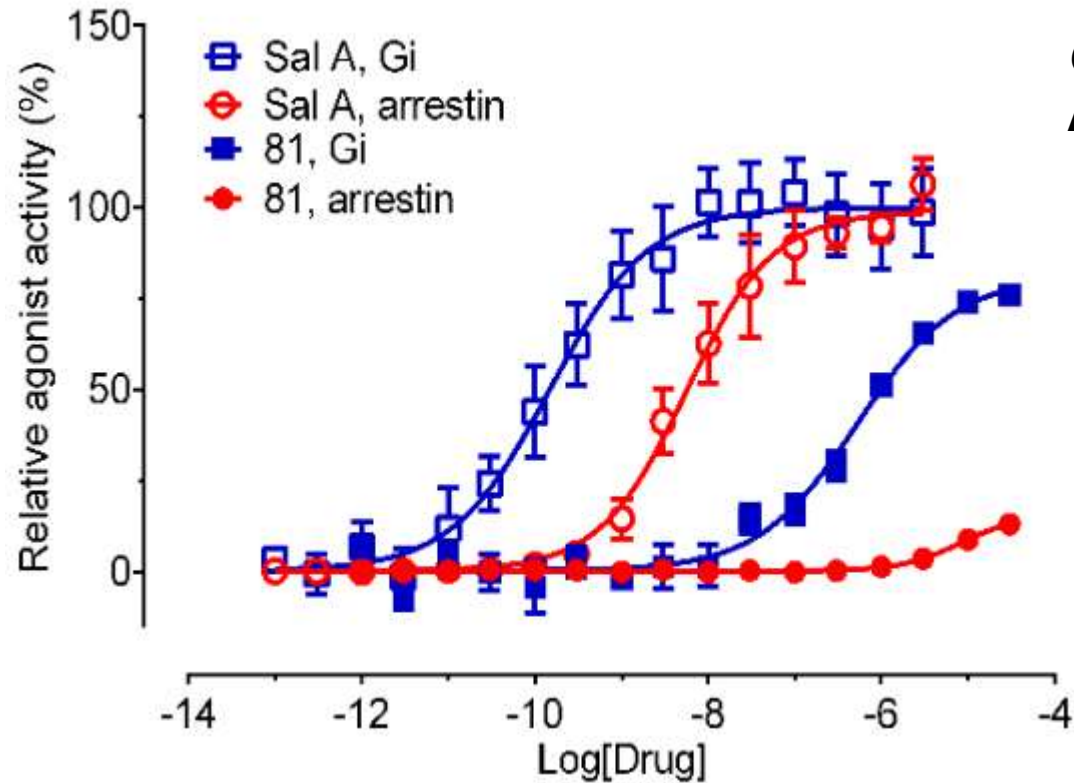
- Best  $K_i < 200$  nm, Ligand Efficiency **LE > 0.45**
- Agonists and antagonists, best  $EC_{50} = 260$  nm

**Analog by catalog for the top leads**

- 30 compounds tested (in 6 scaffolds)
- **10** sub-micromolar compounds!
- **Best  $K_i < 90$  nm!**
- **Functional Profiles >**



# Discovery of a new Biased Agonist



Comp #81:

Binding Affinity  $K_i = 160$  nM

$G_i$  pathway:  $EC_{50} = 530$  nM

Arrestin :  $EC_{50} > 10000$  nM

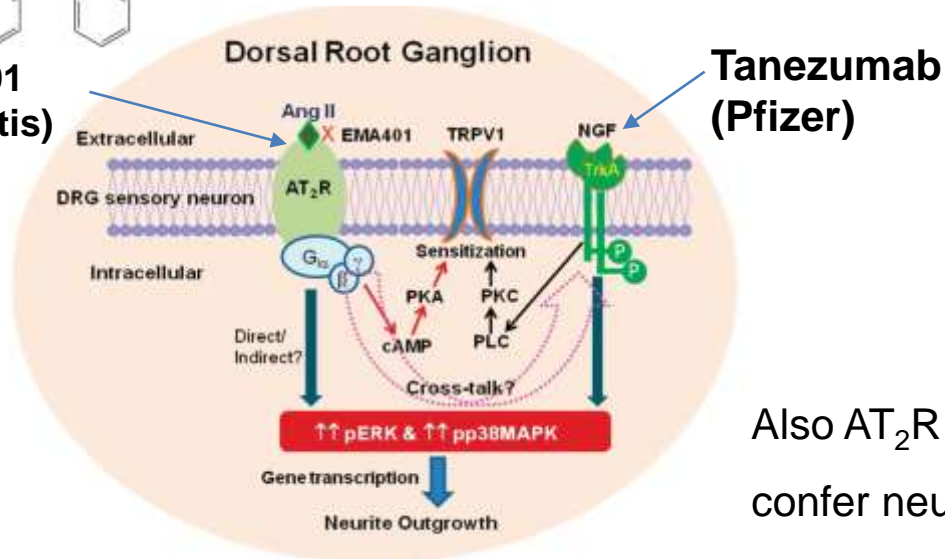
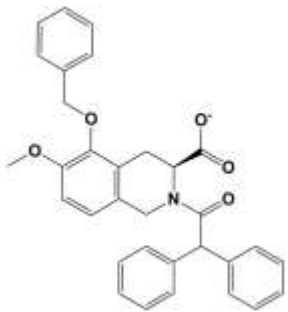
Bias factor:  $\Delta\Delta\log(\tau/K_A) > 6.0$   
( $G_i$  vs arrestin pathways)

Ongoing optimizations of best scaffolds

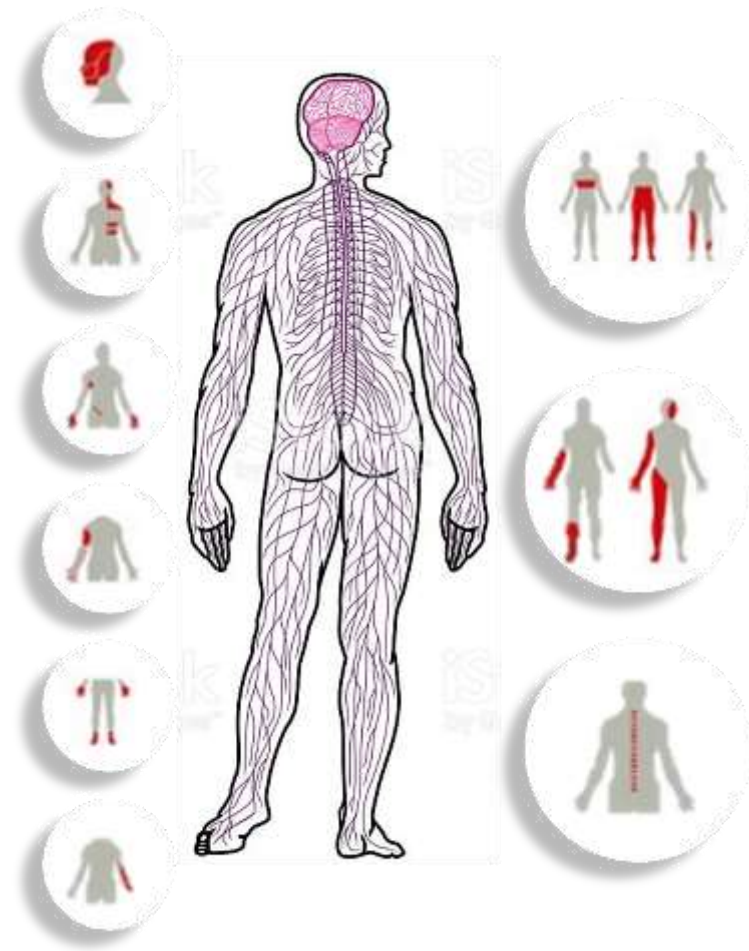
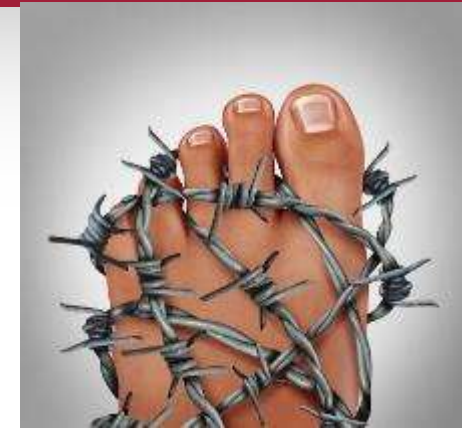


# Angiotensin AT<sub>2</sub>R antagonists for neuropathic pain relief

- AT<sub>2</sub>R regulates nerve sensitization and neurite outgrowth in neuropathic pain
- Blocking AT<sub>2</sub>R in PNS can reduce the pain
- **EMA401** is in Phase II trials for post-herpetic neuropathic pain (Novartis)



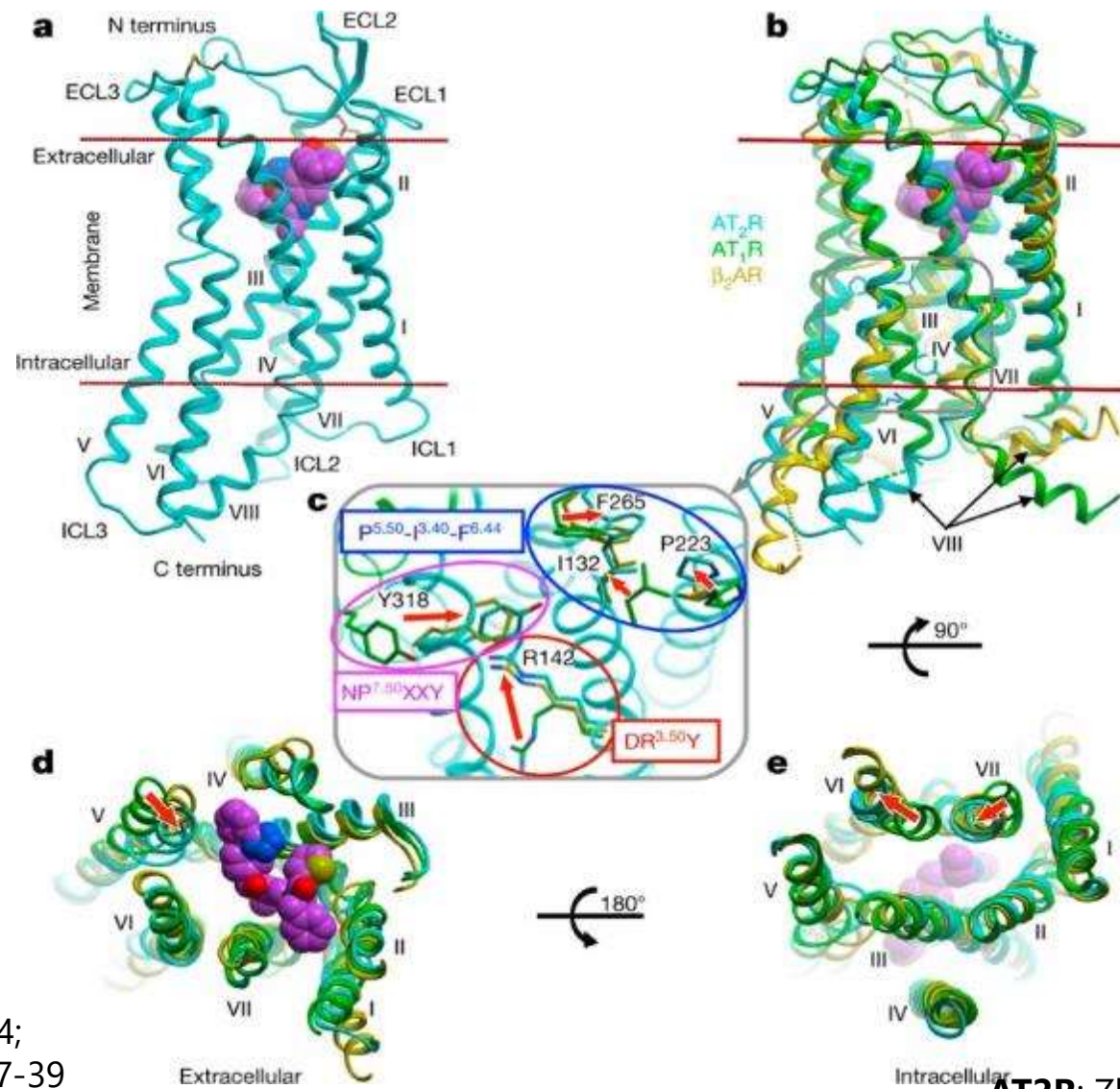
Also AT<sub>2</sub>R agonists (C21) confer neuroprotection in CNS (stroke)



Smith et al, *Pain* 2016



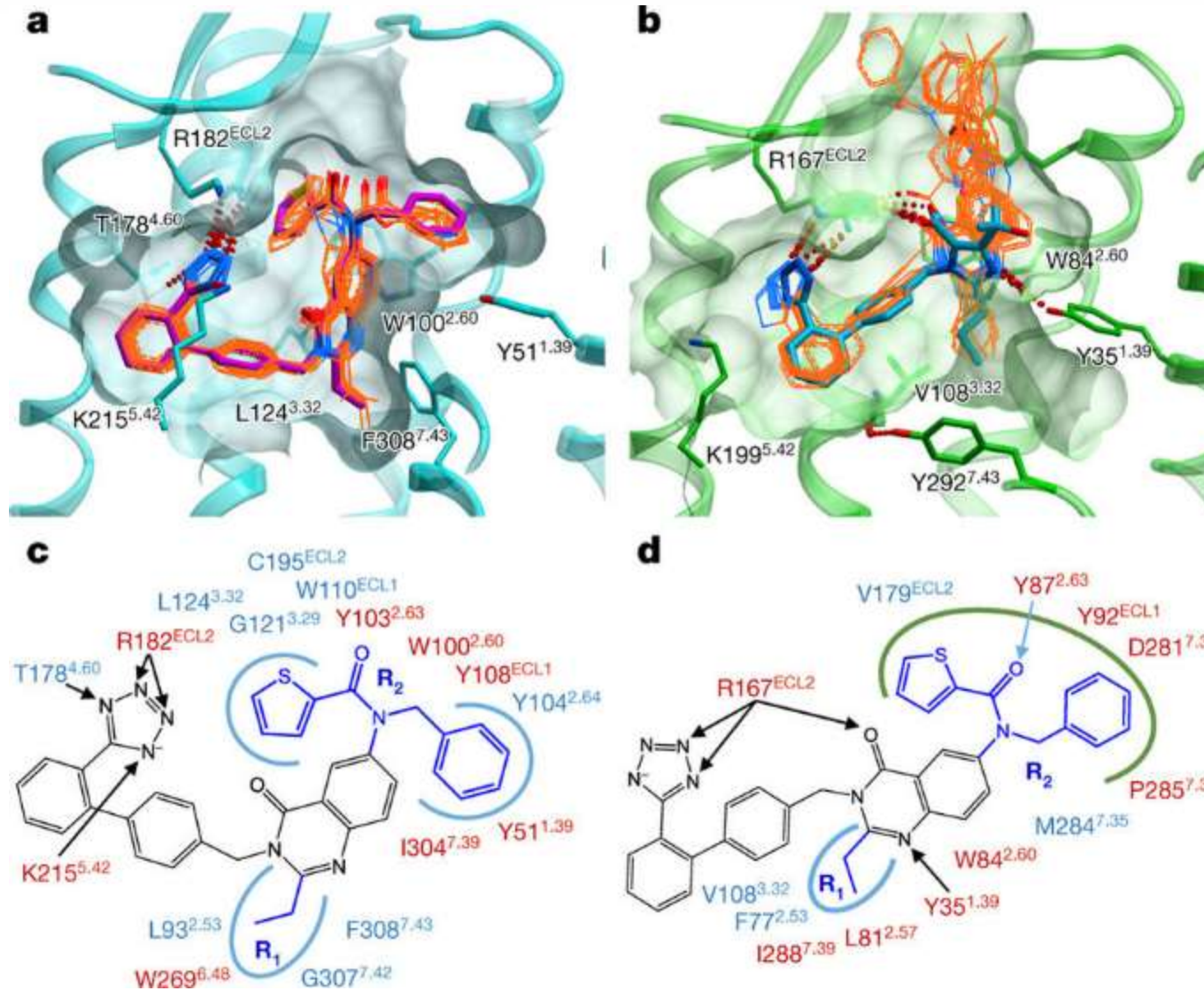
# Crystal Structures of AT<sub>1</sub>R and AT<sub>2</sub>R give new insights into function ...



**AT<sub>1</sub>R:** Zhang H, *Cell*. 2015;161(4):833-44;  
Zhang H, *JBC* 2015;290(49):29127-39

**AT<sub>2</sub>R:** Zhang H, *Nature*. 2017;544(7650):327-32;

# ... and AT<sub>1</sub>R vs. AT<sub>2</sub>R selectivity



AT<sub>2</sub>R: Zhang H, *Nature*. 2017;544(7650):327-32;



# Lead-like AT<sub>2</sub>R selective hits from initial VLS

Model tested and refined with EMA401

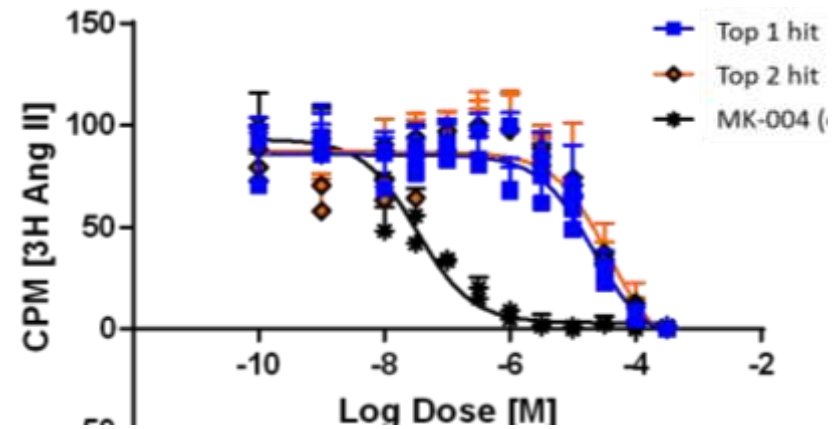
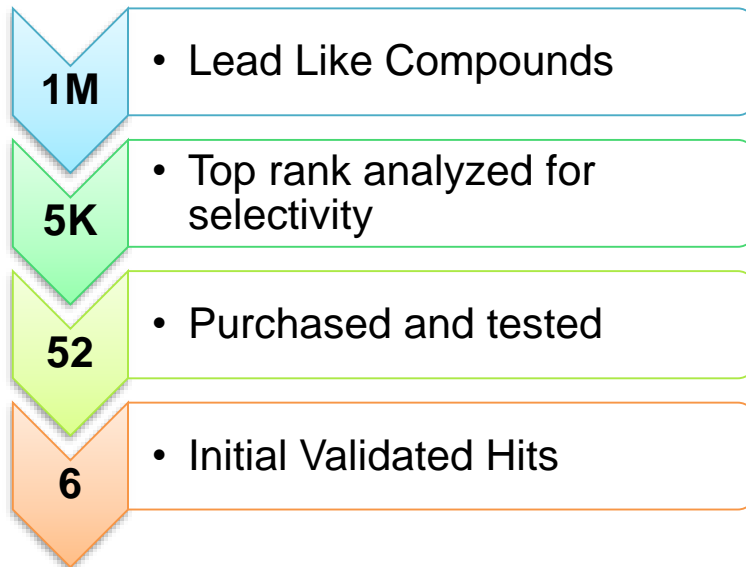
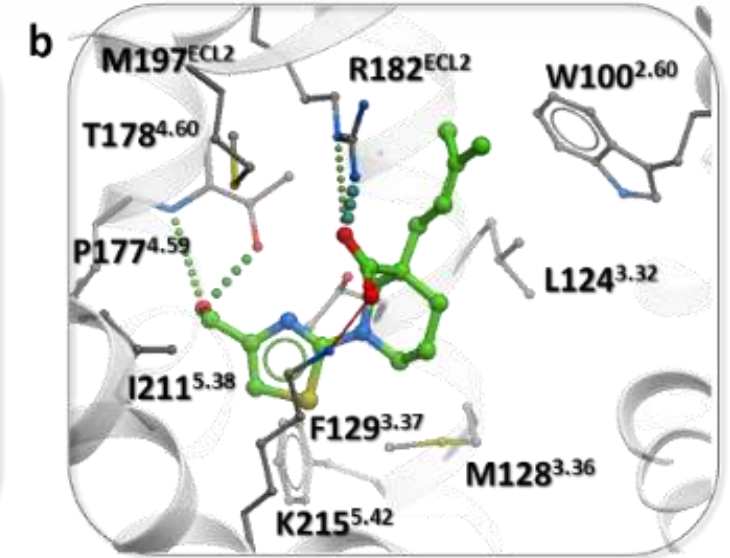
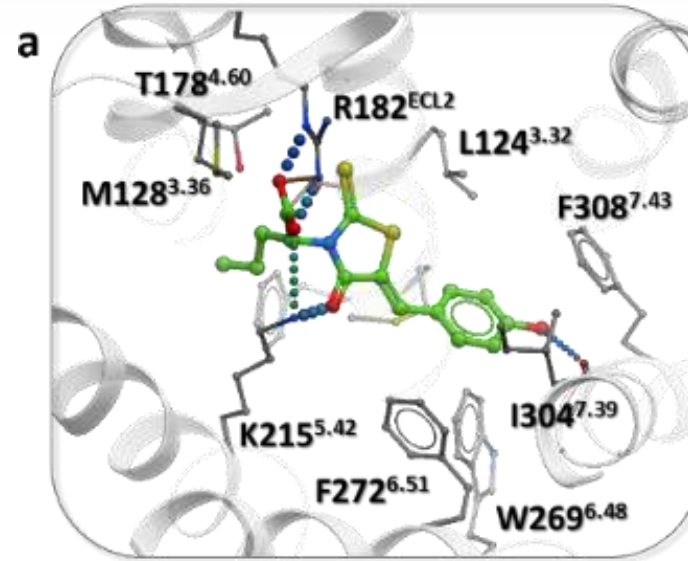
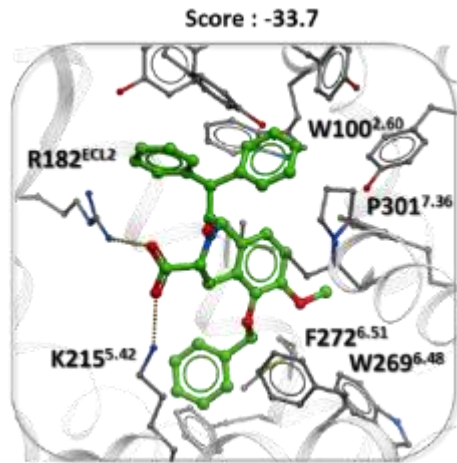
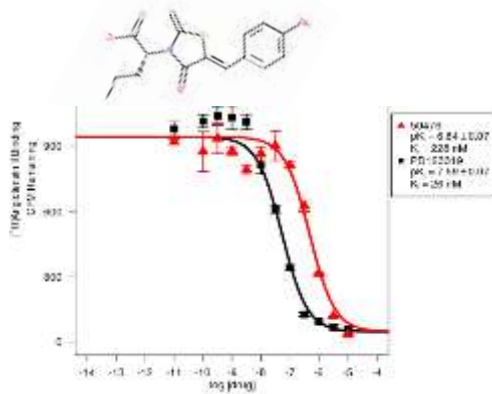


Fig.10 Radioligand competition binding assay for the MK-4 and top two hit compounds.

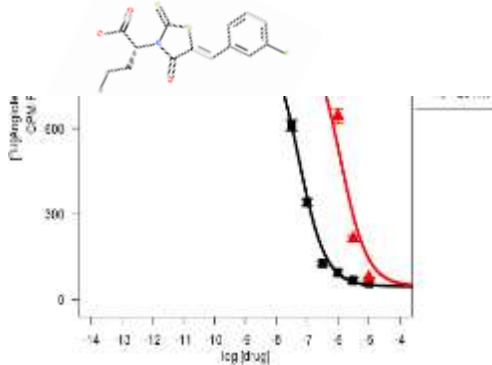
# New SAR Results

Confidential

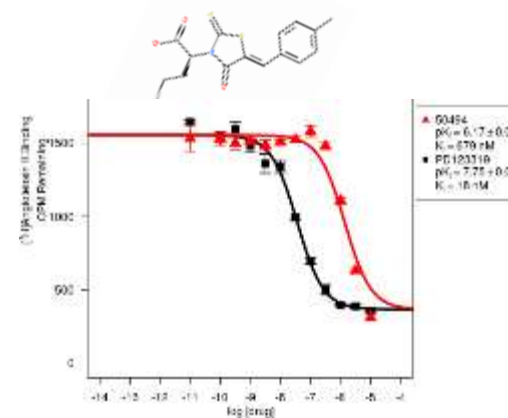
**BRI-6001**



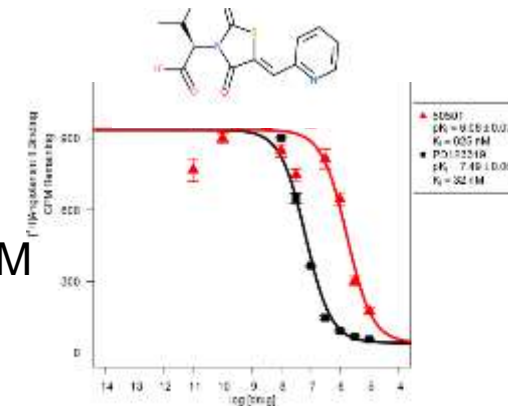
**BRI-6003**



**BRI-6011**



**BRI-6018**

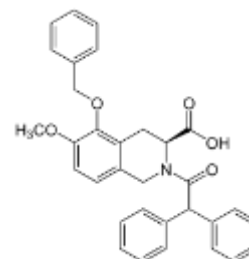


Compound # (Name)	$pK_i \pm SEM$	$K_i$ , nM	LE
<b>50476 (BRI-6001)</b>	<b>6.64 ± 0.07</b>	<b>228</b>	<b>0.42</b>
<b>50486 (BRI-6103)</b>	<b>6.24 ± 0.07</b>	<b>581</b>	<b>0.40</b>
50491 (BRI-6108)	5.65 ± 0.07	2254	0.36
50492 (BRI-6109)	5.43 ± 0.08	3739	0.35
<b>50494 (BRI-6111)</b>	<b>6.17 ± 0.06</b>	<b>680</b>	<b>0.39</b>
<b>50501 (BRI-6118)</b>	<b>6.08 ± 0.07</b>	<b>825</b>	<b>0.41</b>
50502 (BRI-6119)	5.25 ± 0.09	5681	0.32
50477 (BRI-6002)	5.67 ± 0.08	2158	0.38
<b>50533 (EMA401)</b>	<b>7.60 ± 0.04</b>	<b>25</b>	<b>0.28</b>
50534 (C21)	7.44 ± 0.04	36	0.33

Ligand Efficiency,  $LE = 1.4 * pK_i / N_{atom}$

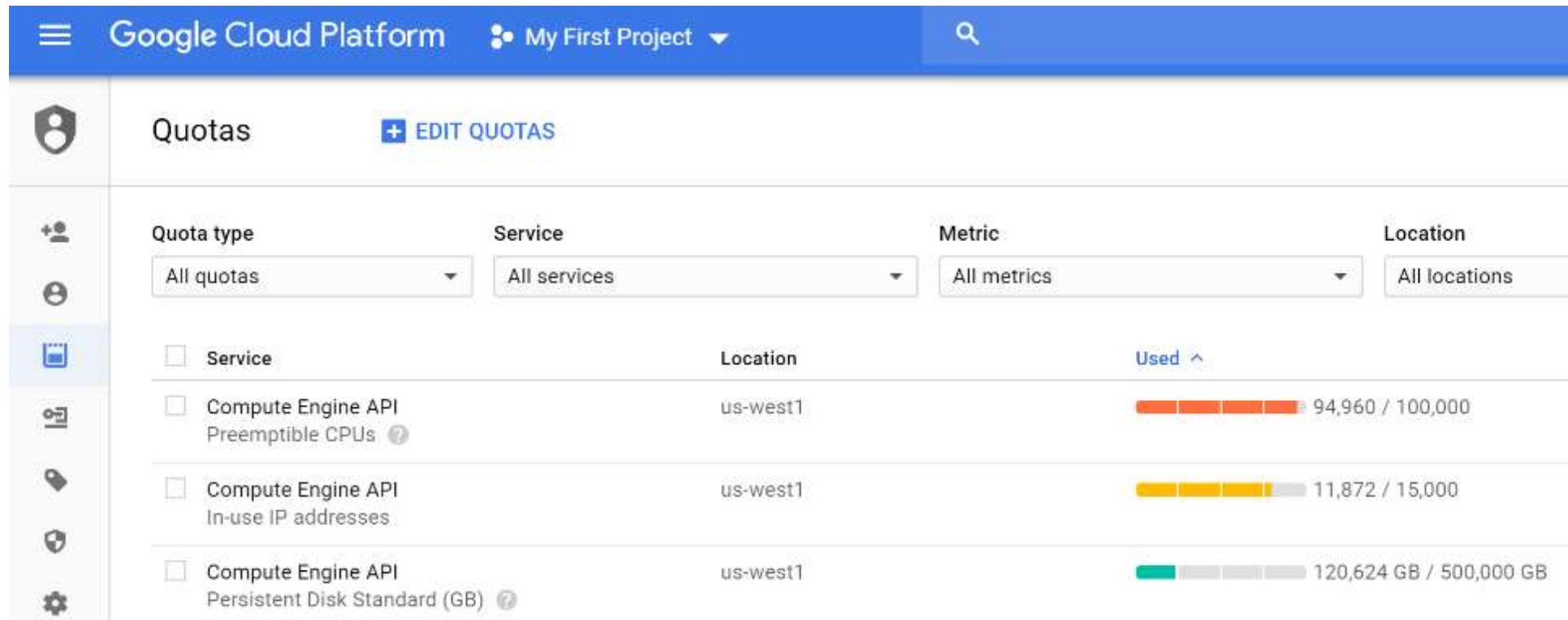
- Four sub-micromolar hits
- Very high Ligand Efficiency (LE)
- AT<sub>2</sub>R selective: no AT<sub>1</sub>R binding at 30 μM
- Amenable for lead optimization

Control: PD123319 (**EMA401**)  
 $N_{atom} = 38$






# Google Cloud screening of 500M compounds (NIH Commons Pilot Project)

- Used new ZINC library of “Wait-OK” compounds (4-6 weeks, 80% success)
- Google Cloud screening used ~100,000 CPUs at once
- Completed in 24 hours
- Would take ~5 month on our servers



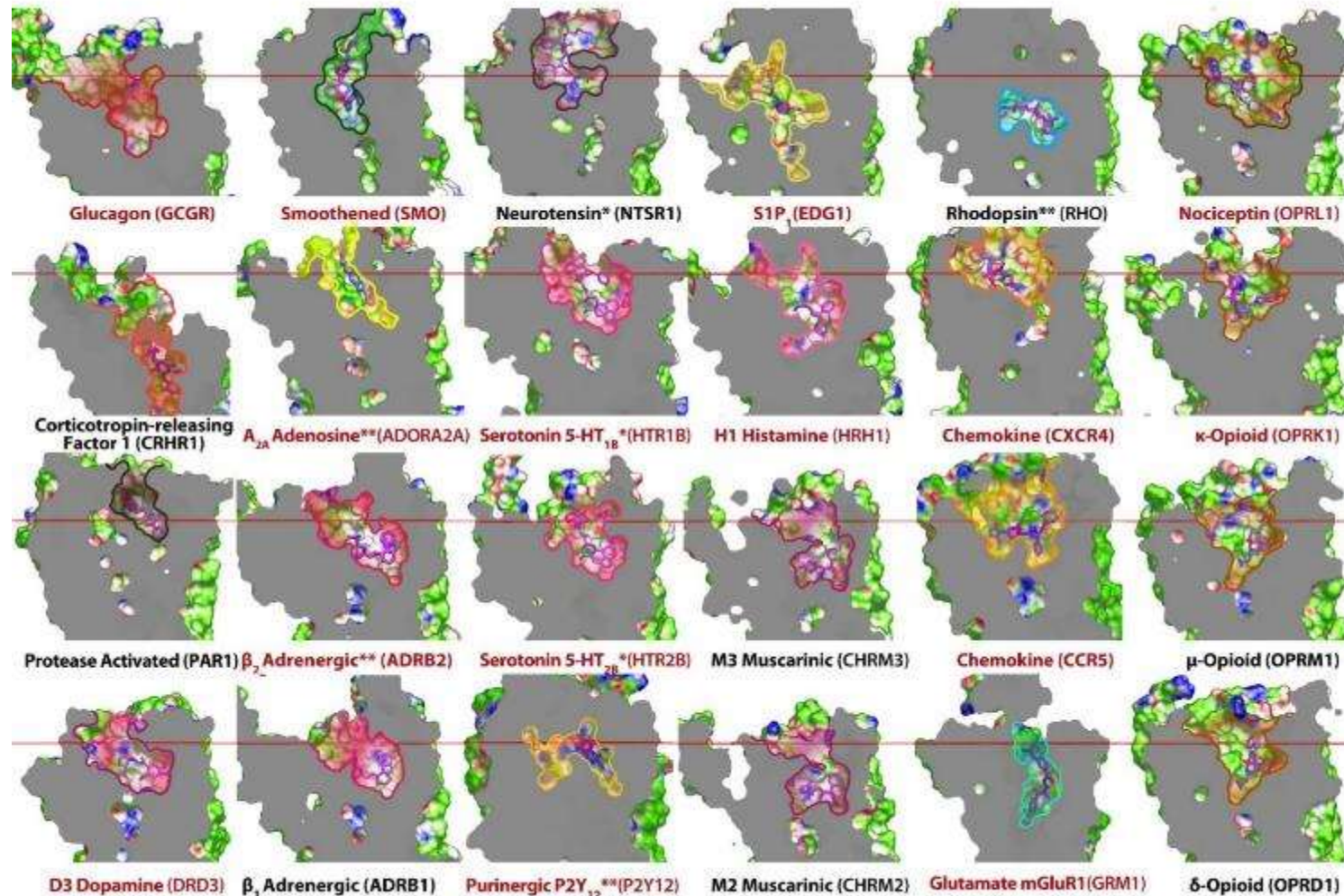
The screenshot shows the Google Cloud Platform interface for the 'My First Project' under the 'Quotas' section. It displays a table of quota types, services, and locations with their respective usage metrics. The 'Used' column is sorted in descending order.

Quota type	Service	Metric	Location	Used
All quotas	All services	All metrics	All locations	
<input type="checkbox"/>	Service		Location	Used ^
<input type="checkbox"/>	Compute Engine API Preemptible CPUs		us-west1	 94,960 / 100,000
<input type="checkbox"/>	Compute Engine API In-use IP addresses		us-west1	 11,872 / 15,000
<input type="checkbox"/>	Compute Engine API Persistent Disk Standard (GB)		us-west1	 120,624 GB / 500,000 GB



# Other Hot GPCR Targets for Ligand Discovery

>50 GPCRs with structures solved and ~10 more coming this year...



\* Solved in agonist-bound partially activated state

\*\* Both agonist-bound and antagonist-bound structures known

# Summary & Outlook : **Structure-based VLS**

- **Structure-based VLS allows effective and fast discovery of novel chemotypes for GPCR targets**
- **High hit rates (20-40%), sub-uM affinities, high Ligand Efficiencies**
- **Subtype- and functional selectivity can be improved via SAR and lead optimization**
- **Targeting new allosteric pockets (e.g. sodium site)**

Ray Stevens & Vadim Cherezov

# Thanks!

GPCR  
Consortium

R33 DA038858; P01 DA035764  
R21 EY027620 (BRAIN Initiative)



## Computational Group



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- ❖ Beili Wu & Quang Zhao (SIMM)
- ❖ Uli Zachariae (U. Dundee)
- ❖ Ken Jacobson (NIH/NIDDK)
- ❖ Patrick Giguere (Ottawa U)
- ❖ Rob Lane (Monash U)
- ❖ Ivy Carroll (RTI)
- ❖ Irene Coin (Leipzig U)
- ❖ Dirk Trauner (NYU)
- ❖ Ruben Abagyan (UCSD/Molsoft)

